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Immune Epitope Database and Analysis Resource Program

2022 Annual IEDB Compendium

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Introduction

The Immune Epitope Database and Analysis Resource (IEDB) is a public repository of immune epitope data and is sponsored by the National Institute for Allergy and Infectious Diseases (NIAID). The IEDB development started in December 2003 and it became available to the public in a beta test phase on 15 February 2006. The IEDB primarily contains data related to antibody and T cell epitopes for humans, non-human primates, rodents, and other animal species. The IEDB also makes available a variety of analytical and epitope prediction tools and resources within its Analysis Resource.

This Compendium of the Immune Epitope Database and Analysis Resource, and consists of three sections. The first section contains a list of the antibody and T cell epitope information in the database as of January 2023. The second section describes the features of the IEDB 4.4 website and the Analysis Resource 2.27. The third section lists the scientific publications in 2021 for which the IEDB played a contributory role.

Since the publication of last year's 2021 Annual Compendium, the quantity of data available in the IEDB continues to increase significantly. As of January 2023, the IEDB contains data for over 2,000,000 epitopes, 75,000 antigens, 6,000,000 assays, and 186,000 receptors from more than 23,000 references. The focus of the IEDB remains on peptidic and non-peptidic epitope data relating to infectious diseases, allergens, autoimmune diseases, and transplant/alloantigens; with exceptions in respect to cancer and HIV.

1 Antibody and T Cell Epitopes

Many new references and species were added to the IEDB in 2022, as demonstrated in Table 1.1. The table lists the number of distinct B cell and T cell epitopes in the database by source species at the end of 2021 and 2022. Of the 4,442 species/strains listed, 196 were added in 2022. It should be noted that the source organism is the species/strain from which the epitopes originate, and may not be from an infecting organism or vaccine target. The curation of MHC binding peptides, cross-reactive epitopes, and autoimmune epitopes has resulted in the appearance of human (*Homo sapiens*) and mouse (*Mus musculus*) epitopes on the list.

In the table below, the leftmost column labeled "New" indicates with an "X" if epitopes for the species/strain was added to the IEDB in 2022. A font color of red is used to highlight the information in the row. The Organism ID matches the NCBI taxonomy ID if it exists. Otherwise, it represents an IEDB-assigned identifier, which are eight-digit numbers starting with "1000". These are easily distinguishable from the NCBI identifiers which are six digits or less. The columns labeled "B 21", "T 21", "B 22", and "T 22" indicate the cumulative number of distinct B cell and T cell epitopes in the database at the end of 2021 and 2022, respectively. The two rightmost columns display the differences in the B and T cell epitope counts from 2021 to 2022. In 2022, the number of B cell epitopes increased by 4,966 from 162,929 to 167,895 and the number of T cell epitopes increased by 319,080 from 1,030,426 to 1,349,506.

Table 1.1 Summary of B and T cell epitopes contained in the IEDB

ID	NEW 2022	ORGANIS M ID	SPECIES STRAIN	B 21	T 21	B 22	T 22	DELT A B	DELTA T
1		2	Bacteria	56		56			
2		106	Runella slithyformis		1		1		
3		108	Spirosoma linguale	1		1			
4		139	Borrelia burgdorferi	93	46	94	46	1	
6		158	Treponema denticola		2		2		
7		160	Treponema pallidum	7	2	7	2		
9		171	Leptospira	2		2			
10		173	Leptospira interrogans	10	4	10	4		
11		195	Campylobacter coli		3		3		
12		197	Campylobacter jejuni	128	30	128	30		
13	X	199	Campylobacter concisus				10		10
16		210	Helicobacter pylori	88	62	88	66		4
17		234	Brucella	1	28	1	28		
18		235	Brucella abortus	7	39	8	39	1	
19		236	Brucella ovis		3		3		
20		263	Francisella tularensis	4	16	4	16		
21		266	Paracoccus denitrificans	3		3			
22		271	Thermus aquaticus	1		1			
23		274	Thermus thermophilus	1	1	1	1		
24		280	Xanthobacter autotrophicus		1		1		
25		286	Pseudomonas	1	1	1	1		
26		287	Pseudomonas aeruginosa	98	259	98	259		
27		292	Burkholderia cepacia	3	2	3	2		
28		294	Pseudomonas fluorescens		17		17		
30		300	Pseudomonas mendocina		2		2		
31		303	Pseudomonas putida	1	11	1	11		
32		312	Pseudomonas sp. RS-16	1		1			
33		316	Stutzerimonas stutzeri		3		3		
34		319	Pseudomonas savastanoi pv. phaseolicola	1		1			
35		346	Xanthomonas citri		1		1		
36		354	Azotobacter vinelandii		2		2		
37		358	Agrobacterium tumefaciens	1	2	1	2		
38		382	Sinorhizobium meliloti		1		1		
39		384	Rhizobium leguminosarum		1		1		
40		446	Legionella pneumophila	1	1	1	1		
41		470	Acinetobacter baumannii	14	10	14	10		
42		471	Acinetobacter calcoaceticus		1		1		
43		480	Moraxella catarrhalis	10		11		1	
44		485	Neisseria gonorrhoeae	77	15	77	15		
45		487	Neisseria meningitidis	102	23	102	23		

46		491	Neisseria meningitidis serogroup B	47	1	47	1		
47	X	502	Kingella denitrificans				1		1
48		511	Alcaligenes faecalis	3	2	3	2		
49		520	Bordetella pertussis	352	218	352	218		
50		521	Bordetella avium		1		1		
51	X	524	Acidiphilium cryptum				1		1
52	X	539	Eikenella corrodens				16		16
53		543	Enterobacteriaceae	4		4			
54		544	Citrobacter	1		1			
55		546	Citrobacter freundii	2		2			
56		548	Klebsiella aerogenes		1		1		
57		550	Enterobacter cloacae		10		10		
58		554	Pectobacterium carotovorum	2		2			
59		556	Dickeya chrysanthemi	1		1			
60		562	Escherichia coli	282	188	286	189	4	1
61		573	Klebsiella pneumoniae	33	12	33	12		
62		582	Morganella morganii	1		1			
63		583	Proteus	2		2			
64		584	Proteus mirabilis	7		7			
66		590	Salmonella	3		3			
67		596	Salmonella enterica subsp. enterica serovar Muenchen	1		1			
68		599	Salmonella sp.	1		1			
69		605	Salmonella enterica subsp. enterica serovar Pullorum		1		1		
70		611	Salmonella enterica subsp. enterica serovar Heidelberg	9		9			
71		615	Serratia marcescens	1	3	1	3		
72		621	Shigella boydii		5		5		
73		622	Shigella dysenteriae	6	2	6	2		
74		623	Shigella flexneri	77	11	77	11		
75		624	Shigella sonnei		1	10	1	10	
76		630	Yersinia enterocolitica	6	29	6	29		
77		632	Yersinia pestis	24	324	24	324		
78		633	Yersinia pseudotuberculosis		2		2		
80		644	Aeromonas hydrophila	9	3	9	3		
81		645	Aeromonas salmonicida	59		59			
82		659	Photobacterium phosphoreum		1		1		
83		666	Vibrio cholerae	40	92	40	92		
84		670	Vibrio parahaemolyticus	1	57	1	57		
85		671	Vibrio proteolyticus	1		1			
86		672	Vibrio vulnificus	2	66	2	66		
87		674	Vibrio mimicus	2		2			
88		686	Vibrio cholerae O1 biovar El Tor	3	1	3	1		
89		703	Plesiomonas shigelloides		2		2		

90		714	Aggregatibacter actinomycetemcomitans	1	4	1	4		
91		715	Actinobacillus pleuropneumoniae		2		2		
92		727	Haemophilus influenzae	36	72	36	72		
93		730	[Haemophilus] ducreyi	3	1	3	1		
94		731	Histophilus somni		1		1		
95		738	Glaesserella parasuis	1		1			
96		747	Pasteurella multocida	1	10	1	10		
97		769	Anaplasma centrale		2		2		
98		770	Anaplasma marginale	25	14	25	14		
99		774	Bartonella bacilliformis	35		35			
100		777	Coxiella burnetii	2	311	8	311	6	
101		779	Ehrlichia ruminantium	3	23	3	23		
102		781	Rickettsia conorii		13		13		
103		782	Rickettsia prowazekii		23		23		
104		783	Rickettsia rickettsii		1		1		
105		784	Orientia tsutsugamushi	87		88		1	
106		785	Rickettsia typhi		3		3		
107		787	Rickettsia australis		1		1		
108		788	Rickettsia canadensis		1		1		
109		813	Chlamydia trachomatis	241	69	242	69	1	
110		817	Bacteroides fragilis	27	4	27	7		3
111	X	818	Bacteroides thetaiotaomicron				1		1
113	X	821	Phocaeicola vulgatus				22		22
114	X	823	Parabacteroides distasonis				1		1
115		837	Porphyromonas gingivalis	111	22	112	22	1	
116	X	851	Fusobacterium nucleatum				140		140
117	X	853	Faecalibacterium prausnitzii				3		3
119		920	Acidithiobacillus ferrooxidans		1		1		
120		944	Ehrlichia canis	8	1	8	1		
121		945	Ehrlichia chaffeensis	14	1	14	1		
123		948	Anaplasma phagocytophilum	45		47		2	
124		955	Wolbachia pipientis	1		1			
125		1006	Marivirga tractuosa	1	1	1	1		
126		1018	Capnocytophaga ochracea		1		1		
127		1019	Capnocytophaga sputigena		1		1		
128		1022	Beggiatoa alba	1		1			
129		1063	Cereibacter sphaeroides		1		1		
130		1076	Rhodopseudomonas palustris		2		2		
131		1126	Microcystis aeruginosa	1		1			
132		1224	Pseudomonadota	3		3			
133		1245	Leuconostoc mesenteroides	1		1			
134		1254	Pediococcus acidilactici	1		1			
135		1260	Finegoldia magna		1		1		

136		1265	Ruminococcus flavefaciens		1		1		
137		1270	Micrococcus luteus	1		1			
138		1280	Staphylococcus aureus	194	52	219	74	25	22
140		1292	Staphylococcus warneri	1		1			
141		1299	Deinococcus radiodurans	3		3			
142		1301	Streptococcus	1		1			
143	X	1302	Streptococcus gordonii				5		5
144		1303	Streptococcus oralis	12	1	12	1		
146		1305	Streptococcus sanguinis		1		1		
147		1307	Streptococcus suis	4		4			
148		1309	Streptococcus mutans	146	89	146	89		
149		1310	Streptococcus sobrinus	1		1			
150		1311	Streptococcus agalactiae	19		19			
151		1313	Streptococcus pneumoniae	96	31	97	31	1	
152		1314	Streptococcus pyogenes	420	125	420	125		
153		1317	Streptococcus downei	2	1	2	1		
154		1319	Streptococcus sp. 'group B'	4		4			
155		1320	Streptococcus sp. 'group G'	14	5	14	5		
156		1334	Streptococcus dysgalactiae	3	7	3	7		
157		1336	Streptococcus equi	22	14	22	14		
159		1349	Streptococcus uberis	8		8			
160		1351	Enterococcus faecalis	2		2			
161		1352	Enterococcus faecium	6		6			
162		1358	Lactococcus lactis		2		2		
163		1386	Bacillus		1		1		
164		1390	Bacillus amyloliquefaciens	18	3	18	3		
165		1392	Bacillus anthracis	403	209	403	209		
166		1396	Bacillus cereus	1	15	1	15		
167		1402	Bacillus licheniformis		10		10		
168		1404	Priestia megaterium		1		7		6
169		1405	Bacillus mycoides		4		4		
170		1406	Paenibacillus polymyxa	1		1			
171		1408	Bacillus pumilus		1		1		
172		1422	Geobacillus stearothermophilus	2	1	2	1		
173		1423	Bacillus subtilis	2	4	2	4		
174		1428	Bacillus thuringiensis	7	10	9	10	2	
175		1435	Bacillus thuringiensis serovar san diego		2		2		
176		1454	Schinkia azotoformans		1		1		
177		1467	Lederbergia lenta		1		1		
178		1491	Clostridium botulinum	207	111	207	111		
179		1492	Clostridium butyricum		1		1		
180		1496	Clostridioides difficile	98	2	100	2	2	
181		1502	Clostridium perfringens	41	2	43	2	2	

182		1513	Clostridium tetani	82	282	82	282		
183		1520	Clostridium beijerinckii	1		1			
184	X	1531	Enterocloster clostridioformis				2		2
185	X	1582	Lactacaseibacillus casei			1		1	
186		1585	Lactobacillus delbrueckii subsp. bulgaricus	1		1			
187	X	1590	Lactiplantibacillus plantarum				8		8
188		1599	Latilactobacillus sakei		1		1		
189		1604	Lactobacillus amylovorus		1		1		
190	X	1605	Ligilactobacillus animalis				9		9
191		1613	Limosilactobacillus fermentum	1		1			
192		1638	Listeria ivanovii	1	2	1	2		
193		1639	Listeria monocytogenes	25	104	25	104		
195		1641	Listeria grayi		1		1		
196		1642	Listeria innocua	2	2	2	2		
197		1646	Renibacterium salmoninarum	12		12			
198	X	1660	Schaalia odontolytica				8		8
199		1661	Trueperella pyogenes	7		7			
200		1664	Arthrobacter glacialis		1		1		
201		1681	Bifidobacterium bifidum		1		1		
202		1685	Bifidobacterium breve		1		1		
203		1716	Corynebacterium		1		2		1
204		1717	Corynebacterium diphtheriae	15	55	15	55		
205		1718	Corynebacterium glutamicum		26		26		
206	X	1719	Corynebacterium pseudotuberculosis			6	5	6	5
207		1747	Cutibacterium acnes		2		2		
208		1752	Propionibacterium freudenreichii subsp. shermanii		1		1		
209		1763	Mycobacterium	29	14	29	14		
210		1764	Mycobacterium avium	6	18	6	18		
211		1765	Mycobacterium tuberculosis variant bovis	50	330	50	330		
212		1766	Mycolicibacterium fortuitum		1		1		
213		1767	Mycobacterium intracellulare		1		1		
214		1768	Mycobacterium kansasii	9	12	9	12		
215		1769	Mycobacterium leprae	153	409	153	409		
216		1770	Mycobacterium avium subsp. paratuberculosis	106	15	106	15		
217		1771	Mycolicibacterium phlei		1		1		
218		1772	Mycolicibacterium smegmatis		29		29		
219		1773	Mycobacterium tuberculosis	677	2649	681	2656	4	7
220		1774	Mycobacteroides chelonae	1		1			
221		1778	Mycobacterium gordonae	1		1			
222		1781	Mycobacterium marinum		2		2		
223		1783	Mycobacterium scrofulaceum	4		4			

224		1795	Mycolicibacterium neoaurum		1		1		
225		1804	Mycolicibacterium gilvum		1		1		
226		1809	Mycobacterium ulcerans		11		11		
228		1828	Rhodococcus fascians		2		2		
229		1830	Rhodococcus ruber		1		1		
230		1833	Rhodococcus erythropolis		3		3		
231		1836	Saccharopolyspora erythraea	4		4			
232		1895	Streptomyces avidinii	1		1			
233		1902	Streptomyces coelicolor		2		2		
234		1916	Streptomyces lividans	11	1	11	1		
235		1943	Streptomyces griseoruber		1		1		
236		1946	Streptomyces kasugaensis		1		1		
237		2039	Tropheryma whipplei	1		1			
238		2097	Mycoplasma genitalium		1		1		
239		2099	Mesomycoplasma hyopneumoniae	3		3			
240		2100	Mesomycoplasma hyorhinis	1		1			
241		2104	Mycoplasma pneumoniae	20	6	20	6		
243		2133	Spiroplasma citri		1		1		
244		2147	Acholeplasma	1		1			
245		2148	Acholeplasma laidlawii		2		2		
246		2173	Methanobrevibacter smithii		1		1		
247		2190	Methanocaldococcus jannaschii		1		1		
248		2234	Archaeoglobus fulgidus	1	1	1	1		
249		2242	Halobacterium salinarum	1		1			
250		2261	Pyrococcus furiosus		1		1		
252		2287	Saccharolobus solfataricus	1		1			
253		2336	Thermotoga maritima	2		2			
254		2439	Plasmid ColB2	1		1			
255		2465	Plasmid F	1		1			
256	X	2702	Gardnerella vaginalis				9		9
258		2736	Verrucomicrobium spinosum	1		1			
259		2759	Eukaryota	1		1			
260		2864	Dinophyceae	3		3			
261		2903	Emiliana huxleyi		1		1		
262		3039	Euglena gracilis	1		1			
263	X	3055	Chlamydomonas reinhardtii			1			1
264		3068	Volvox carteri f. nagariensis		1		1		
265		3318	Pinaceae	2		2			
266		3346	Pinus pinea	37		37			
267		3352	Pinus taeda	4		4			
268		3369	Cryptomeria japonica	45	300	45	300		
269		3469	Papaver somniferum	1		1			
270		3505	Betula pendula	91	337	91	337		

271		3513	Quercus alba		5		5		
272		3517	Alnus glutinosa		17		17		
273		3559	Chenopodium album	3	2	3	2		
274		3562	Spinacia oleracea	4		4			
276		3617	Fagopyrum esculentum	40		40			
277	X	3625	Actinidia chinensis			1		1	
278		3645	Bertholletia excelsa	7	24	7	24		
279		3656	Cucumis melo	13		13			
280		3677	Trichosanthes kirilowii	4		4			
281		3702	Arabidopsis thaliana	6	10	20	10	14	
282		3704	Armoracia rusticana	5		6		1	
283		3707	Brassica juncea	9		9			
285		3711	Brassica rapa	1		1			
286		3726	Raphanus sativus	1		1			
287		3728	Sinapis alba	2		2			
288		3750	Malus domestica	27	48	28	48	1	
289		3755	Prunus dulcis	19		19			
290		3758	Prunus domestica	4		4			
291		3760	Prunus persica	19	51	50	51	31	
292		3816	Abrus precatorius	99		99			
293		3818	Arachis hypogaea	497	263	711	263	214	
294		3847	Glycine max	649	8	651	8	2	
295		3864	Lens culinaris	19		24		5	
296		3885	Phaseolus vulgaris	10	3	10	3		
298		3972	Viscum album	17		17			
299		3981	Hevea brasiliensis	212	43	213	43	1	
300		3988	Ricinus communis	90	6	101	6	11	
301		4012	Rhus		1		1		
302		4026	Acer pseudoplatanus	2		2			
303		4039	Daucus carota		46	1	46	1	
304		4045	Apium graveolens		14	2	14	2	
305		4081	Solanum lycopersicum	3		3			
306		4097	Nicotiana tabacum	5		5			
307		4100	Nicotiana benthamiana	5		5			
309	X	4120	Ipomoea batatas			1		1	
310		4146	Olea europaea	76	19	76	19		
311		4163	Digitalis	1		1			
312		4182	Sesamum indicum	11		12		1	
313		4212	Ambrosia artemisiifolia	5	215	6	216	1	1
314		4214	Ambrosia trifida		10		10		
315		4215	Ambrosia artemisiifolia var. elatior	9	12	9	12		
316		4220	Artemisia vulgaris	4	70	4	70		
317		4232	Helianthus annuus	18		18			

318		4479	Poaceae	1		1			
319		4498	Avena sativa		16		16		
320		4509	Dactylis glomerata		12		12		
321		4513	Hordeum vulgare		61		61		
322		4522	Lolium perenne	44	123	44	123		
323		4530	Oryza sativa	2		6		4	
324		4545	Poa pratensis	34	35	34	35		
325		4550	Secale cereale	51	82	51	82		
326		4558	Sorghum bicolor		1		1		
327		4565	Triticum aestivum	655	623	655	624		1
328		4567	Triticum turgidum subsp. durum		1		1		
329		4568	Triticum monococcum		1		1		
331		4577	Zea mays	3	1	5	1	2	
332		4606	Lolium arundinaceum	1		1			
333		4615	Ananas comosus	3		3			
334		4641	Musa acuminata	50	1	50	1		
335		4751	Fungi	6		6			
336		4754	Pneumocystis carinii	2	1	2	1		
337	X	4784	Phytophthora capsici			1		1	
338	X	4792	Phytophthora parasitica			1		1	
339		4903	Cyberlindnera jadinii	12		12			
340		4920	Millerozyma farinosa		1		1		
341		4932	Saccharomyces cerevisiae	37	17	37	17		
342		4954	Zygosaccharomyces bailii		1		1		
343		5037	Histoplasma capsulatum		1		81		80
344		5039	Blastomyces dermatitidis		1		1		
345		5052	Aspergillus	7		8		1	
346		5057	Aspergillus clavatus		1		1		
347		5059	Aspergillus flavus	6	1	6	1		
348		5060	Aspergillus giganteus		4		4		
349		5061	Aspergillus niger		3		3		
350		5064	Aspergillus restrictus	1		1			
351		5067	Aspergillus parasiticus		1		1		
352		5076	Penicillium chrysogenum	45	19	45	19		
353		5077	Penicillium citrinum	13		13			
354		5141	Neurospora crassa	6	3	6	3		
355		5207	Cryptococcus neoformans	4	3	4	3		
356		5325	Trametes versicolor		1		1		
357		5334	Schizophyllum commune	1	1	1	1		
358		5346	Coprinopsis cinerea		1		1		
359		5476	Candida albicans	114	82	114	82		
360		5480	Candida parapsilosis	1		1			
361		5482	Candida tropicalis		2		2		

362		5501	Coccidioides immitis		1		1		
363		5503	Curvularia lunata	20	10	20	10		
364		5507	Fusarium oxysporum		4		4		
365		5518	Fusarium graminearum	9		9			
366		5551	Trichophyton rubrum	2	27	2	27		
367		5554	Cutaneotrichosporon cutaneum	1		1			
368		5580	Aureobasidium pullulans		1		1		
369		5599	Alternaria alternata	8	38	8	38		
370		5658	Leishmania	4		4			
371		5659	Leishmania amazonensis	5	15	5	15		
372		5660	Leishmania braziliensis	12		20		8	
373		5661	Leishmania donovani	30	108	30	108		
374		5664	Leishmania major	32	186	32	186		
375		5665	Leishmania mexicana	3	3	3	3		
376		5667	Leishmania aethiopica	16		16			
377		5671	Leishmania infantum	108	9	108	9		
378		5679	Leishmania panamensis	3	18	3	18		
379		5682	Leishmania pifanoi		20		20		
380		5691	Trypanosoma brucei	13	6	13	6		
381		5692	Trypanosoma congolense	1		1			
382		5693	Trypanosoma cruzi	164	460	164	460		
383		5702	Trypanosoma brucei brucei	5		9		4	
384		5722	Trichomonas vaginalis	80	1	80	1		
385		5741	Giardia intestinalis	1	13	1	13		
386		5755	Acanthamoeba castellanii	14	3	14	3		
387		5759	Entamoeba histolytica	30	14	30	14		
388		5762	Naegleria gruberi	1		1			
389		5801	Eimeria acervulina	1		1			
390		5802	Eimeria tenella	8		8			
391		5807	Cryptosporidium parvum	4	31	4	31		
392		5811	Toxoplasma gondii	99	103	99	103		
393		5821	Plasmodium berghei	31	57	31	57		
394		5823	Plasmodium berghei ANKA	7	65	9	66	2	1
395		5824	Plasmodium brasilianum	2		2			
396		5825	Plasmodium chabaudi	4	4	4	4		
397		5827	Plasmodium cynomolgi	2		2			
399		5833	Plasmodium falciparum	3392	1061	3411	1061	19	
400		5834	Plasmodium falciparum RO-33	8	1	8	1		
401		5835	Plasmodium falciparum CAMP/Malaysia	36	3	36	3		
402		5836	Plasmodium falciparum CDC/Honduras	18		18			
403		5837	Plasmodium falciparum FC27/Papua New Guinea	47	60	47	60		
404		5838	Plasmodium falciparum FCR-3/Gambia	94		94			

405	5839	Plasmodium falciparum K1	18	19	18	19		
406	5840	Plasmodium falciparum LE5		2		2		
407	5841	Plasmodium falciparum Mad20/Papua New Guinea	8	47	8	47		
408	5842	Plasmodium falciparum NF7/Ghana	4		4			
409	5843	Plasmodium falciparum NF54	21	31	33	37	12	6
410	5846	Plasmodium falciparum T4/Thailand	25	1	25	1		
411	5848	Plasmodium falciparum isolate WELLCOME	25	51	25	51		
412	5850	Plasmodium knowlesi	2	17	2	17		
413	5851	Plasmodium knowlesi strain H	41	10	41	10		
414	5852	Plasmodium knowlesi strain Nuri		10		10		
415	5855	Plasmodium vivax	387	176	388	176	1	
416	5857	Plasmodium fragile		5		5		
417	5858	Plasmodium malariae	1		1			
418	5861	Plasmodium yoelii	19	39	19	39		
419	5865	Babesia bovis	5	8	14	14	9	6
420	5866	Babesia bigemina		3	4	3	4	
421	5868	Babesia microti	5		7		2	
422	5872	Theileria equi	8		8			
423	5874	Theileria annulata	1		1			
424	5875	Theileria parva	28	20	28	27		7
426	5970	Exophiala dermatitidis		1		1		
427	6035	Encephalitozoon cuniculi		5		5		
428	6100	Aequorea victoria	5	5	5	5		
429	6181	Schistosoma	3		3			
430	6182	Schistosoma japonicum	1207	53	1215	54	8	1
431	6183	Schistosoma mansoni	3162	52	3162	52		
432	6184	Schistosoma bovis	2		2			
434	6192	Fasciola hepatica	300	28	300	28		
435	6198	Opisthorchis viverrini	4		4			
436	6203	Taenia ovis	8		8			
437	6204	Taenia solium	29	4	29	4		
438	6206	Taenia saginata	13		13			
439	6207	Taenia crassiceps	11	1	11	1		
440	6210	Echinococcus granulosus	41	2	47	6	6	4
441	6211	Echinococcus multilocularis	22	14	29	22	7	8
442	6238	Caenorhabditis briggsae	1		2		1	
443	6239	Caenorhabditis elegans	2	10	3	10	1	
445	6252	Ascaris lumbricoides	11		11			
446	6253	Ascaris suum	1	290	1	290		
447	6265	Toxocara canis	3		3			
448	6269	Anisakis simplex	40	28	40	28		
449	6279	Brugia malayi	10	5	10	5		

450		6280	Brugia pahangi	1		1			
451		6282	Onchocerca volvulus	4394	39	4418	39	24	
452		6289	Haemonchus contortus	3		3			
453		6293	Wuchereria bancrofti	13	8	13	8		
454		6299	Litomosoides carinii	3		3			
455	X	6313	Angiostrongylus cantonensis			32		32	
456		6334	Trichinella spiralis	18	7	18	7		
457		6339	Heligmosomoides polygyrus	3		3			
458		6421	Hirudo medicinalis		3		3		
459		6454	Haliotis rufescens	1		1			
460		6491	Conus geographus	30		30			
461		6492	Conus magus	2		2			
462		6493	Conus striatus	11		11			
464		6536	Helix pomatia	1		1			
465		6594	Macrocallista nimbosa	1		1			
466		6661	Artemia franciscana	2		2			
467		6687	Penaeus monodon	10	23	10	23		
468		6689	Penaeus vannamei	158	2	158	2		
469		6690	Penaeus aztecus	51	31	51	31		
470		6728	Procambarus clarkii	3		3			
471		6761	Scylla serrata	13		13			
473		6850	Limulus polyphemus		1		1		
474		6853	Tachypleus tridentatus	1		1			
475		6858	Androctonus australis	9		9			
476		6860	Androctonus mauritanicus mauritanicus	1		1			
477		6878	Centruroides noxius	10		10			
478		6887	Tityus serrulatus	92		92			
479		6925	Latrodectus tredecimguttatus		1		1		
480		6938	Ornithodoros moubata	20		20			
481		6941	Rhipicephalus microplus	2		2			
482		6945	Ixodes scapularis	11		11			
483		6954	Dermatophagoides farinae	58	315	58	315		
484		6956	Dermatophagoides pteronyssinus	155	700	157	700	2	
485		6958	Euroglyphus maynei		10		10		
486		6973	Blattella germanica	23	501	23	501		
487		6978	Periplaneta americana	30	6	33	9	3	3
488	X	7004	Locusta migratoria			1		1	
489		7088	Lepidoptera		1		1		
490		7108	Spodoptera frugiperda		4		4		
491		7130	Manduca sexta		3		3		
492		7137	Galleria mellonella	2		2			
493		7154	Chironomus thummi	2	3	2	3		
494		7155	Chironomus thummi thummi	58	27	58	27		

495		7160	Aedes albopictus	7		7			
496		7165	Anopheles gambiae	8	2	8	2		
497		7227	Drosophila melanogaster	11	17	15	17	4	
498		7234	Drosophila persimilis	1		1			
499		7386	Sarcophaga peregrina		1		1		
500		7441	Dolichovespula maculata	11	20	11	20		
501		7444	Vespa basalis	1		1			
502		7453	Vespula maculifrons	1		1			
503		7454	Vespula vulgaris	2	95	2	95		
504		7460	Apis mellifera	15	108	15	108		
505		7469	Apis mellifera ligustica		1		1		
506		7515	Ctenocephalides felis		2		2		
507		7719	Ciona intestinalis	2		3		1	
508		7726	Styela plicata	6		6			
509	X	7741	Branchiostoma belcheri			1		1	
510		7742	Vertebrata	4		4			
511		7777	Chondrichthyes	1		1			
512		7787	Tetronarce californica	151	75	151	75		
513		7788	Torpedo marmorata	3	3	3	3		
514		7955	Danio rerio	2	3	6	3	4	
515		7957	Carassius auratus	3	1	4	1	1	
516	X	7959	Ctenopharyngodon idella			1		1	
517		7962	Cyprinus carpio	6	1	6	1		
518		8005	Electrophorus electricus	4		4			
519		8018	Oncorhynchus keta		6		6		
520		8022	Oncorhynchus mykiss	1		1			
521		8030	Salmo salar	35		35			
522		8049	Gadus morhua	44		44			
523		8053	Gadus morhua callarias	10		10			
524		8090	Oryzias latipes	2	1	2	1		
525		8255	Paralichthys olivaceus	1		1			
526		8342	Anura	1		1			
527		8355	Xenopus laevis	4	5	4	5		
528		8364	Xenopus tropicalis	2	2	2	2		
529		8554	Heloderma suspectum		1		1		
530		8613	Bungarus fasciatus	1		1			
531		8616	Bungarus multicinctus	17	3	17	3		
532		8620	Dendroaspis polylepis polylepis	22		22			
533		8654	Naja nigricollis	3	2	3	2		
535		8656	Naja atra	14		26		12	
536		8657	Naja oxiana	4		4			
537		8658	Naja pallida	1	1	1	1		
538		8663	Notechis scutatus	2		2			

539	8667	Oxyuranus scutellatus scutellatus	4		4			
540	8671	Pseudechis porphyriacus	3		3			
541	8704	Vipera ammodytes	1		1			
542	8722	Bothrops asper	28		28			
543	8724	Bothrops jararaca	5		5			
544	8725	Bothrops atrox	13		13			
545	8726	Bothrops jararacussu	13		13			
546	8732	Crotalus durissus terrificus	31		31			
547	8753	Lachesis muta muta	45		45			
548	8801	Struthio camelus	2		2			
549	8839	Anas platyrhynchos	2		2			
550	8845	Anser cygnoides	2		2			
551	8855	Cairina moschata	2	1	2	1		
552	8932	Columba livia		10		10		
553	8996	Numida meleagris	1		1			
554	9014	Colinus virginianus	1	3	1	3		
555	9031	Gallus gallus	406	2194	610	2194	204	
556	9057	Phasianus colchicus colchicus	1		1			
558	9103	Meleagris gallopavo	9		9			
559	9258	Ornithorhynchus anatinus		1		1		
561	9322	Macropus sp.	1		1			
562	9337	Trichosurus vulpecula	45		45			
563	9397	Chiroptera	1		1			
565	9447	Lemur catta	1		1			
566	9479	Platyrrhini	1		1			
567	9483	Callithrix jacchus	1	4	1	4		
568	9488	Saguinus mystax	1		1			
569	9491	Saguinus imperator	1		1			
570	9502	Alouatta caraya	1		1			
571	9505	Aotus trivirgatus	1		1			
572	9509	Ateles geoffroyi		1		1		
573	9510	Ateles paniscus	1		1			
574	9511	Ateles sp.	1		1			
575	9515	Sapajus apella	1		1			
576	9519	Lagothrix lagotricha	1		1			
577	9521	Saimiri sciureus	1		1			
578	9534	Chlorocebus aethiops		5		5		
579	9538	Erythrocebus patas		1		1		
580	9541	Macaca fascicularis	6	4	7	4	1	
581	9544	Macaca mulatta	3	5	3	5		
583	9548	Macaca radiata	14	1	14	1		
584	9555	Papio anubis	1		1			
585	9556	Papio cynocephalus	1		1			

586		9580	Hylobates lar		1		1		
587		9593	Gorilla gorilla		2		2		
588		9595	Gorilla gorilla gorilla		6		6		
589		9597	Pan paniscus		1		1		
590		9598	Pan troglodytes	7	25	7	25		
591		9600	Pongo pygmaeus		2		2		
592		9601	Pongo abelii		2		2		
593		9606	Homo sapiens	34420	797430	34977	1058562	557	261132
594		9615	Canis lupus familiaris	24	1924	24	1924		
595		9627	Vulpes vulpes	1		1			
596		9666	Mustela lutreola	1		1			
597		9685	Felis catus	42	117	42	117		
598		9721	Cetacea	1		1			
599		9755	Physeter catodon	2	34	2	34		
600		9796	Equus caballus	56	57	56	57		
601		9798	Equus przewalskii		1		1		
602		9823	Sus scrofa	126	55	127	55	1	
603		9825	Sus scrofa domesticus		5		5		
604		9844	Lama glama	1		1			
605		9860	Cervus elaphus	6		6			
606		9864	Cervus canadensis nelsoni	5		5			
607		9874	Odocoileus virginianus	2		2			
608		9913	Bos taurus	1345	855	1360	1515	15	660
609		9925	Capra hircus	11		11			
610		9940	Ovis aries	96	17	96	17		
611		9978	Ochotona princeps		1		1		
612		9986	Oryctolagus cuniculus	66	10	66	10		
613		10029	Cricetulus griseus	9		10		1	
614		10036	Mesocricetus auratus	58		58			
615		10042	Peromyscus maniculatus		1		1		
616		10090	Mus musculus	761	43643	809	81302	48	37659
617		10092	Mus musculus domesticus		12		12		
618		10114	Rattus	10		10			
619		10116	Rattus norvegicus	296	33149	304	33154	8	5
620		10141	Cavia porcellus	37	86	37	86		
621	X	10224	Saccoglossus kowalevskii			1		1	
622		10243	Cowpox virus		26		26		
623		10244	Monkeypox virus	5	2	5	2		
624		10245	Vaccinia virus	11	725	11	725		
625		10247	Vaccinia virus WR 65-16		4		4		
626		10248	Vaccinia virus LC16M8		5		5		
627		10249	Vaccinia virus Copenhagen	31	851	31	856		5
628		10251	Vaccinia virus IHD-J	1		1			

629		10253	Vaccinia virus Tian Tan		21		21		
630		10254	Vaccinia virus WR	31	6823	31	6823		
631		10255	Variola virus		250	1	250	1	
632		10258	Orf virus	1		1			
633		10261	Fowlpox virus		7		7		
634		10273	Myxoma virus		4		4		
635		10276	Swinepox virus		2		2		
636		10298	Human alphaherpesvirus 1	183	585	183	586		1
637		10299	Human alphaherpesvirus 1 strain 17	44	64	44	64		
638		10301	Human alphaherpesvirus 1 strain Angelotti		4		4		
639		10303	Human alphaherpesvirus 1 strain HFEM	2		2			
640		10304	Human alphaherpesvirus 1 strain F	17	5	17	5		
641		10306	Human alphaherpesvirus 1 strain KOS	17	4	19	4	2	
642		10308	Human alphaherpesvirus 1 strain Patton	3		3			
643		10309	Human alphaherpesvirus 1 strain SC16	5	2	5	2		
644		10310	Human alphaherpesvirus 2	295	230	295	234		4
645		10312	Human herpesvirus 2 strain 186	1	1	1	13		12
646		10313	Human herpesvirus 2 strain 333	2	5	2	5		
647		10315	Human herpesvirus 2 strain HG52		32		32		
648		10317	Cercopithecine alphaherpesvirus 2		1		1		
649		10320	Bovine alphaherpesvirus 1	7	40	7	40		
650		10323	Bovine herpesvirus type 1.1 (strain Cooper)	9	17	9	17		
651		10324	Bovine herpesvirus type 1.1 (strain P8-2)	1		1			
652		10325	Macacine alphaherpesvirus 1	366		366			
653		10326	Equid alphaherpesvirus 1	7	218	7	218		
654		10331	Equid alphaherpesvirus 4	6		6			
655		10335	Human alphaherpesvirus 3	10	131	10	131		
656		10338	Human herpesvirus 3 strain Dumas		29		29		
657		10345	Suid alphaherpesvirus 1	14	3	14	3		
658		10349	Suid herpesvirus 1 (strain NIA-3)	4		4			
659		10359	Human betaherpesvirus 5	126	1321	131	1327	5	6
660		10360	Human herpesvirus 5 strain AD169	236	535	238	540	2	5
661		10363	Human herpesvirus 5 strain Towne	96	26	101	26	5	
662		10366	Murid betaherpesvirus 1	2	64	2	64		
663		10367	Murine cytomegalovirus (strain Smith)		32		32		
665		10369	Human herpesvirus 6 (strain GS)	2		2			
666		10370	Human herpesvirus 6 (strain Uganda-1102)	2	3	2	3		
667		10372	Human betaherpesvirus 7	2	4	2	4		
668		10373	Macaca mulatta cytomegalovirus		76		76		
669		10376	Human gammaherpesvirus 4	2423	708	2443	711	20	3

670		10377	Human herpesvirus 4 strain B95-8	187	389	187	401		12
671		10378	Human herpesvirus 4 strain RAJI	1		1			
672		10381	Saimiriine gammaherpesvirus 2		3		3		
673		10390	Gallid alphaherpesvirus 2		1		1		
674		10407	Hepatitis B virus	388	768	401	878	13	110
675		10408	Hepatitis B virus subtype adw2	82	61	82	61		
676		10409	Hepatitis B virus adr4		9		9		
677		10410	Hepatitis B virus adw/991	2	2	2	2		
678		10411	Hepatitis B virus alpha1		19		19		
679		10412	Hepatitis B virus adw/Indonesia/PIDW420		3		3		
680		10414	Hepatitis B virus LSH/chimpanzee		2		2		
681		10415	Hepatitis B virus adw/Okinawa/PODW282		5		5		
682		10418	Hepatitis B virus subtype ayw	57	185	57	185		
683		10419	Hepatitis B virus subtype adyw	4	45	4	45		
684		10430	Woodchuck hepatitis virus 1	3		3			
685		10433	Woodchuck hepatitis virus 8		34		34		
686		10479	Thermoproteus tenax virus 1	1		1			
687		10497	African swine fever virus	9	31	23	34	14	3
688		10498	African swine fever virus BA71V	58	106	58	106		
689		10514	Canine adenovirus 2	2		2			
690		10515	Human adenovirus 2	28	62	28	62		
691		10519	Human adenovirus 7	4		4			
692		10521	Human adenovirus 14	4		4			
693		10524	Human adenovirus 41	1		1			
694		10529	Human adenovirus 31		7		7		
695		10530	Murine adenovirus 1	1	2	1	2		
696		10533	Human adenovirus 1		1	1	1	1	
697		10541	Human adenovirus 11		12		12		
698		10562	Bos taurus papillomavirus 4	10	3	10	3		
699		10566	Human papillomavirus	1	1	1	1		
700		10573	human papillomavirus 13	1	1	1	1		
702		10580	human papillomavirus 11	9	43	9	43		
703		10583	Human papillomavirus type 1a	8	15	8	15		
704		10585	human papillomavirus 31	13	7	13	7		
705		10586	human papillomavirus 33	6	9	6	9		
706		10587	human papillomavirus 35	1	1	1	1		
707		10588	human papillomavirus 39	1	1	1	1		
708		10589	Human papillomavirus type 41		1		1		
709		10592	human papillomavirus 44		1		1		
710		10593	human papillomavirus 45	3	9	3	9		
711		10595	human papillomavirus 51	1	1	1	1		
712		10596	human papillomavirus 56	1	5	1	5		

713	10598	human papillomavirus 58	49	34	52	34	3	
714	10600	Human papillomavirus type 6b	11	29	11	29		
715	10611	human papillomavirus 30	1		1			
716	10614	Human papillomavirus 3		3		3		
717	10615	human papillomavirus 40	1	1	1	1		
718	10617	Human papillomavirus 4		4		4		
719	10618	human papillomavirus 52	1	36	1	36		
720	10620	Human papillomavirus type 7	1	3	1	3		
721	10623	Kappapapillomavirus 2	5	7	5	7		
722	10631	BK virus strain AS		1		1		
723	10632	JC polyomavirus	9	96	9	96		
724	10636	Murine polyomavirus strain A2	1	3	1	3		
725	10638	Kilham polyomavirus		1		1		
726	10665	Tequatrovirus T4	10	20	10	20		
727	10678	Punavirus P1		6		6		
728	10703	Corynephage beta	2		2			
729	10710	Lambdavirus lambda		6		6		
730	10754	Lederbergvirus P22	6		6			
731	10760	Escherichia phage T7	1		1			
732	10783	Aleutian mink disease parvovirus (STRAIN G)	3		3			
733	10786	Feline panleukopenia virus		4		4		
734	10788	Canine parvovirus	24	21	24	21		
735	10790	Canine parvovirus strain CPV-D CORNELL 320	1		1			
736	10791	Canine parvovirus strain N	47		47			
737	10793	Mink enteritis virus strain Abashiri	1		1			
738	10796	Porcine parvovirus	41		44		3	
739	10798	Human parvovirus B19	94	44	94	44		
740	10804	adeno-associated virus 2	42	153	42	153		
741	10847	Sinheimvirus phiX174	1		1			
742	10863	Enterobacteria phage f1	1	1	1	1		
743	10864	Enterobacteria phage fd	7	1	7	1		
744	10879	Cystovirus phi6	7		7			
745	10884	Mammalian orthoreovirus 1 Lang	3		3			
746	10886	Mammalian orthoreovirus 3 Dearing	1		1			
747	10891	Reovirus sp.		1		1		
748	10900	Bluetongue virus (serotype 10 / American isolate)	3		3			
749	10903	Bluetongue virus 17	4		4			
750	10904	Bluetongue virus (serotype 1 / isolate Australia)	7		7			
751	10906	Bluetongue virus 10	3		3			
752	10915	Porcine rotavirus serotype 5/strain OSU	1		1			
753	10917	Porcine rotavirus strain Gottfried	2		2			

754	10919	Porcine rotavirus strain YM	2	1	2	1		
755	10923	Simian rotavirus A/SA11	8	5	8	5		
756	10927	Bovine rotavirus	9	1	9	1		
757	10933	Bovine rotavirus strain RF	12	7	12	7		
758	10934	Bovine rotavirus strain UK/G6		1		1		
759	10941	Human rotavirus A	14	84	14	84		
760	10950	Human rotavirus (SEROTYPE 2 / STRAIN DS1)	1		1			
761	10952	Human rotavirus strain KU	9		9			
762	10957	Human rotavirus strain P	1	4	1	4		
763	10958	Human rotavirus strain RRV	1		1			
764	10960	Human rotavirus G4 strain St. Thomas 3	6		6			
765	10962	Human rotavirus strain WA	10	7	10	7		
766	10995	Infectious bursal disease virus	12		12			
767	10996	Infectious bursal disease virus 52/70		10		10		
768	10997	Infectious bursal disease virus 002-73/AUS	1		1			
769	11002	Infectious pancreatic necrosis virus	1		1			
770	11021	Eastern equine encephalitis virus	48	2	48	2		
771	11022	Eastern equine encephalitis virus (STRAIN VA33[TEN BROECK])		1		1		
772	11029	Ross River virus	1	1	1	1		
773	11032	Ross river virus (STRAIN T48)	11		11			
774	11033	Semliki Forest virus	36	5	36	5		
775	11034	Sindbis virus	26		26			
776	11036	Venezuelan equine encephalitis virus	2	6	2	6		
777	11037	Venezuelan equine encephalitis virus (strain TC-83)	25		25			
778	11038	Venezuelan equine encephalitis virus (strain Trinidad donkey)	15		15			
779	11039	Western equine encephalitis virus		1		1		
780	11041	Rubella virus	81	100	81	100		
781	11043	Rubella virus strain M33	3	11	3	11		
782	11044	Rubella virus vaccine strain RA27/3	2	6	2	6		
783	11045	Rubella virus strain Therien	20	35	20	35		
784	11047	Equine arteritis virus	3		3			
785	11048	Lactate dehydrogenase-elevating virus	14		14			
786	11049	Lelystad virus	66	12	66	12		
787	11053	dengue virus type 1	91	631	91	641		10
788	11057	Dengue virus 1 Thailand/AHF 82-80/1980		1		1		
789	11059	Dengue virus 1 Nauru/West Pac/1974	6	50	6	50		
790	11060	dengue virus type 2	199	744	201	755	2	11
791	11062	Dengue virus 2 Malaysia M2	1		1			
792	11064	Dengue virus 2 Jamaica/1409/1983	267	1254	267	1254		
793	11065	Dengue virus 2 Thailand/NGS-C/1944	20	146	21	146	1	

794		11066	Dengue virus 2 Puerto Rico/PR159-S1/1969	32	21	32	21		
795		11067	Dengue virus 2 Tonga/EKB194/1974		3		3		
796		11069	dengue virus type 3	129	679	135	695	6	16
797		11070	dengue virus type 4	85	469	86	497	1	28
798		11072	Japanese encephalitis virus	50	173	50	177		4
799		11073	Japanese encephalitis virus strain SA-14	6	2	6	2		
800		11075	Japanese encephalitis virus strain JAOARS982	4	4	4	4		
801		11076	Japanese encephalitis virus strain Nakayama	2	2	2	2		
802		11077	Kunjin virus	12	1	12	1		
803		11078	Kunjin virus (STRAIN MRM61C)		4		4		
804		11079	Murray Valley encephalitis virus	21	9	21	9		
805		11080	Saint Louis encephalitis virus	9	1	9	1		
806		11081	St. Louis encephalitis virus (strain MS1-7)	7		7			
807		11082	West Nile virus	52	451	68	451	16	
808		11083	Powassan virus	14	3	14	3		
809		11084	Tick-borne encephalitis virus	18	147	18	147		
810		11085	Langat virus		1		1		
811		11086	Louping ill virus	1		1			
812		11087	Tick-borne encephalitis virus (STRAIN SOFJIN)	6		6			
813		11089	Yellow fever virus	1855	1365	1855	1365		
814		11090	Yellow fever virus 17D	2	688	2	688		
815		11092	Kumlinge virus	26		26			
816		11096	Classical swine fever virus	17	4	20	4	3	
817		11098	Classical swine fever virus - Brescia	4		4			
818		11099	Bovine viral diarrhea virus 1	2	12	4	12	2	
819		11100	Bovine viral diarrhea virus 1-NADL	4	3	4	3		
820		11104	Hepatitis C virus (isolate 1)	37	256	37	256		
821		11105	Hepatitis C virus (isolate BK)	3	90	3	90		
822		11108	Hepatitis C virus (isolate H)	166	84	166	84		
823		11110	Hepatitis C virus HCT18	1		1			
824		11113	Hepatitis C virus isolate HC-J6	9	3	9	3		
825		11115	Hepatitis C virus isolate HC-J8	1	1	1	1		
826		11116	Hepatitis C virus (isolate Japanese)	6	19	6	19		
827		11120	Infectious bronchitis virus	10	9	11	9	1	
828		11127	Avian infectious bronchitis virus (strain M41)	4	3	4	3		
829		11128	Bovine coronavirus	7		7			
830	X	11135	Feline infectious peritonitis virus			1		1	
831		11137	Human coronavirus 229E	5503	10	5518	20	15	10
832		11138	Murine hepatitis virus	3	31	3	31		
833		11142	Murine hepatitis virus strain A59	26	4	26	4		
834		11144	Murine hepatitis virus strain JHM	25	21	25	21		

835		11149	Transmissible gastroenteritis virus	15	1	15	1		
836		11150	Porcine transmissible gastroenteritis coronavirus strain FS772/70	2		2			
837		11151	Porcine transmissible gastroenteritis coronavirus strain Purdue	14		14			
838		11152	Turkey coronavirus	1		1			
839		11153	Canine coronavirus		3		3		
840		11169	Mumps virus strain Kilham	1		1			
841		11173	Mumps virus strain SBL-1	6		6			
842		11177	Newcastle disease virus (STRAIN AUSTRALIA-VICTORIA/32)	4		4			
843		11178	Newcastle disease virus strain Beaudette C/45	12		12			
844		11180	Newcastle disease virus strain D26/76	4		4			
845		11186	Newcastle disease virus strain Queensland/66	4		4			
846		11194	Sendai virus (strain Enders)	7	11	7	11		
847		11195	Sendai virus (strain Fushimi)		1		1		
848		11196	Sendai virus (strain Harris)	2		2			
849		11198	Sendai virus (Z)		2		2		
850		11214	Human parainfluenza virus 2 (strain Toshiba)	3		3			
851		11216	Human respirovirus 3	3		3			
852		11217	Human parainfluenza 3 virus (strain NIH 47885)	8	12	8	12		
853		11233	Canine distemper virus strain Onderstepoort	6	34	6	34		
854		11234	Measles morbillivirus	64	39	64	39		
855		11235	Measles virus strain Edmonston	167	210	167	210		
856		11236	Measles virus strain Halle		34		34		
857		11241	Rinderpest morbillivirus	2	3	2	3		
858		11243	Rinderpest virus (strain L)	6		6			
859		11246	Bovine orthopneumovirus	3	2	3	2		
860		11249	Bovine respiratory syncytial virus (strain RB94)	4		4			
861		11250	Human orthopneumovirus	36	208	36	208		
862		11251	Human respiratory syncytial virus (subgroup B / strain 18537)	2	1	4	1	2	
863		11252	Human respiratory syncytial virus (strain RSB642)	2		2			
865		11255	Human respiratory syncytial virus (strain RSB6190)	6		6			
866		11256	Human respiratory syncytial virus (strain RSB6256)	11		11			
867		11259	Human respiratory syncytial virus A2	111	245	119	245	8	
868		11260	Human respiratory syncytial virus A strain Long	129	94	129	94		
869		11263	Murine orthopneumovirus		14		14		
870		11276	Vesicular stomatitis virus		10		11		1
871		11277	Vesicular stomatitis Indiana virus	9	6	9	6		
872		11278	Vesicular stomatitis Indiana virus strain Glasgow		1		1		
873		11279	Vesicular stomatitis Indiana virus strain Mudd-Summers	1		1			

875		11285	Vesicular stomatitis Indiana virus strain San Juan		1		1		
877		11288	Viral hemorrhagic septicemia virus 07-71	12	1	12	1		
878		11290	Infectious hematopoietic necrosis virus	8		8			
879		11292	Lyssavirus rabies	15	26	15	26		
880		11293	Rabies virus AVO1		3		3		
881		11294	Rabies virus CVS-11	6		7		1	
882		11295	Rabies virus ERA	10	27	10	27		
883		11296	Rabies virus HEP-FLURY	7	1	7	1		
884		11298	Rabies virus Nishigahara RCEH	1		1			
885		11307	Sonchus yellow net nucleorhabdovirus		1		1		
886		11318	Thogotovirus dhoriense	1		1			
887		11320	Influenza A virus	78	660	85	668	7	8
889		11384	Influenza A virus (A/chicken/FPV/Weybridge(H7N7))		2		2		
890		11408	Influenza A virus (STRAIN A/EQUINE/NEW MARKET/76)		1		1		
891		11448	Influenza A virus (A/parrot/Ulster/73(H7N1))		1		1		
892		11484	Influenza A virus (A/whale/Maine/1/84(H13N9))	3		3			
893		11520	Influenza B virus	5	460	5	463		3
894		11521	Influenza B virus (B/Ann Arbor/1/1986)		3		3		
895		11541	Influenza B virus (B/Oregon/5/80)	16		16			
896		11550	Influenza B virus (B/Yamagata/1/73)		3		3		
897		11552	Influenza C virus		1		2		1
898		11553	Influenza C virus (C/Ann Arbor/1/50)	16		16			
899		11577	La Crosse virus	2	1	2	1		
900		11578	La Crosse virus L74		2		2		
901		11583	HoJo virus		1		1		
902		11588	Rift Valley fever virus	13	67	13	67		
903		11590	Toscana virus	3		3			
904		11602	Hantaan virus 76-118	14	132	14	132		
905		11605	Puumala virus Hallnas B1		1		1		
906		11607	Sapporo rat virus	1	4	1	4		
907		11610	Seoul virus SR11	1		1			
908		11621	Lassa virus GA391		55		55		
909		11622	Lassa virus Josiah	10	451	20	451	10	
910		11624	Lymphocytic choriomeningitis virus (strain Armstrong)		1158		1158		
911		11625	Lymphocytic choriomeningitis virus (strain Pasteur)		5		5		
912		11626	Lymphocytic choriomeningitis virus (strain Traub)		3		3		
913		11627	Lymphocytic choriomeningitis virus (strain WE)		74		74		
914		11636	Reticuloendotheliosis virus	2	5	2	5		
915		11642	Simian foamy virus	1		1			

916		11660	Caprine arthritis encephalitis virus	14	1	14	1		
917		11661	Caprine arthritis encephalitis virus strain Cork	9		9			
918		11662	Caprine arthritis encephalitis virus G63	6		6			
919		11665	Equine infectious anemia virus	93	112	93	112		
920		11670	Equine infectious anemia virus (CLONE 1369)		1		1		
921		11671	Equine infectious anemia virus (STRAIN WSU5)		19		19		
922		11673	Feline immunodeficiency virus		30		30		
923		11674	Feline immunodeficiency virus (isolate Petaluma)	2	4	2	4		
924		11675	Feline immunodeficiency virus (isolate San Diego)		1		1		
925		11676	Human immunodeficiency virus 1	319	537	319	538		1
926		11678	Human immunodeficiency virus type 1 BH10	2	29	2	29		
927		11679	Human immunodeficiency virus type 1 (CLONE 12)		7		7		
929		11682	Human immunodeficiency virus type 1 (BH5 ISOLATE)		1		1		
930		11683	Human immunodeficiency virus type 1 (Z2/CDC-Z34 ISOLATE)		1		1		
932		11685	HIV-1 M:B_ARV2/SF2	1	44	1	44		
933		11686	Human immunodeficiency virus type 1 (BRU ISOLATE)	1	4	1	4		
934		11687	Human immunodeficiency virus type 1 (CDC-451 ISOLATE)		3		3		
935		11688	Human immunodeficiency virus type 1 (JRCSF ISOLATE)	4	37	4	37		
936		11689	Human immunodeficiency virus type 1 (ELI ISOLATE)		7		7		
937		11690	Human immunodeficiency virus type 1 (SF33 ISOLATE)		2		2		
938		11691	Human immunodeficiency virus type 1 (SF162 ISOLATE)	2		2			
939		11694	Human immunodeficiency virus type 1 (JH3 ISOLATE)		3		3		
940		11695	Human immunodeficiency virus type 1 (NDK ISOLATE)		8		8		
941		11696	HIV-1 M:B_MN	11	27	11	27		
942		11697	Human immunodeficiency virus type 1 (MAL ISOLATE)		6		6		
943		11698	Human immunodeficiency virus type 1 (NEW YORK-5 ISOLATE)	2	2	2	2		
944		11699	Human immunodeficiency virus type 1 (OY1 ISOLATE)		3		3		
945		11701	Human immunodeficiency virus type 1 (RF/HAT ISOLATE)	1	19	1	19		
946		11703	Human immunodeficiency virus type 1 (STRAIN UGANDAN / ISOLATE U455)		14		14		
947		11704	Human immunodeficiency virus type 1 (MFA ISOLATE)		2		2		
948		11705	Human immunodeficiency virus type 1 (WMJ2 ISOLATE)		1		1		
949		11706	HIV-1 M:B_HXB2R	15	18	15	18		
950		11708	Human immunodeficiency virus type 1 (ZAIRE 6 ISOLATE)		1		1		
951		11709	Human immunodeficiency virus 2	1	8	1	8		

952		11711	Simian immunodeficiency virus - mac		1		1		
953		11713	Human immunodeficiency virus type 2 (ISOLATE D194)		1		1		
954		11714	Human immunodeficiency virus type 2 (ISOLATE BEN)		12		12		
955		11715	Human immunodeficiency virus type 2 (ISOLATE CAM2)		2		2		
956		11716	Human immunodeficiency virus type 2 (ISOLATE D205,7)		2		2		
957		11717	Human immunodeficiency virus type 2 (ISOLATE GHANA-1)		7		7		
958		11718	Human immunodeficiency virus type 2 (ISOLATE SBLISY)		2		2		
959		11720	Human immunodeficiency virus type 2 (ISOLATE ROD)		2		2		
960		11723	Simian immunodeficiency virus		245		245		
961		11730	Simian immunodeficiency virus (AGM3 ISOLATE)		1		1		
964		11733	Simian immunodeficiency virus (MM142-83 ISOLATE)		102		102		
965		11734	Simian immunodeficiency virus (MM251 ISOLATE)		1		1		
966		11735	Simian immunodeficiency virus - mac K6W		125		125		
967		11736	Simian immunodeficiency virus (K78 ISOLATE)		29		29		
968		11737	Simian immunodeficiency virus (F236/SMH4 ISOLATE) (SOOTY MANGABEY)		17		17		
969		11738	Simian immunodeficiency virus (PBJ/BC13 ISOLATE) (SOOTY MANGABEY)		2		2		
970		11742	Visna lentivirus (strain 1514)	1		1			
971		11743	Visna lentivirus (strain 1514 / clone LV1-1KS1)	4		4			
972		11746	Jaagsiekte sheep retrovirus	1		1			
973		11757	Mouse mammary tumor virus		12		12		
974		11764	Baboon endogenous virus strain M7	1		1			
975		11769	Feline leukemia virus strain A/Glasgow-1	6		6			
976		11780	Snyder-Theilen feline sarcoma virus	1		1			
977		11786	Murine leukemia virus	1	25	1	25		
978		11790	AKT8 murine leukemia virus	1		1			
979		11791	AKR (endogenous) murine leukemia virus		13		13		
980		11795	Friend murine leukemia virus	6	53	6	53		
981		11801	Moloney murine leukemia virus		5		5		
982		11812	Murine sarcoma virus 3611	4		4			
983		11824	Gardner-Arnstein feline leukemia oncovirus B	13		13			
984		11827	Human endogenous retrovirus	1	1	1	1		
985		11861	Avian erythroblastosis virus	1		1			
986		11864	Avian leukosis virus	5		5			
987		11886	Rous sarcoma virus	4	4	4	4		
988		11901	Bovine leukemia virus	108	40	108	40		
990		11908	Human T-cell leukemia virus type I	236	280	236	280		

991		11909	Human T-lymphotropic virus 2	63	2	63	2		
992		11926	Human T-cell lymphotropic virus type 1 (strain ATK)	30	118	30	118		
993		11927	Human T-cell lymphotropic virus type 1 (Caribbean isolate)		6		6		
994		11928	Human T-cell lymphotropic virus type 1 (isolate MT-2)	1		1			
995		11942	Simian retrovirus 1	1		1			
996		11963	Human spumaretrovirus	1		1			
997		11966	Aids-associated retrovirus		2		2		
998		11976	Rabbit hemorrhagic disease virus	4	6	4	6		
999		11978	Feline calicivirus	4	1	4	1		
1000		11980	Feline calicivirus strain Japanese F4	6		7		1	
1001		11981	Feline calicivirus strain F9	2		2			
1002		11983	Norwalk virus	7	8	7	8		
1003		11984	Southampton virus	1		1			
1004		12022	Escherichia phage MS2	1		1			
1005		12060	Echovirus E9	1		1			
1006		12063	Human parechovirus 1	7		7			
1007		12064	Enterovirus E	17		17			
1008		12065	Bovine enterovirus strain VG-5-27	6		6			
1009		12067	Coxsackievirus A9	30		30			
1010		12071	Coxsackievirus B1	13		17		4	
1011		12072	Coxsackievirus B3	7	18	7	18		
1012		12073	Coxsackievirus B4	18	51	18	51		
1013		12075	Swine vesicular disease virus	1		1			
1014		12076	Swine vesicular disease virus (STRAIN H/3 '76)	5		5			
1015		12077	Swine vesicular disease virus (STRAIN UKG/27/72)	3		3			
1016		12078	Echovirus E11	1		1			
1017		12080	Poliovirus 1	6	1	6	1		
1018		12081	Human poliovirus 1 Mahoney	62	31	62	31		
1019		12082	Human poliovirus 1 strain Sabin	8		8			
1020		12086	Poliovirus 3	5	2	5	2		
1021		12088	Poliovirus type 3 (strains P3/LEON/37 AND P3/LEON 12A[1]B)	4	15	4	15		
1022		12092	Hepatovirus A	30	22	30	22		
1023		12098	Human hepatitis A virus Hu/Australia/HM175/1976	87	96	87	96		
1024		12104	Encephalomyocarditis virus		1		1		
1025		12107	Mengo virus	5	13	5	13		
1026		12110	Foot-and-mouth disease virus	33	6	33	6		
1027		12111	Foot-and-mouth disease virus A	3	7	4	7	1	
1028		12112	Foot-and-mouth disease virus (strain A10-61)	8		8			

102 9		12113	Foot-and-mouth disease virus (strain A5)	3		3			
103 0		12114	Foot-and-mouth disease virus (strain A12)	21	1	21	1		
103 1		12115	Foot-and-mouth disease virus (strain A24 Cruzeiro)	17	32	17	32		
103 2		12116	Foot and mouth disease virus C	6	26	6	26		
103 3		12117	Foot-and-mouth disease virus (strain C3 Indaial)	2		2			
103 4		12118	Foot-and-mouth disease virus O	49	13	50	13	1	
103 5		12120	Foot-and-mouth disease virus (strain C1-Santa Pau)	1		1			
103 6		12121	Foot-and-mouth disease virus C1	1	4	1	4		
103 7		12123	Foot-and-mouth disease virus SAT 3		1		1		
103 8		12124	Theiler's encephalomyelitis virus	1	18	1	18		
103 9		12125	Theiler's encephalomyelitis virus (STRAIN BEAN 8386)	16	52	16	52		
104 0		12126	Theiler's encephalomyelitis virus (STRAIN DA)	3	2	3	2		
104 1		12130	rhinovirus A2	11		11			
104 2		12131	rhinovirus B14	17	1	17	1		
104 3		12132	rhinovirus A89	3		7		4	
104 4		12134	Human rhinovirus 1A		19		19		
104 5		12146	Tomato bushy stunt virus (STRAIN BS-3)	3		3			
104 6		12161	Beet yellows virus	5		5			
104 7		12162	Citrus tristeza virus	8		8			
104 8		12211	Plum pox virus	1	1	1	1		
104 9		12213	Plum pox virus isolate NAT		1		1		
105 0		12216	Potato virus Y	2		2			
105 1		12219	Potato virus Y strain N	1		1			
105 2		12220	Potato virus Y strain O	7		7			
105 3		12232	Zucchini yellow mosaic virus	1		1			
105 4		12242	Tobacco mosaic virus	142	13	142	13		
105 5		12243	Tobacco mosaic virus (vulgare)	1		1			
105 6		12246	Tobacco mosaic virus strain Dahlemense	2		2			
105 7		12260	Bean pod mottle virus	2		2			
105 8		12264	Cowpea mosaic virus		1		1		
105 9		12274	Grapevine fanleaf virus	1		1			
106 0		12305	Cucumber mosaic virus	1		1			
106 1		12455	Borna disease virus	8	4	8	4		
106 2		12475	Hepatitis delta virus	56	66	56	67		1
106 3		12509	Human herpesvirus 4 type 2	2	10	2	10		
106 4		12557	Seoul virus 80-39		3		3		
106 5		12618	Chicken anemia virus	3		3			

106 6		12637	Dengue virus	4429	588	4429	588		
106 7		12639	Duck hepatitis B virus	262	20	262	20		
106 8		12643	Ectromelia virus		45		45		
106 9		12663	Feline coronavirus	64	2	65	3	1	1
107 0		12701	Human astrovirus 2	2		2			
107 1		12721	Human immunodeficiency virus		58		58		
107 2		12750	Visna/maedi virus EV1	1		1			
107 3		12760	Murine hepatitis virus strain 4		5		5		
107 4		12814	Respiratory syncytial virus	5	5	5	5		
107 5		12870	Variola major virus		17		17		
107 7		13101	Juniperus ashei	13		13			
107 8		13187	Parietaria officinalis	2		2			
107 9		13286	Theromyzon tessulatum	1		1			
108 0		13373	Burkholderia mallei	3	50	3	50		
108 1		13415	Chamaecyparis obtusa	5	68	5	68		
108 2		13451	Corylus avellana	33	57	35	57	2	
108 3		13469	Cupressus sempervirens		8		8		
108 4		13557	Hapalemur griseus		1		1		
108 5		13616	Monodelphis domestica	1	4	1	4		
108 6		13618	Myrmecia pilosula	2		2			
108 7		13677	Scomber scombrus	1		1			
108 8		13684	Parastagonospora nodorum		2		2		
108 9		13687	Sphingomonas	1	5	1	5		
109 1	X	13818	<i>Adiantum capillus-veneris</i>			1		1	
109 2		15368	Brachypodium distachyon	1		1			
109 3		15957	Phleum pratense	64	1158	64	1158		
109 4		16719	Juglans nigra	1		1			
109 6		27990	Plasmodium vivax-like sp.	3	1	3	1		
109 7	X	28035	<i>Staphylococcus lugdunensis</i>				5		5
109 8		28037	Streptococcus mitis		1		1		
109 9		28038	Latilactobacillus curvatus		1		1		
110 0		28080	Campylobacter upsaliensis	3		3			
110 1		28108	Alteromonas macleodii		1		1		
110 2	X	28116	<i>Bacteroides ovatus</i>				6		6
110 3		28130	Prevotella disiens		1		1		
110 4		28131	Prevotella intermedia		2		2		
110 7		28151	Serratia proteamaculans		1		1		
110 8		28172	Vibrio metschnikovii	1		1			
110 9		28227	Malacoplasma penetrans	40		40			

111 0		28276	Human adenovirus 15	1		1			
111 1		28280	Human adenovirus E4	1		1			
111 2		28282	Human adenovirus 12	11	3	11	3		
111 3		28284	Human adenovirus 40	2	1	2	1		
111 4		28285	Human adenovirus 5	27	186	27	186		
111 5		28295	Porcine epidemic diarrhea virus	16		77		61	
111 6		28300	Heron hepatitis B virus		1		1		
111 7		28314	Aleutian mink disease virus	10		10			
111 8		28327	Murine rotavirus		3		3		
111 9		28344	Porcine reproductive and respiratory syndrome virus	90	163	97	163	7	
112 0		28375	Soil-borne wheat mosaic virus	1		1			
112 1		28446	Anaerotignum propionicum		2		2		
112 2		28448	Komagataeibacter xylinus		1		1		
112 3		28450	Burkholderia pseudomallei	65	13	65	13		
112 4		28479	Phalaris aquatica		12		12		
112 5		28869	Ovine respiratory syncytial virus	1		1			
112 6		28871	Taterapox virus		4		4		
112 7		28873	Camelpox virus		2		2		
112 8		28875	Rotavirus A	2	2	2	2		
112 9		28901	Salmonella enterica		147		147		
113 0		28903	Mycoplasmopsis bovis	21		21			
113 1		28909	Cynodon dactylon	27	48	27	48		
113 2		29073	Ursus maritimus	1		1			
113 3		29159	Crassostrea gigas	7		26		19	
113 4		29170	Ancylostoma caninum	1		1			
113 5		29176	Neospora caninum	3	17	3	17		
113 6		29271	Dasheen mosaic virus	1		1			
113 7		29292	Pyrococcus abyssi		1		1		
113 8		29320	Paenarthrobacter nicotinovorans	1		1			
113 9		29339	Bacillus thuringiensis serovar kurstaki	3		3			
114 0		29355	Ruminiclostridium cellobioparum	1		1			
114 1		29362	Ruminiclostridium papyrosolvans		1		1		
114 2	X	29380	Staphylococcus caprae				173		173
114 3		29388	Staphylococcus capitis	1		1	17		17
114 4		29430	Acinetobacter haemolyticus	1		1			
114 5		29438	Pseudomonas savastanoi		1		1		
114 6		29442	Pseudomonas tolaasii		1		1		
114 7		29447	Xanthomonas albilineans		1		1		

114 8		29459	Brucella melitensis	19	95	19	95		
114 9		29461	Brucella suis	1	8	1	8		
115 0	X	29466	Veillonella parvula				2		2
115 1		29477	Salmonella enterica subsp. enterica serovar Essen	1		1			
115 2		29491	Aeromonas salmonicida subsp. salmonicida		1		1		
115 3		29507	Leptospira kirschneri	1		1			
115 4		29518	Borrelia afzelii	6	1	6	1		
115 5		29519	Borrelia garinii	6	29	6	29		
115 6	X	29523	Bacteroides sp.				1		1
115 7		29661	Anthoxanthum odoratum		3		3		
115 8		29679	Holcus lanatus	14	3	14	3		
115 9		29715	Ambrosia psilostachya		10		10		
116 0		29760	Vitis vinifera	1		1			
116 1		29780	Mangifera indica	1		1			
116 2		29908	Sporothrix schenckii		2		2		
116 3		29916	Fusarium sp.	3		3			
116 4		29918	Cladosporium herbarum		18		18		
116 5		29960	Penaeus indicus	2		2			
116 6		30011	Pyrophorus plagiophthalmus		1		1		
116 7		30069	Anopheles stephensi	1		1			
116 9	X	30522	Bos indicus x Bos taurus			1			1
117 0		30538	Vicugna pacos	2		2			
117 1		31271	Plasmodium chabaudi chabaudi		3		3		
117 2		31273	Plasmodium vivax strain Belem	9	31	9	31		
117 3		31276	Perkinsus marinus		1		1		
117 4		31285	Trypanosoma brucei gambiense	2		2			
117 5		31286	Trypanosoma brucei rhodesiense	1	9	1	9		
117 6		31330	Ephydatia fluviatilis		1		1		
117 7		31525	Human herpesvirus 4 strain CAO		1		1		
117 8		31531	Vaccinia virus L-IPV		8		8		
117 9		31545	Human adenovirus D8	1	1	1	1		
118 0		31552	Human papillomavirus type 6	12	4	12	4		
118 1		31560	Infectious bursal disease virus E	1		1			
118 2		31569	Human rotavirus (serotype 2 / strain RV-5)	1		1			
118 3		31608	Simian virus 5 (isolate canine/CPI+)	1		1			
118 4		31611	Bovine respiratory syncytial virus (strain 391-2)	3		3			
118 5		31615	Tacaribe virus strain V5		1		1		
118 6		31616	Tacaribe virus strain V7		1		1		

118 7	31621	Four Corners hantavirus	2	8	2	8		
118 8	31631	Human coronavirus OC43	9397	11	9409	23	12	12
118 9	31634	Dengue virus 2 Thailand/16681/84	36	62	42	62	6	
119 0	31635	Dengue virus 2 16681-PDK53	4	80	4	80		
119 1	31636	Dengue virus 2 China/D2-04		5		5		
119 2	31637	Dengue virus 2 Thailand/TH-36/1958		4		4		
119 3	31641	Yellow fever virus 1899/81		36		36		
119 4	31644	Hepatitis C virus HCV-KF	1	1	1	1		
119 5	31645	Hepatitis C virus (isolate Taiwan)	3	15	3	15		
119 6	31647	Hepatitis C virus subtype 1b	612	277	612	277		
119 7	31649	Hepatitis C virus subtype 2a	80	22	80	22		
119 8	31650	Hepatitis C virus subtype 2b	103	4	103	4		
119 9	31653	Hepatitis C virus subtype 4a	1	1	1	1		
120 0	31654	Hepatitis C virus subtype 5a	1	1	1	1		
120 1	31655	Hepatitis C virus subtype 6a		1		1		
120 2	31678	Human immunodeficiency virus type 1 (WMJ1 isolate)		16		16		
120 4	31682	Simian immunodeficiency virus - mac1A11		18		18		
120 5	31683	Simian immunodeficiency virus - stm		7		7		
120 6	31684	Simian immunodeficiency virus (isolate AGM / clone GRI-1)		2		2		
120 8	31704	Coxsackievirus A16	5	1	5	1		
120 9	31708	rhinovirus A16	9	30	9	30		
121 0	31715	Bean-pod mottle virus (strain Kentucky G7)	9	1	9	1		
121 1	31721	Beet necrotic yellow vein virus	10		10			
121 2	31767	Hepatitis E virus (strain Burma)	160		160			
121 3	31768	Hepatitis E virus (strain Mexico)	31		31			
121 4	32008	Burkholderia	1		1			
121 5	32019	Campylobacter fetus subsp. fetus	3		3			
121 6	32022	Campylobacter jejuni subsp. jejuni	2		2			
121 7	32025	Helicobacter hepaticus		1		1		
121 8	32049	Picosynechococcus sp. PCC 7002		1		1		
121 9	32201	Carya illinoensis	19		43		24	
122 0	32278	Metapenaeus ensis	1	6	1	6		
122 1	32603	Human betaherpesvirus 6A	2	7	2	7		
122 2	32604	Human betaherpesvirus 6B	2	4533	2	4533		
122 3	32605	Buffalopox virus		1		1		
122 4	32606	Rabbitpox virus		2		2		
122 5	32614	Convict Creek 107 virus	1	3	1	3		
122 7	32644	unidentified		56955		64533		7578

122 8	X	33025	Phascolarctobacterium faecium				1		1
123 0	X	33039	[Ruminococcus] torques				1		1
123 2		33075	Acidobacterium capsulatum		1		1		
123 3		33090	Viridiplantae	1		1			
123 4		33127	Parietaria judaica	65	2	65	2		
123 5		33178	Aspergillus terreus	2		2			
123 6		33703	Suid herpesvirus 1 strain Kaplan	4		4			
123 7		33706	Caviid betaherpesvirus 2	3	3	3	3		
123 8	X	33707	Marek's disease herpesvirus (strain RB-1b)				4		4
123 9		33708	Murid gammaherpesvirus 4	1	54	1	54		
124 0		33717	Bluetongue virus (serotype 13 / isolate USA)	2		2			
124 1		33718	Bluetongue virus (serotype 17 / isolate USA)	4		4			
124 2		33727	Marburg virus - Musoke, Kenya, 1980	3	84	3	84		
124 3		33728	Lake Victoria marburgvirus - Popp	3	15	3	15		
124 4		33734	Feline infectious peritonitis virus (strain 79-1146)	15	14	15	14		
124 5		33741	Dengue virus 1 Singapore/S275/1990		244	15	244	15	
124 6		33745	Hepatitis C virus genotype 4	1	10	1	10		
124 7		33746	Hepatitis C virus genotype 5		2		2		
124 8		33747	Simian T-lymphotropic virus 1		14		14		
124 9		33892	Mycobacterium tuberculosis variant bovis BCG	13	296	13	296		
125 0		33934	Anoxybacillus flavithermus		1		1		
125 1		33959	Lactobacillus johnsonii	1		1			
125 3		33990	Rickettsia bellii	1	3	1	3		
125 4		34054	Yersinia enterocolitica (type O:8)		1		1		
125 5		34245	Zinnia elegans		1		1		
125 6		34610	Amblyomma variegatum	1		69		68	
125 7		34613	Ixodes ricinus	2		2			
125 8		34631	Rhipicephalus appendiculatus	2		70		68	
125 9		34632	Rhipicephalus sanguineus	1		1			
126 0		34828	Eulemur mongoz	1		1			
126 1		34862	Otospermophilus beecheyi		1		1		
126 2		35241	Lactococcus phage Tuc2009	1		1			
126 3		35258	Lambdapapillomavirus 2		25		25		
126 4		35269	Woodchuck hepatitis virus	4	6	4	6		
126 5		35275	Murine endogenous retrovirus		14		14		
126 6		35288	Grapevine virus A	12		12			
126 7		35292	Foot-and-mouth disease virus SAT 2	1	1	1	1		

126 8	35293	Echovirus E12		1		1		
126 9	35297	Striped jack nervous necrosis virus	1		1			
127 0	35305	California encephalitis virus		1		1		
127 1	35327	Bluetongue virus 1	5		5			
127 2	35329	Bluetongue virus 11	6		6			
127 3	35330	Bluetongue virus 13	1		1			
127 4	35331	Bluetongue virus 15	7		7			
127 5	35336	Rotavirus G4		1		1		
127 6	35345	Lactococcus phage TP901-1	3		3			
127 7	35670	Naja naja	3		3			
127 8	35725	Macrophomina phaseolina		1		1		
127 9	35788	Rickettsia africae		1		1		
128 0	35791	Rickettsia massiliae		1		1		
128 2	35793	Rickettsia sibirica		6		6		
128 3	35795	Ehrlichia muris	2	3	2	3		
128 5	36080	Mucor circinelloides		3		3		
128 6	36087	Trichuris trichiura	15		15			
128 7	36329	Plasmodium falciparum 3D7	1183	392	1197	392	14	
128 8	36351	Human herpesvirus 6 strain Z29	1	146	1	146		
128 9	36352	Human herpesvirus 4 type 1	4	14	4	14		
129 0	36372	Feline immunodeficiency virus (strain UK8)		1		1		
129 1	36374	Visna/maedi virus EV1 KV1772	2		2			
129 2	36375	Human immunodeficiency virus type 1 (KB-1 isolate)		2		2		
129 3	36386	Louping ill virus (strain 31)	1		1			
129 4	36409	Rinderpest virus (strain RBOK)	33	5	33	5		
129 5	36412	Human parainfluenza 1 virus (strains A1426 / 86-315 / 62M-753)		1		1		
129 6	36418	Influenza A virus (A/chicken/Brescia/1902(H7N7))		1		1		
129 7	36420	H1N1 swine influenza virus		13		13		
129 8	36421	African horse sickness virus 4	42	4	42	4		
129 9	36439	Bovine rotavirus strain NCDV/G6	1		1			
130 0	36470	Streptococcus sp. 'group A'	7		7			
130 1	36596	Prunus armeniaca	4		4			
130 2	36809	Mycobacteroides abscessus		19		19		
130 3	36826	Clostridium botulinum A	46		46			
130 4	36827	Clostridium botulinum B	29		29			
130 5	36829	Clostridium botulinum D	2		2			
130 6	36830	Clostridium botulinum E	3	3	3	3		
130 7	36831	Clostridium botulinum F	3		3			

130 8	X	36845	<i>Clostridium intestinale</i>				2		2
130 9		36855	<i>Brucella canis</i>		3		3		
131 0		36911	<i>Clavispora lusitanae</i>	1	1	1	1		
131 1		36914	<i>Lodderomyces elongisporus</i>		3		3		
131 2		36936	<i>Lepidoglyphus destructor</i>	5	10	5	10		
131 4		37111	Human papillomavirus 28		1		1		
131 5		37112	Human papillomavirus 29		1		1		
131 6		37115	human papillomavirus 59	2	1	2	1		
131 7		37120	human papillomavirus 67	2		2			
131 8		37121	human papillomavirus 69		1		1		
131 9		37124	Chikungunya virus	87	22	89	22	2	
132 0		37128	Potato mop-top virus	8		8			
132 1		37132	Rabies virus Ontario fox	3		3			
132 2		37137	Simian rotavirus A/SA11-both	20	2	20	2		
132 3		37207	Rio Segundo hantavirus	1		1			
132 4		37296	Human gammaherpesvirus 8	26	489	26	489		
132 5		37325	Muscovy duck parvovirus	11		11			
132 6		37326	<i>Nocardia brasiliensis</i>	6		6			
132 7		37329	<i>Nocardia farcinica</i>		1		1		
132 8	X	37332	<i>Nocardia seriolae</i>			4		4	
132 9		37347	<i>Tupaia belangeri</i>		1		1		
133 0		37546	<i>Glossina morsitans morsitans</i>	2		2			
133 2		37731	<i>Secale strictum</i> subsp. <i>africanum</i>		2		2		
133 3		37734	<i>Enterococcus casseliflavus</i>		1		1		
133 4		37762	<i>Escherichia coli</i> B	1		1			
133 5		37769	<i>Cryptococcus gattii</i> VGI		1		1		
133 6		38020	<i>Callitrichinae</i> sp.	3		3			
133 7		38033	<i>Chaetomium globosum</i>	1		1			
133 8		38171	Avian reovirus strain S1133	6		6			
133 9		38251	Goose parvovirus	10		10			
134 0		38289	<i>Corynebacterium jeikeium</i>		1		1		
134 1		38323	<i>Bartonella henselae</i>	1		1			
134 2		38873	<i>Fraxinus excelsior</i>		3		3		
134 3		38973	Influenza A virus (A/Memphis/4/1973(H3N2))		2		2		
134 4		39002	Puumala virus sotkamo/v-2969/81	196		196			
134 5		39015	Human T-cell lymphotropic virus type 1 (african isolate)		2		2		
134 6		39054	Enterovirus A71	129	1	129	1		
134 8		39152	<i>Methanococcus maripaludis</i>	1		1			

134 9		39414	Plantago lanceolata		2		2		
135 0		39442	Mus musculus musculus		1		1		
135 1		39457	Human papillomavirus type 70		1		1		
135 3		39584	Juniperus virginiana	1		1			
135 4	X	39778	Veillonella dispar				5		5
135 5		39803	Qubevirus durum		2		2		
135 6		39947	Oryza sativa Japonica Group	5	1	11	1	6	
135 7	X	39950	Dialister pneumosintes				2		2
135 8		40051	Bluetongue virus	12	1	12	1		
135 9		40214	Acinetobacter johnsonii		1		1		
136 0		40216	Acinetobacter radioresistens		1		1		
136 1		40271	Hepatitis C virus genotype 2	42	6	42	6		
136 2		40324	Stenotrophomonas maltophilia	1		1			
136 3		40674	Mammalia	6	2	6	2		
136 4		40697	Blomia tropicalis	31	6	31	6		
136 5		41514	Salmonella enterica subsp. arizonae serovar 62:z4,z23:-		4		4		
136 6		41846	Echovirus E30	2		2			
136 7		41856	Hepatitis C virus genotype 1	27	178	27	178		
136 8		41857	Influenza A virus H3N2	6	50	6	50		
136 9		41858	Simian foamy virus-gorilla	1		1			
137 0		41953	Pseudo-nitzschia	1		1			
137 1		41997	Enterococcus saccharolyticus		1		1		
137 2		42097	Isla Vista hantavirus	1	2	1	2		
137 3		42182	Hepatitis C virus genotype 6		7		7		
137 4		42229	Prunus avium	3	1	3	1		
137 5		42345	Phoenix dactylifera		3		3		
137 6		42358	Hantavirus Monongahela-3		5		5		
137 7		42374	Candida dubliniensis		2		2		
137 8		42769	Coxsackievirus A10	4		4			
137 9		42782	Coxsackievirus A20	1		1			
138 2		42789	enterovirus D68	16		16			
138 3		42792	Hepatitis C virus subtype 3g	1		1			
138 4		42862	Rickettsia felis		1		1		
138 5		42897	Shigella flexneri 2a	36		36			
138 6		43304	Mycolicibacterium peregrinum	2		2			
138 7	X	43335	Populus alba			1		1	
138 8	X	43358	Human astrovirus 8			2		2	
139 0		43765	Corynebacterium amycolatum		1		1		

139 1		43767	Prescottella equi	18	3	18	3		
139 2		43852	Toxicodendron	1	1	1	1		
139 3		43853	Toxicodendron radicans	1		1			
139 4		44026	Sepik virus		1		1		
139 5		44088	Canarypox virus		8		8		
139 6		44104	Vibrio cholerae 569B	37		37			
139 7		44271	Leishmania chagasi	3		3			
139 8		44275	Leptospira interrogans serovar Copenhageni	2		2			
139 9		44276	Leptospira interrogans serovar Pomona	2	3	2	3		
140 0		44386	Haemaphysalis longicornis	1		1			
140 1		44561	Murine type C retrovirus		1		1		
140 2		44689	Dictyostelium discoideum	1	1	1	1		
140 3		44755	New York hantavirus		2		2		
140 4		45029	Bluetongue virus 16	3		3			
140 5		45201	Mannheimia haemolytica serotype 1	82		82			
140 6		45240	human papillomavirus 68		1		1		
140 7		45409	Feline immunodeficiency virus (isolate wo)	3		3			
140 8		45410	Hepatitis B virus adw4/Brazil/isolate w4b		12		12		
140 9		45455	Macacine gammaherpesvirus 4		27		27		
141 0		45582	[Candida] saitoana	1		1			
141 1		45617	Human endogenous retrovirus K	14		14			
141 2		45659	Human adenovirus B3	29	2	29	2		
141 4		46015	Autographa californica nucleopolyhedrovirus		1		1		
141 5		46221	Porcine circovirus	1	49	1	49		
141 6		46242	Spodoptera litura nucleopolyhedrovirus		1		1		
141 7		46245	Drosophila pseudoobscura pseudoobscura	1		1			
141 8		46290	Foot-and-mouth disease virus C3	4	1	4	1		
141 9		46457	Cycloclasticus oligotrophus		1		1		
142 1	X	46503	Parabacteroides merdae				1		1
142 2		46506	Bacteroides stercoris		1		1		
142 3		46771	Simian virus 12	1		1			
142 4		46835	Fasciola gigantica	3		3			
142 5		46920	Rio Mamore hantavirus	1	1	1	1		
142 6		46921	Human adenovirus D13	1		1			
142 7		46941	Human adenovirus 46	1		1			
142 9		47466	Borrelia miyamotoi	73		73			
143 0		47516	Echovirus E3	1		2		1	

143 2	X	47845	Brachybacterium alimentarium				15		15
143 3		47929	Macacine betaherpesvirus 3		101		101		
143 4		48409	Salmonella enterica subsp. enterica serovar Virchow		1		1		
143 5		48483	Reclinomonas americana		1		1		
143 6		48935	Novosphingobium aromaticivorans		1		1		
143 7		49011	Hesperocyparis arizonica	2	6	2	6		
143 8		49511	Piper longum	1		1			
143 9	X	49827	Glycyrrhiza glabra			1		1	
144 1		50557	Insecta	7		7			
144 2		51022	Ancylostoma duodenale	8		8			
144 3		51031	Necator americanus	25		26		1	
144 4		51033	human papillomavirus 73		1		1		
144 5		51240	Juglans regia	34	19	34	19		
144 6		53182	Feline foamy virus	7		7			
144 7		53258	Variola minor virus		93		93		
144 8		53326	Ancylostoma ceylanicum	3		3			
144 9		53751	Echinacea purpurea	1		1			
145 0		54290	GB virus C	16		16			
145 1		54315	Bovine viral diarrhea virus 2	1		1			
145 2		54388	Salmonella enterica subsp. enterica serovar Paratyphi A		4		4		
145 3		54390	Micrurus corallinus	125		125			
145 4		54736	Salmonella bongori		1		1		
145 6		55513	Pistacia vera		6		6		
145 7		55532	Influenza A virus (A/swine/Quebec/1192/1986(H1))		1		1		
145 8		55601	Vibrio anguillarum	1		1			
145 9		55635	Inula helenium		1		1		
146 0		55951	Grapevine leafroll-associated virus 3	1		1			
146 1		56636	Aeropyrum pernix	3	2	3	2		
146 2	X	56665	Fusarium kyushuense			1		1	
146 3		56950	Border disease virus strain Moredun	3		3			
146 4		57068	Acanthisitta chloris		1		1		
146 5		57266	Plasmodium falciparum 7G8	43	59	43	59		
146 6		57270	Plasmodium falciparum Palo Alto/Uganda	38		38			
146 7		57278	Human herpesvirus 7 strain JI		1		1		
146 9		57372	Mycoplasma suis	3		3			
147 0		57482	European bat 1 lyssavirus		1		1		
147 1		57486	Mus musculus molossinus		1		1		
147 2	X	57579	Adeno-associated virus - 4				4		4

147 3		57667	Simian-Human immunodeficiency virus	1	5	1	5		
147 4		57678	Leptospira interrogans serovar Lai	10	10	10	10		
147 5		57975	Burkholderia thailandensis		2		2		
147 6		58024	Spermatophyta	4	1	4	1		
147 7	X	58029	Cephalotaxus harringtonia			1		1	
147 8		58216	Loxosceles gaucho	1		1			
147 9		58217	Loxosceles laeta	2		2			
148 0		58218	Loxosceles intermedia	83		83			
148 1		58291	Rhizopus microsporus		1		1		
148 2		59201	Salmonella enterica subsp. enterica		18		18		
148 3		59202	Salmonella enterica subsp. salamae		1		1		
148 4		59203	Salmonella enterica subsp. arizonae		1		1		
148 5		59205	Salmonella enterica subsp. houtenae		1		1		
148 6		59300	Getah virus	9		9			
148 7		59301	Mayaro virus	11		11			
148 8		59375	Influenza A virus (A/South Carolina/1/1918(H1N1))	9	1	9	1		
148 9		59538	Pantholops hodgsonii		1		1		
149 0		59729	Taeniopygia guttata		1		1		
149 1	X	59779	Paracoccus marcusii				1		1
149 2		59799	Angomonas deanei		2		2		
149 3		60189	Rhipicephalus decoloratus	6		69		63	
149 5		60552	Burkholderia vietnamiensis		2		2		
149 6		60876	Pixuna virus		1		1		
149 7		60879	Cabassou virus		1		1		
149 8		60893	Desulfobacca acetoxidans		1		1		
149 9	X	61235	Fusarium equiseti			1		1	
150 0		61466	Gnathostoma binucleatum	10		10			
150 1		61673	Porcine endogenous retrovirus	2	11	2	11		
150 2		62319	Halococcus saccharolyticus		1		1		
150 3		62322	Shewanella baltica		1		1		
150 4		62330	Fagopyrum tataricum	8		8			
150 5		62446	Influenza A virus (A/Argentina/3779/94(H3N2))		1		1		
150 6		62478	Influenza A virus (A/Kitakyushu/93(H3N2))		1		1		
150 7		62488	Influenza A virus (A/Nanchang/58/1993(H3N2))		1		1		
150 8		62496	Influenza A virus (A/New_York/17/94(H3N2))		1		1		
150 9		62503	Influenza A virus (A/Ohio/3/95(H3N2))		1		1		
151 0		62512	Influenza A virus (A/Shangdong/5/94(H3N2))		1		1		

151 1		62541	Influenza A virus (A/Christchurch/2/1988(H3N2))		1		1		
151 2		62549	Influenza A virus (A/Fukuoka/C29/1985(H3N2))	5		5			
151 3		62559	Influenza A virus (A/Los Angeles/1987(H3N2))	5		5			
151 4		62564	Influenza A virus (A/New_York/15/94(H3N2))		1		1		
151 5		62575	Influenza A virus (A/Sichuan/2/1987(H3N2))		5		5		
151 6		62596	Influenza A virus (A/USSR/26/1985(H3N2))		1		1		
151 8		63106	Influenza A virus (A/Wuhan/359/1995(H3N2))	5		5			
151 9		63363	Aquifex aeolicus	2	7	2	7		
152 0		63421	Swine hepatitis E virus	12		12			
152 1		63673	Turbo cornutus	2		2			
152 2		63737	Nostoc punctiforme PCC 73102		2		2		
152 3		63746	Hepatitis C virus (isolate H77)	183	210	183	210		
152 4		64284	Saboya virus		1		1		
152 5		64289	Carey Island virus		2		2		
152 6		64293	Tembusu virus	3	11	4	24	1	13
152 7		64320	Zika virus	1279	747	1281	747	2	
152 8		64382	Human Endogenous Retrovirus IDDMK1,2-22	3		3			
152 9		64495	Rhizopus arrhizus	4	1	4	1		
153 0		65690	AK7 murine leukemia virus		1		1		
153 1		65699	Neisseria meningitidis serogroup A	5		5			
153 2		65743	Blackcurrant reversion virus	2		2			
153 3		66692	Shouchella clausii KSM-K16		1		1		
153 4		66976	Legionella pneumophila serogroup 1		7		7		
153 5		67351	Streptomyces californicus	1		1			
153 7		68621	Classical swine fever virus strain Riems		5		5		
153 8		68825	Rasamsonia emersonii		1		1		
153 9		68887	Torque teno virus	5	94	5	94		
154 0		68909	Deinococcus geothermalis	1		1			
154 1		69008	Juniperus oxycedrus		3		3		
154 2		69153	Human enterovirus 71 (strain BRCR)	1		1			
154 3		69156	Murine cytomegalovirus (strain K181)	2	15	2	15		
154 4	X	69218	Enterobacter cancerogenus				3		3
154 5		69245	Lechiguanas virus		9		9		
154 7		69247	Oran virus		2		2		
154 8		69248	Pergamino virus		1		1		
154 9		69820	Spodoptera litura		1		1		
155 0		70146	Measles virus strain Edmonston-B	2	78	2	78		

155 1		70149	Measles virus strain Edmonston-Zagreb	1		1			
155 2		70175	Androctonus australis hector	24		24			
155 3		70203	Fibrovirus fs1		1		1		
155 4		70865	Murine rotavirus EDIM		27		27		
155 5		71238	Pseudomonas sp. G-179		1		1		
155 6		71366	Staphylococcus phage PVL	1		1			
155 7		71421	Haemophilus influenzae Rd KW20		3		3		
156 0		72132	Rotavirus G1	1	1	1	1		
156 1		72539	Physalis mottle virus	2		2			
156 2		72556	Achromobacter piechaudii	1		1			
156 3		72590	Salmonella sp. 'group B'	6		6			
156 4		72664	Eutrema salsugineum	2		2			
156 5		73036	Rotavirus G3		1		1		
156 6		73239	Plasmodium yoelii yoelii	23	42	23	42		
156 7		73482	Foot-and-mouth disease virus (strain O1)	38	1	38	1		
156 8		73484	Human immunodeficiency virus type 2 (isolate KR)		3		3		
156 9	X	74437	Alectryonella plicatula			9		9	
157 0		74537	Vladivostok virus	1		1			
157 1		74722	Stachybotrys chartarum	11		11			
157 2		74942	Hantavirus CRF355		1		1		
157 4		75325	Orf virus strain D1701		36		36		
157 5	X	75658	Campylobacter lanienae				2		2
157 6		75922	Mycolicibacterium tusciae		2		2		
157 8		76777	Malassezia sympodialis	1		1			
157 9		76832	Myroides odoratimimus		1		1		
158 2		77009	Hordeum vulgare subsp. spontaneum		2		2		
158 3		77044	Rosellinia necatrix		1		1		
158 4		77095	Segatella bryantii		2		2		
158 5		77153	Muscovy duck reovirus	1		1			
158 6		77643	Mycobacterium tuberculosis complex		5		5		
158 7		78245	Xanthobacter autotrophicus Py2		1		1		
158 8	X	79327	Schmidtea mediterranea			1		1	
158 9		79692	Human respiratory syncytial virus B1	1	25	1	25		
159 0		79695	Influenza A virus (A/Memphis/1/71H-A/Bellamy/42N)		1		1		
159 1		79698	Merluccius bilinearis	1		1			
159 2		79889	Bovine herpesvirus type 1.1	2		2			
159 3		79923	Clonorchis sinensis	2	1	2	1		
159 4		80365	Laminaria digitata	2		2			

159 5		80859	Streptomyces ribosidificus		1		1		
159 6		80956	Pomacentrus moluccensis		1		1		
159 8		81475	Frateuria aurantia	1		1			
159 9		81847	Trichophyton quinckeanum	1		1			
160 0		81985	Capsella rubella	1		1			
160 1		82070	Squilla maritima	1		1			
160 2		82300	adeno-associated virus 5	9		9			
160 3		82348	Streptococcus pluranimalium		1		1		
160 4		82372	Influenza A virus (A/Sydney/05/97-like(H3N2))		1		1		
160 5		82470	Bovine viral diarrhea virus strain Oregon C24V	5	3	5	3		
160 6		82639	Coxsackievirus B2	2		2			
160 7		82658	Lordsdale virus		1		1		
160 8		82659	Sapporo virus-Manchester		1		1		
160 9		82823	Bovine respiratory syncytial virus strain Ielystad	1		1			
161 0		82824	Bovine respiratory syncytial virus strain snook	1	75	1	75		
161 1		82830	Epstein-barr virus strain ag876	16	1	16	1		
161 2		82831	Equid herpesvirus type 2 strain 86/87	1		1			
161 4		83192	Topografov hantavirus	1		1			
161 5		83206	Influenza A virus (A/swine/Scotland/410440/94(H1N2))		5		5		
161 6		83262	Mycobacteroides immunogenum		5		5		
161 7		83331	Mycobacterium tuberculosis CDC1551		42		42		
161 8		83332	Mycobacterium tuberculosis H37Rv	62	990	63	1007	1	17
161 9		83333	Escherichia coli K-12	33	19	34	19	1	
162 0		83334	Escherichia coli O157:H7	3	123	3	123		
162 1		83455	Myxococcus stipitatus		1		1		
162 2		83554	Chlamydia psittaci	102	6	102	6		
162 3		83555	Chlamydia abortus	32		32			
162 4		83556	Chlamydia felis	12		12			
162 5		83557	Chlamydia caviae	14		14			
162 6		83558	Chlamydia pneumoniae	111	24	111	24		
162 7		83559	Chlamydia suis	19		19			
162 8		83560	Chlamydia muridarum	17	3	17	3		
162 9		83810	Thosea asigna virus		1		1		
163 0	X	83816	Methanobrevibacter ruminantium			7	8	7	8
163 1		85057	Trypanosoma cruzi cruzi		1		1		
163 2		85106	Adeno-associated virus - 1		11		17		6
163 3		85223	Laurus nobilis		1		1		

163 4	X	85363	<i>Aconitum carmichaelii</i>			1		1	
163 5		85552	<i>Scylla paramamosain</i>	95		128		33	
163 6		85569	<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhimurium str. DT104	1		1			
163 7		85698	<i>Achromobacter xylosoxidans</i>	1		1			
163 8		85708	Porcine circovirus 2	43	1	43	1		
163 9		85709	Porcine circovirus type 2-B	18		18			
164 0		85777	<i>Agelas mauritiana</i>		1		1		
164 1	X	85831	<i>Bacteroides acidifaciens</i>				2		2
164 2		85929	<i>Sphaerulina musiva</i>		1		1		
164 3		85962	<i>Helicobacter pylori</i> 26695	40	10	40	10		
164 4		85963	<i>Helicobacter pylori</i> J99		53		53		
164 5		85991	<i>Chlamydia pecorum</i>	142		142			
164 6		86049	<i>Cladophialophora carrionii</i>		2		2		
164 7		86107	Coxsackievirus A6	1		1			
164 8		86385	Porcine circovirus type 2-D	1		1			
164 9		86600	<i>Discosoma</i> sp.		1		1		
165 0		86782	Amur virus		35		35		
165 2		87883	<i>Burkholderia multivorans</i>	1	2	1	2		
165 3		88036	<i>Selaginella moellendorffii</i>		1		1		
165 4		88085	<i>Lachesis stenophrys</i>	13		13			
165 5		88086	<i>Protobothrops elegans</i>	7		7			
165 6		88087	<i>Protobothrops flavoviridis</i>	3		3			
165 7		88103	Influenza A virus (A/Hong Kong/532/1997(H5N1))		1		1		
165 8		88104	Influenza A virus (A/Hong Kong/483/1997(H5N1))	3	10	3	10		
165 9	X	88431	<i>Dorea longicatena</i>				1		1
166 0		88776	Influenza A virus (A/Brevig Mission/1/1918(H1N1))	1	1	1	1		
166 1		89059	<i>Ligilactobacillus acidipiscis</i>		1		1		
166 2		89169	Influenza A virus (A/Chicken/Pennsylvania/13552-1/98 (H7N2NSB))	1		1			
166 3		89172	Influenza A virus (A/quail/Arkansas/16309-7/94 (H7N3NSA))	1		1			
166 5		89462	<i>Bubalus bubalis</i>	7	1	7	1		
166 7		90370	<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhi	22	119	22	119		
166 8		90371	<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhimurium	16	76	16	76		
166 9		91465	<i>Pseudomonas</i> sp. MIS38		1		1		
167 0		91626	<i>Mucor ambiguus</i>		1		1		
167 1		92050	<i>Macruronus magellanicus</i>	2		2			
167 2		92637	<i>Metarhizium acridum</i>		1		1		

167 3		92652	Shrimp white spot syndrome virus	3		3			
167 4		93061	Staphylococcus aureus subsp. aureus NCTC 8325	5	1	5	1		
167 5		93062	Staphylococcus aureus subsp. aureus COL	17		17			
167 6		93621	Heteropneustes fossilis		1		1		
167 7		93678	TTV-like mini virus		1		1		
167 8		93838	Influenza A virus (A/goose/Guangdong/1/1996(H5N1))	1	10	1	10		
167 9		94432	Human rotavirus MP409	1	1	1	1		
168 0		94966	Bluetongue virus 12	15		15			
168 1		94967	Bluetongue virus 4	7		7			
168 3		95486	Burkholderia cenocepacia	1	1	1	1		
168 4		95602	Eriocheir sinensis	10		10			
168 5		95888	Influenza A virus (A/mallard duck/PA/10218/84(H5N2))	8		8			
168 7		97138	Clostridium sp. MD294		3		3		
168 8		97253	Eubacterium plexicaudatum		4		4		
168 9		97348	Influenza A virus (A/Chicken/Hong Kong/G23/97(H9N2))		1		1		
169 0		97353	Influenza A virus (A/Chicken/Hong Kong/739/94(H9N2))		1		1		
169 1		97477	Acetomicrobium mobile		1		1		
169 2		98360	Salmonella enterica subsp. enterica serovar Dublin		1		1		
169 3		99287	Salmonella enterica subsp. enterica serovar Typhimurium str. LT2		78		78		
169 4		99586	Echis ocellatus	5		5			
169 5	X	99806	Taxus cuspidata			1		1	
169 6		99875	Leishmania donovani donovani	1		1			
169 7		99883	Tetraodon nigroviridis		2		2		
169 8		100226	Streptomyces coelicolor A3(2)	1	1	1	1		
169 9		100835	Influenza A virus (A/Chicken/Hong Kong/258/97 (H5N1))	1		1			
170 0		101350	Porcine rotavirus strain A253	2		2			
170 1		102617	Helicobacter pylori SS1	4	3	4	3		
170 2	X	102618	Helicobacter pylori NCTC 11637 = CCUG 17874 = ATCC 43504 = JCM 12093			16	2	16	2
170 3		102793	H5N1 subtype	27	40	27	40		
170 4		102796	H9N2 subtype	3	1	3	1		
170 5		102862	Proteus penneri	1		1			
170 6		103448	Pleistophora sp. LS		1		1		
170 7		103903	Coxsackievirus B3 (strain Nancy)	13	32	13	32		
170 8		103905	Coxsackievirus B4 (strain E2)	7	41	7	41		
170 9		103906	Coxsackievirus B4 (strain JVB / Benschoten / New York/51)		76		76		
171 1		103920	Guinea pig cytomegalovirus (strain 22122 / ATCC VR682) (GPCMV)	1		1			

171 2		103922	Human enterovirus 71 (strain 7423/MS/87)	4		4			
171 3		103929	Rabies virus strain Pasteur vaccin	2	1	5	1	3	
171 4		103930	Rhesus cytomegalovirus strain 68-1		18		18		
171 5		104102	Acetobacter tropicalis		1		1		
171 6		104355	Gloeophyllum trabeum		1		1		
171 7		105751	Aeromonas bestiarum		1		1		
171 8		106276	Mimachlamys nobilis	7		7			
171 9		106654	Acinetobacter nosocomialis		1		1		
172 0		106820	Hepatitis B virus subtype adr	23	25	23	25		
172 1		106821	Hepatitis B virus subtype adw	17	76	17	76		
172 2		107404	Influenza B virus (B/Beijing/184/93)		1		1		
172 3		107406	Influenza B virus (B/Chiba/447/98)		1		1		
172 4		107493	Influenza A virus (A/Cordoba/3278/96(H3N2))		1		1		
172 5		107558	Influenza A virus (A/France/75/97(H3N2))		1		1		
172 6		107819	Clostridium perfringens D	45		45			
172 7		108098	Human mastadenovirus B	1		1			
172 8		110195	Foot-and-mouth disease virus Asia 1	10	3	10	3		
172 9		110321	Sinorhizobium medicae		1		1		
173 0		112509	Hordeum vulgare subsp. vulgare		26		26		
173 1		114497	Cordyceps fumosorosea		1		1		
173 2		114727	H1N1 subtype	23	49	23	49		
173 3		114729	H2N2 subtype		1		1		
173 4		114742	Pythium insidiosum	2		2			
173 5		115711	Chlamydia pneumoniae AR39	1		1			
173 6		115713	Chlamydia pneumoniae CWL029	73	48	73	48		
173 7		117125	Cucumber mosaic virus (strain Pepo)	1		1			
173 8		117187	Fusarium verticillioides	1		1			
173 9		117204	African horse sickness virus 3	21		21			
174 0	X	117260	Rosa acicularis			1		1	
174 1		118161	Pleurocapsa sp. PCC 7319		1		1		
174 2		119210	H3N2 subtype	6	23	6	24		1
174 3		119211	H3N8 subtype	1		1			
174 4		119215	H7N3 subtype	2		2			
174 5		119218	H7N7 subtype		1		1		
174 6		119220	H5N2 subtype	1		1			
174 7		119602	Streptococcus dysgalactiae subsp. equisimilis	22		22			
174 8		119856	Francisella tularensis subsp. tularensis		5		5		

174 9	119912	Salmonella enterica subsp. enterica serovar Choleraesuis		2		2		
175 0	120505	Baboon cytomegalovirus		1		1		
175 1	121224	Pediculus humanus corporis	2		2			
175 2	121723	Photobacterium sp. SKA34		1		1		
175 3	121759	Paracoccidioides brasiliensis	4	16	4	16		
175 4	122291	Kappapapillomavirus 1	2		2			
175 5	122355	Pseudomonas psychrophila	1		1			
175 6	122586	Neisseria meningitidis MC58	18		18			
175 7	122928	Norovirus GI	2		2			
175 8	122929	Norovirus GII	2	19	2	19		
175 9	123734	Hypoderma bovis	1		1			
176 1	126793	Plasmodium vivax Sal-1	24		24			
176 2	126794	Vaccinia virus Ankara		127		127		
176 3	127886	Halalkalibacter hemicellulosilyticus		1		1		
176 4	127906	Vibrio cholerae O1	9		9			
176 5	127999	Tanacetum parthenium		1		1		
176 6	128947	Ebola virus - Gabon (1994-1997)		18		18		
176 7	128948	Sudan virus - Boniface, Sudan, 1976	3	1	3	1		
176 8	128949	Sudan ebolavirus - Maleo (1979)		30		30		
176 9	128951	Ebola virus - Zaire (1995)		3		3		
177 0	128952	Ebola virus - Mayinga, Zaire, 1976	70	13	72	18	2	5
177 1	128982	Influenza A virus (A/Swine/Indiana/9K035/99 (H1N2))		2		2		
177 2	128987	Grass carp reovirus		1		1		
177 3	128999	Tai Forest virus - Cote d'Ivoire, Cote d'Ivoire, 1994	1	3	1	3		
177 4	129000	Ebola virus - Eckron (Zaire, 1976)		12		12		
177 5	129003	Reston ebolavirus - Reston	2	27	2	27		
177 6	129052	Catelliglobospora koreensis		1		1		
177 7	129138	Pseudomonas amygdali pv. morsprunorum	2		2			
177 9	129727	Bovine papular stomatitis virus		2		2		
178 0	129788	Ruditapes philippinarum	3		3			
178 1	129875	Human mastadenovirus A		1		1		
178 2	129951	Human mastadenovirus C	1	3	1	3		
178 3	130663	Fowl aviadenovirus 4	3		3			
178 4	130760	Influenza A virus (A/Hong Kong/1073/99(H9N2))	1	25	1	25		
178 5	130763	Influenza A virus (A/Hong Kong/156/97(H5N1))	4	26	4	26		
178 6	131110	Schaalia radingae		1		1		
178 7	132475	Yaba-like disease virus		3		3		

178 8	132487	Measles virus strain Schwarz	1	1	1	1		
178 9	132504	Influenza A virus (A/X-31(H3N2))	78	193	78	472		279
179 1	132783	Influenza A virus (A/Netherlands/626/89 (H3N2))		1		1		
179 2	132841	Influenza A virus (A/Netherlands/785e/90 (H3N2))		1		1		
179 3	132884	Influenza A virus (A/Netherlands/458/98 (H3N2))		1		1		
179 4	133704	Porcine circovirus 1	6		6			
179 5	133926	Olsenella uli		1		1		
179 6	134537	Paraburkholderia fungorum	1		1			
179 8	135661	Influenza A virus (A/environment/Hong Kong/437-10/99 (H5N1))		1		1		
179 9	135720	Neisseria meningitidis serogroup C	3		3			
180 0	136371	Lasallia papulosa	1		1			
180 1	136474	Influenza A virus (A/Swine/Colorado/23619/99(H3N2))		1		1		
180 2	136966	SEN virus		7		7		
180 3	137578	Influenza A virus (A/England/939/69 x A/PR/8/34)		2		2		
180 4	137885	Influenza A virus (A/parakeet/Chiba/1/97(H9N2))		4		4		
180 5	137886	Influenza A virus (A/parakeet/Narita/92A/98(H9N2))		2		2		
180 6	138072	Candidatus Hamiltonella defensa		1		1		
180 7	138948	Enterovirus A	8		8			
180 8	138949	Enterovirus B	7		7			
180 9	138950	Enterovirus C	13	3	13	3		
181 0	138951	Enterovirus D	7		7			
181 2	142948	Influenza A virus (A/ruddy turnstone/Delaware/67/1998(H12N4))		1		1		
181 3	144549	Mycolicibacterium fortuitum subsp. fortuitum		1		1		
181 4	145262	Methanothermobacter thermautotrophicus		1		1		
181 5	145306	Influenza A virus (A/swine/Hong Kong/10/98(H9N2))	5		5			
181 6	145307	Influenza A virus (A/swine/Hong Kong/9/98(H9N2))	2		2			
181 7	147095	Influenza A virus (A/Swine/Italy/25823/94(H3N2))		2		2		
181 8	147272	Paspalum notatum		16		16		
181 9	147711	Rhinovirus A		21		21		
182 0	148360	Watermelon mosaic virus 2 (isolate USA)	2		2			
182 1	148363	Helicoverpa armigera nucleopolyhedrovirus G4	2		2			
182 2	149539	Salmonella enterica subsp. enterica serovar Enteritidis	1	3	1	3		
182 3	150080	Norovirus isolates	1		1			
182 4	150127	Influenza B virus (B/Osaka/983/97-V3)	1		1			

182 5		150340	Vibrio antiquarius		15		15		
182 6		150846	Enterovirus 5865/sin/000009	2		2			
182 7		151250	Kali turgidum	1	22	1	22		
182 8		152219	Menangle virus	2		2			
182 9	X	152682	Sphingomonas melonis				10		10
183 0		152794	Corynebacterium efficiens		5		5		
183 1		153496	Kozakia baliensis		1		1		
183 2		153969	Influenza A virus (A/Hong Kong/497/97(H3N2))		1		1		
183 3		154540	Influenza A virus (A/Kayano/57(H2N2))		1		1		
183 4		155091	Secale cereale subsp. afghanicum		2		2		
183 5		155218	Influenza A virus (A/Hong Kong/482/97(H5N1))	7	1	7	1		
183 6		155220	Influenza A virus (A/Hong Kong/542/97(H5N1))	1		1			
183 7		155222	Influenza A virus (A/Hong Kong/488/97(H5N1))		1		1		
183 8		155225	Influenza A virus (A/Hong Kong/507/97(H5N1))		5		5		
183 9		155226	Influenza A virus (A/Hong Kong/514/97(H5N1))		1		1		
184 0		155227	Influenza A virus (A/Hong Kong/516/97(H5N1))		2		2		
184 1		155864	Escherichia coli O157:H7 str. EDL933	4	32	4	32		
184 2		155917	Influenza A virus (A/NWS/33HA-A/tern/Australia/G70C/75NA(H1N9))	1		1			
184 3		156230	Karenia brevis	1		1			
184 5		157703	Murine polyomavirus strain A3		4		4		
184 6		157914	Ziziphus mauritiana	4		4			
184 7		158306	Influenza A virus (A/swine/Cotes d'Armor/800/00(H1N2))		1		1		
184 8		158465	Human calicivirus Hu/NLV/GII/MD145-12/1987/US	1	1	5	1	4	
184 9		158474	Bovine viral diarrhea virus strain Trangie Y546	1		1			
185 0	X	158836	Enterobacter hormaechei				8		8
185 1		158879	Staphylococcus aureus subsp. aureus N315		5		5		
185 2		158899	Collimonas fungivorans		2		2		
185 3		159091	Pseudomonas sp. KIE171		1		1		
185 4		159449	Embleya scabrispora	1		1			
185 5		159471	Influenza A virus (A/eq/Sussex/89/(H3N8))	4		4			
185 6		159479	Saaremaa hantavirus		21		21		
185 7		160490	Streptococcus pyogenes M1 GAS	13	2	13	2		
185 8		160491	Streptococcus pyogenes str. Manfredo	3	35	3	35		
185 9		160691	Tiger frog virus		1		1		
186 0		160753	Simian immunodeficiency virus 17E-Fr		2		2		

186 1	X	160808	Acidithiobacillus ferrivorans				1		1
186 2		161502	Influenza A virus (A/swine/Italy/1509-6/97(H1N1))		15		15		
186 3		161600	Yellow grouper nervous necrosis virus	1		1			
186 4		161727	Foot-and-mouth disease virus A10/Holland	11	14	11	14		
186 5		161934	Beta vulgaris	5		5			
186 6		162145	Human metapneumovirus	8	130	8	130		
186 7		162326	Influenza A virus (A/Hong Kong/1143/99(H3N2))	1		1			
186 8		162425	Aspergillus nidulans		1		1		
186 9		163164	Wolbachia endosymbiont of Drosophila melanogaster		1		1		
187 0		164326	Influenza A virus (A/Hong Kong/1131/1998(H1N1))	1	1	1	1		
187 1		164756	Mycobacterium sp. MCS		73		73		
187 2		164757	Mycobacterium sp. JLS		77		77		
187 3		165179	Segatella copri		5		5		
187 4		165420	Influenza A virus (A/Turkey/MO/24093/99(H1N2))		4		4		
187 5		165512	Influenza A virus (A/goose/Guangdong/3/1997(H5N1))	3	1	3	1		
187 8		167758	TTV-like virus DXL1		10		10		
187 9		168278	Influenza A virus (A/swine/Finistere/127/99(H3N2))		4		4		
188 0		168807	Escherichia coli O127:H6	1		1			
188 1		169166	Influenza A virus (A/swine/Italy/1521/98(H1N2))		4		4		
188 2		169169	Influenza A virus (A/swine/Cotes d'Armor/1482/1999(H1N1))		1		1		
188 3		169173	Choclo virus		3		3		
188 5		169963	Listeria monocytogenes EGD-e	5	64	5	64		
188 7		170187	Streptococcus pneumoniae TIGR4		2		2		
188 8		170325	Canid alphaherpesvirus 1		1		1		
188 9		170342	Influenza A virus (A/swine/Italy/1654-1/99(H1N2))		1		1		
189 0		170500	H1N9 subtype	1		1			
189 1		170525	Measles virus genotype D4	1		1			
189 2		170529	Measles virus genotype D7	1		1			
189 3		170955	Amur virus Solovey/AP63/1999		2		2		
189 4		171101	Streptococcus pneumoniae R6	2	12	2	12		
189 5		171264	Measles virus genotype D3	1		1			
189 6		171370	human papillomavirus 86	1		1			
189 7		171425	Influenza B virus (B/Kobe/87/2001)	1		1			
189 8		171631	Fusarium oxysporum species complex	8		8			
189 9		171929	Anacardium occidentale	65	10	65	10		
190 0		172148	Alkhumra hemorrhagic fever virus		4		4		

190 1	172149	Influenza A virus (A/swine/Quebec/192/81(H1N1))	1		1			
190 2	173473	Influenza A virus (A/swine/Hong Kong/2106/98(H9N2))		2		2		
190 3	173477	Influenza A virus (A/swine/Hong Kong/3297/98(H9N2))		3		3		
190 5	176275	Beauveria bassiana		1		1		
190 6	176279	Staphylococcus epidermidis RP62A		1		1		
190 7	176672	Influenza A virus (A/Duck/Hong Kong/ww461/2000(H5N1))		1		1		
190 9	177128	Influenza A virus (A/SW/MN/12883/00(H1N2))		2		2		
191 0	177131	Influenza A virus (A/SW/KS/13481-T/00(H1N2))		1		1		
191 1	177188	Influenza A virus (A/SW/MN/3227/00(H1N2))		1		1		
191 2	177416	Francisella tularensis subsp. tularensis SCHU S4	44	92	44	92		
191 3	178773	Norwalk-like virus sp.		1		1		
191 4	178876	Cryptococcus neoformans var. grubii	1		1			
191 5	180454	Anopheles gambiae str. PEST	6	2	6	2		
191 6	180498	Jatropha curcas	6		6			
191 7	182082	Chlamydia pneumoniae TW-183	7		7			
191 8	182781	Influenza A virus (A/Goose/Hong Kong/385.3/2000(H5N1))		1		1		
191 9	182785	Mycobacterium tuberculosis subsp. tuberculosis		1		1		
192 0	183741	Influenza A virus (A/Chicken/Shanghai/F/98(H9N2))	5		5			
192 1	183764	Influenza A virus (A/Puerto Rico/8/34/Mount Sinai(H1N1))	7	100	7	100		
192 2	183778	Influenza A virus (A/quail/Hong Kong/SF550/00(H6N1))		2		2		
192 3	183783	Influenza A virus (A/duck/Hong Kong/3096/99(H6N2))		2		2		
192 4	184816	Influenza B virus (B/Kadoma/122/99)	2		2			
192 5	184922	Giardia lamblia ATCC 50803		249		249		
192 6	185431	Trypanosoma brucei brucei TREU927		1		1		
192 7	185579	Hepatitis E virus type 1	3		3			
192 8	185580	Hepatitis E virus type 4	1		1			
192 9	185794	Influenza A virus (A/mallard/NZ/1/97(H5N2))		1		1		
193 0	185894	rhinovirus A15	3		3			
193 1	185905	rhinovirus A34		54		54		
193 2	185907	rhinovirus A39		1		1		
193 3	185949	Sphingomonas aurantiaca		1		1		
193 5	186460	Influenza A virus (A/swine/New Jersey/11/1976(H1N1))	1		1			
193 6	186538	Zaire ebolavirus	188	901	188	904		3
193 7	186539	Reston ebolavirus	1	11	1	11		
193 8	186540	Sudan ebolavirus	8	292	8	292		

1939	186541	Tai Forest ebolavirus		1		1		
1940	187164	Influenza A virus (A/Swine/Minnesota/55551/2000(H1N2))		1		1		
1941	187410	Yersinia pestis KIM10+		11		11		
1942	187420	Methanothermobacter thermautotrophicus str. Delta H		1		1		
1943	188538	Human parainfluenza virus 1 strain Washington/1964		11		11		
1944	188763	Panine betaherpesvirus 2		4		4		
1945	189518	Leptospira interrogans serovar Lai str. 56601	1		1			
1946	189918	Mycobacterium sp. KMS		69		69		
1947	190063	Fowl avianinfluenza virus C	1		1			
1948	191090	Influenza A virus (A/Weiss/1943(H1N1))		1		1		
1949	191218	Bacillus anthracis str. A2012		29		29		
1950	192087	Pseudomonas syringae pv. atrofaciens	4		4			
1951	192222	Campylobacter jejuni subsp. jejuni NCTC 11168 = ATCC 700819	13	22	17	22	4	
1952	192535	Influenza A virus (A/Saudi Arabia/7971/2000(H1N1))		1		1		
1953	192549	Influenza A virus (A/Egypt/96/2002(H1N2))		1		1		
1954	192720	Influenza A virus (A/Saudi Arabia/2581/2001(H1N2))		1		1		
1956	194601	Vipera aspis aspis	3		3			
1957	194958	Porcine endogenous retrovirus A	2		2			
1958	194959	Porcine endogenous retrovirus B	1		1			
1959	195055	Human parechovirus 3	4		4			
1960	195099	Campylobacter jejuni RM1221		13		13		
1961	195102	Clostridium perfringens str. 13		1		1		
1962	195700	Avian rotavirus PO-13	6		6			
1963	196164	Corynebacterium efficiens YS-314		4		4		
1964	196403	Baboon endogenous virus	1	1	1	1		
1965	196462	Influenza A virus (A/chicken/California/1002a/00(H6N2))	1	1	1	1		
1966	196600	Vibrio vulnificus YJ016		75		75		
1967	196620	Staphylococcus aureus subsp. aureus MW2		7		7		
1968	196627	Corynebacterium glutamicum ATCC 13032		14		14		
1969	197575	Haemophilus aegyptius	1		1			
1970	197579	Influenza A virus (A/Chicken/Guangdong/11/97(H9N2))		1		1		
1971	197585	Influenza A virus (A/chicken/Shandong/6/96(H9N2))	1		1			
1972	197589	Influenza A virus (A/Quail/Shanghai/8/96(H9N2))		1		1		
1973	197780	Bluetongue virus 8	2	21	2	21		

197 4	198059	Influenza A virus (A/England/627/01(H1N2))		2		2		
197 5	198094	Bacillus anthracis str. Ames		2		2		
197 6	198214	Shigella flexneri 2a str. 301		4		4		
197 7	198215	Shigella flexneri 2a str. 2457T		2		2		
197 8	199306	Coccidioides posadasii		17		17		
197 9	199310	Escherichia coli CFT073	1	2	1	2		
198 0	199738	uncultured Chlamydia sp.	2		2			
198 1	200454	Pseudomonas tremae		1		1		
198 2	201444	Aracatuba virus		5		5		
198 3	202812	Adeno-associated virus - 7		2		2		
198 4	202813	Adeno-associated virus - 8	17	14	17	17		3
198 5	202950	Acinetobacter baylyi		2		2		
198 6	203119	Acetivibrio thermocellus ATCC 27405		1		1		
198 7	203124	Trichodesmium erythraeum IMS101		2		2		
198 8	203126	Influenza A virus (A/Kamata/14/91(H3N2))	1		1			
198 9	203129	Influenza A virus (A/Aichi/2/1994(H3N2))		1		1		
199 0	203172	Camelpox virus CMS		28		28		
199 1	203173	Camelpox virus M-96		133		133		
199 2	203174	Camelpox virus CP1		1		1		
199 4	204038	Dickeya dadantii	2		2			
199 5	204428	Chlamydiota	16		16			
199 6	204711	Theilovirus		10		10		
199 7	204722	Brucella suis 1330	1	29	1	29		
199 9	205913	Bifidobacterium longum DJO10A		2		2		
200 0	205914	Histophilus somni 129PT		1		1		
200 1	205920	Ehrlichia chaffeensis str. Arkansas	23		23			
200 2	205946	Influenza A virus (A/Chicken/Beijing/1/95(H9N2))		1		1		
200 4	206203	Influenza B virus (B/Hong Kong/330/2001)		23		23		
200 5	207401	Influenza A virus (A/finch/Canada/NS1301/2001(H3N8))		1		1		
200 6	208226	Alkaliphilus metalliredigens		1		1		
200 7	208435	Streptococcus agalactiae 2603V/R	1	2	1	2		
200 8	208726	Human hepatitis A virus	4	1	4	1		
200 9	208893	Human respiratory syncytial virus A	31	4	31	4		
201 1	208963	Pseudomonas aeruginosa UCBPP-PA14		23		23		
201 2	208964	Pseudomonas aeruginosa PAO1	2	38	4	38	2	
201 3	209261	Salmonella enterica subsp. enterica serovar Typhi str. Ty2		16		16		

2015		209841	Actinobacillus pleuropneumoniae serovar 7	1		1			
2016		209882	Fusobacterium vincentii ATCC 49256		2		2		
2017		210655	Influenza A virus (A/Duck/Shantou/2144/00(H9N2))		1		1		
2018		210656	Influenza A virus (A/Duck/Shantou/1881/00(H9N2))		1		1		
2019		210659	Influenza A virus (A/Duck/Shantou/2088/01(H9N2))		1		1		
2020		210663	Influenza A virus (A/Duck/Shantou/2134/00(H9N2))		1		1		
2021		211044	Influenza A virus (A/Puerto Rico/8/1934(H1N1))	37	551	39	551	2	
2022		211110	Streptococcus agalactiae NEM316		1		1		
2023		211882	Leptospira interrogans serovar Australis	2		2			
2024		212042	Anaplasma phagocytophilum str. HZ	21		21			
2025		212045	Bacillus anthracis str. Western North America USA6153		1		1		
2026		212361	Anabaena aequalis	1		1			
2027		212717	Clostridium tetani E88		16		16		
2028		214092	Yersinia pestis CO92	53	22	53	22		
2030		214697	Musa acuminata AAA Group	1		1			
2032		214856	Alistipes finegoldii		1		1		
2033		215243	Exophiala oligosperma		1		1		
2034		215358	Larimichthys crocea	3		3			
2036		215862	Influenza A virus (A/Chicken/Nanchang/2-220/2001(H3N6))		1		1		
2038		216466	Streptococcus agalactiae serogroup V	1		1			
2039		216495	Streptococcus agalactiae serogroup III	14		14			
2040		216592	Escherichia coli O42	2		2			
2041		216594	Mycobacterium marinum M		29		29		
2042		216597	Salmonella enterica subsp. enterica serovar Typhimurium str. SL1344		2		2		
2043		216600	Streptococcus pneumoniae 23F	7		7			
2044		216877	Influenza A virus (A/quail/Hong Kong/FY298/00(H9N2))		4		4		
2045		216895	Vibrio vulnificus CMCP6		53		53		
2046		217686	Little cherry virus 1		1		1		
2047		217992	Escherichia coli O6		7		7		
2048		218497	Chlamydia abortus S26/3	28		28			
2049	X	218538	Dialister invisus				1		1
2050		220341	Salmonella enterica subsp. enterica serovar Typhi str. CT18		92		92		
2051		220837	Cryptosporidium sp. MNJ-1		1		1		
2052		220977	Influenza A virus (A/England/10/67(H2N2))		1		1		
2053		221012	Influenza A virus (A/Rio/6/69(H3N2))		1		1		

205 4		221703	Simian foamy virus Pongo pygmaeus pygmaeus	1		1			
205 5		221918	Bovine viral diarrhea virus VEDEVAC	6		6			
205 6	X	222929	Coccidioides posadasii C735 delta SOWgp				10		10
205 7		223337	Tobacco leaf curl Zimbabwe virus		1		1		
205 8		223926	Vibrio parahaemolyticus RIMD 2210633	2	17	2	17		
205 9		223931	Influenza A virus (A/Bangkok/10/1983(H1N1))		1		1		
206 1		223935	Influenza A virus (A/Okuda/1957(H2N2))	1	9	1	9		
206 2		223947	Influenza A virus (A/Hokkaido/20/89(H3N2))		3		3		
206 3		223997	Murine norovirus 1	8	1	8	1		
206 4		224197	Influenza C virus (C/Nara/1/85)		1		1		
206 5		224206	Simian immunodeficiency virus - mon		1		1		
206 6		224308	Bacillus subtilis subsp. subtilis str. 168		1		1		
206 7		224324	Aquifex aeolicus VF5	2	1	2	1		
206 8		224326	Borrelia burgdorferi B31	113	19	142	19	29	
206 9		224819	Influenza A virus (A/Nagasaki/76/98(H3N2))		1		1		
207 0		224914	Brucella melitensis bv. 1 str. 16M	6	14	6	14		
207 1		224964	Influenza B virus (B/Johannesburg/5/99)	1		1			
207 2		225071	Influenza A virus (A/Tokyo/1566/98(H3N2))		1		1		
207 3		225085	Influenza A virus (A/Aichi/2/68 (Ao))		9		9		
207 4		225088	Influenza A virus (A/Kitakyushu/159/93(H3N2))		1		1		
207 5		225089	Influenza A virus (A/equine/NewMarket/D64/79(H3N8))		1		1		
207 6		226186	Bacteroides thetaiotaomicron VPI-5482		1		1		
207 7		226900	Bacillus cereus ATCC 14579	1	1	1	1		
207 8		227377	Coxiella burnetii RSA 493		13		13		
207 9		227941	Chlamydia caviae GPIC	11		11			
208 0		227984	SARS coronavirus Tor2	2723	2205	2725	2205	2	
208 1		228330	SARS coronavirus Urbani	28	67	29	67	1	
208 2		228399	Actinobacillus pleuropneumoniae serovar 1 str. 4074	2	1	2	1		
208 3		228400	Histophilus somni 2336	1		1			
208 4		228407	SARS coronavirus BJ01	67	28	67	28		
208 5		228415	SARS coronavirus CUHK-W1	1		1			
208 6		228928	Influenza A virus (A/Memphis/31/98(H3N2))	6		6			
208 7		229032	Porcine epidemic diarrhea virus CV777	10		89		79	
208 8		229051	Influenza A virus (A/81/HO)		6		6		
208 9		229992	SARS coronavirus Frankfurt 1	5	27	5	27		

2090		231428	Avian infectious bronchitis virus (strain Vic S)	11	8	11	8		
2091		231455	Dyella japonica	1		1			
2092		231851	Influenza A virus (A/Hong Kong/1774/99(H3N2))		1		1		
2093		233262	Bovine enteric coronavirus (strain 98TXSF-110-ENT)		1		1		
2094		233412	[Haemophilus] ducreyi 35000HP	1		1			
2095		233413	Mycobacterium tuberculosis variant bovis AF2122/97		54		54		
2096		234267	Candidatus Solibacter usitatus Ellin6076		1		1		
2097		234603	Sapovirus Mc114		1		1		
2098		234826	Anaplasma marginale str. St. Maries	16	28	16	28		
2099		235279	Helicobacter hepaticus ATCC 51449	2		2			
2100	X	235443	Cryptococcus neoformans var. grubii H99				60		60
2101		235455	Adeno-associated virus 9	10		15	15	5	15
2103		237561	Candida albicans SC5314		3		3		
2104		237609	Pseudomonas alkylphenolica		1		1		
2105		237631	Ustilago maydis 521		2		2		
2106		237895	Cryptosporidium hominis		3		3		
2108		239935	Akkermansia muciniphila		9		9		
2109		240426	Squirrelpox virus		1		1		
2111		242619	Porphyromonas gingivalis W83	3		3			
2113		243160	Burkholderia mallei ATCC 23344		40		40		
2114		243161	Chlamydia muridarum str. Nigg	11	72	11	72		
2115		243164	Dehalococcoides mccartyi 195		1		1		
2116		243230	Deinococcus radiodurans R1 = ATCC 13939 = DSM 20539	2		2			
2117		243232	Methanocaldococcus jannaschii DSM 2661	3	1	3	1		
2118		243243	Mycobacterium avium 104		42		42		
2119		243273	Mycoplasma genitalium G37	8	1	8	1		
2120		243274	Thermotoga maritima MSB8	1		1			
2121		243276	Treponema pallidum subsp. pallidum str. Nichols	283	9	287	9	4	
2122		243277	Vibrio cholerae O1 biovar El Tor str. N16961	4	3	4	3		
2123		243678	Influenza A virus (A/chicken/Jiangsu/JS-1/2002(H9N2))	1	1	1	1		
2124		244320	Escherichia coli O55:H7		1		1		
2125		244367	Foot-and-mouth disease virus C-S8c1	15	18	15	18		
2126		246196	Mycobacterium smegmatis MC2 155		61		61		
2127		246202	Streptococcus sobrinus 6715	4	4	4	4		
2128		246437	Tupaia chinensis	1	1	2	2	1	1
2129		246618	Bifidobacterium thermacidophilum	1		1			

213 0	X	246787	<i>Bacteroides cellulosilyticus</i>				1		1
213 1		246878	Canine parvovirus 2	3		3			
213 2		249372	Influenza A virus (A/duck/Hokkaido/13/00(H9N2))	2		2			
213 3		251297	Influenza A virus (A/Netherlands/127/03(H7N7))		1		1		
213 4		253182	Human Respiratory syncytial virus 9320	1		1			
213 5		253446	Avian orthoreovirus strain 1017-1	1		1			
213 6		253682	Influenza A virus (A/swine/Hong Kong/5190/99(H3N2))		3		3		
213 7		253686	Influenza A virus (A/swine/Hong Kong/74/02(H3N2))		1		1		
213 8		254252	Lactococcus virus P2	2		2			
213 9		254355	Small ruminant lentivirus	1		1			
214 1		255681	Influenza A virus (A/Yokohama/22/2002(H1N2))		2		2		
214 2		255682	Influenza A virus (A/Yokohama/47/2002(H1N2))		1		1		
214 3		256045	Influenza A virus (A/swine/Chiai/77-10/2001(H3N1))		1		1		
214 4		256080	Influenza B virus (B/Kobe/1/2003)	1		1			
214 5	X	256839	<i>Shewanella decolorationis</i>				6		6
214 6		257309	<i>Corynebacterium diphtheriae</i> NCTC 13129	118		118			
214 7		257313	<i>Bordetella pertussis</i> Tohama I		15		15		
214 8		257363	<i>Rickettsia typhi</i> str. Wilmington		6		6		
214 9		260799	<i>Bacillus anthracis</i> str. Sterne	5	4	5	4		
215 0		260806	Influenza A virus (A/FPV/Dutch/1927(H7N7))	1		1			
215 1		260965	Cercopithecine herpesvirus 1 (strain E2490)	18		18			
215 2		261202	Alto Paraguay hantavirus		3		3		
215 3		261204	Itapua hantavirus		10		10		
215 4		261594	<i>Bacillus anthracis</i> str. 'Ames Ancestor'		1		1		
215 5		261658	<i>Cavenderia fasciculata</i>		1		1		
215 6		262307	Measles virus genotype A	4		4			
215 7		262316	<i>Mycobacterium avium</i> subsp. <i>paratuberculosis</i> K-10	4	58	4	58		
215 8		262698	<i>Brucella abortus</i> bv. 1 str. 9-941		1		1		
215 9		262722	<i>Mesomycoplasma hyopneumoniae</i> 7448	1		1			
216 0		262724	<i>Thermus thermophilus</i> HB27	1		1			
216 1		262727	<i>Haemophilus influenzae</i> R2846		6		6		
216 2		262728	<i>Haemophilus influenzae</i> R2866		2		2		
216 3		263683	Bovine herpesvirus 5 strain TX89	2		2			
216 4		263815	<i>Pneumocystis murina</i>	3		3			
216 5		264202	<i>Chlamydia felis</i> Fe/C-56	14		14			

216 6	264510	Influenza A virus (A/chicken/Thailand/LV1NF/2004(H5N1))		1		1		
216 8	264635	Acholeplasma granularum	1		1			
216 9	264732	Moorella thermoacetica ATCC 39073		1		1		
217 0	265120	Influenza A virus (A/turkey/Italy/214845/2002(H7N3))	1		1			
217 1	265619	Ornithodoros erraticus	22		22			
217 2	265669	Listeria monocytogenes serotype 4b str. F2365		7		7		
217 3	265872	Cowpox virus (Brighton Red)		1		1		
217 4	266093	Influenza A virus (A/chicken/Yamaguchi/7/2004(H5N1))	4		4			
217 5	266264	Cupriavidus metallidurans CH34		3		3		
217 6	266779	Chelativorans sp. BNC1		4		4		
217 7	266827	Influenza A virus (A/Thailand/1(KAN-1)/2004(H5N1))	4	16	4	16		
217 8	266834	Sinorhizobium meliloti 1021		3		3		
217 9	266940	Kineococcus radiotolerans SRS30216 = ATCC BAA-149		2		2		
218 0	267212	Neisseria bacilliformis	1		1			
218 1	267377	Methanococcus maripaludis S2	1		1			
218 2	267409	Listeria monocytogenes serotype 1/2a str. F6854		3		3		
218 3	267410	Listeria monocytogenes serotype 4b str. H7858		24		24		
218 4	267671	Leptospira interrogans serovar Copenhageni str. Fiocruz L1-130	11		11			
218 5	269482	Burkholderia vietnamiensis G4		9		9		
218 6	269484	Ehrlichia canis str. Jake	16		16			
218 7	269638	Bovine enterovirus type 2	2		2			
218 8	269798	Cytophaga hutchinsonii ATCC 33406		1		1		
219 0	269801	Bacillus cereus G9241		39		39		
219 1	270338	Human poliovirus 3 strain Sabin	80	1	80	1		
219 2	270473	Pneumonia virus of mice J3666		10		10		
219 3	270478	Zantedeschia mild mosaic virus	1		1			
219 4	270484	Influenza A virus (A/tiger/Thailand/CU-LV/2004(H5N1))		2		2		
219 5	270490	Influenza A virus (A/chicken/Nakorn-Patom/Thailand/CU-K3/2004(H5N1))		1		1		
219 6	270510	Influenza A virus (A/chicken/Vietnam/HD1/2004(H5N1))		3		3		
219 7	270945	Influenza A virus (A/Thailand/4(SP-528)/2004(H5N1))		21		21		
219 8	271108	Bombyx mori nucleopolyhedrovirus	2		2			
219 9	272560	Burkholderia pseudomallei K96243	3	735	3	735		
220 0	272561	Chlamydia trachomatis D/UW-3/CX	51	26	51	26		
220 1	272562	Clostridium acetobutylicum ATCC 824		1		1		

220 2		272563	Clostridioides difficile 630	1		1			
220 3		272564	Desulfitobacterium hafniense DCB-2		3		3		
220 4		272569	Haloarcula marismortui ATCC 43049	1		1			
220 5		272620	Klebsiella pneumoniae subsp. pneumoniae MGH 78578	28		28			
220 7		272624	Legionella pneumophila subsp. pneumophila str. Philadelphia 1	1		1			
220 8		272627	Magnetospirillum magnetotacticum MS-1		5		5		
220 9		272631	Mycobacterium leprae TN		30		30		
221 0		272634	Mycoplasma pneumoniae M129	9		9			
221 1		272636	Adeno-associated virus		18		18		
221 2		272831	Neisseria meningitidis FAM18	1		1			
221 3		272843	Pasteurella multocida subsp. multocida str. Pm70		5		5		
221 4		272943	Cereibacter sphaeroides 2.4.1		1		1		
221 5		272944	Rickettsia conorii str. Malish 7		5		5		
221 6		272947	Rickettsia prowazekii str. Madrid E		8		8		
221 7		272951	Rickettsia sibirica 246		25		25		
221 8		272989	Salmonella enterica subsp. enterica serovar Enteritidis str. LK5		1		1		
221 9		273123	Yersinia pseudotuberculosis IP 32953		1		1		
222 1		273371	Candida orthopsilosis		1		1		
222 2		274828	Influenza A virus (A/duck/Fujian/13/2002(H5N1))		1		1		
222 3		274829	Influenza A virus (A/duck/Fujian/17/2001(H5N1))		2		2		
222 4		277944	Human coronavirus NL63	5623	8	5629	17	6	9
222 5		278137	Mycolicibacterium gilvum Spyr1		53		53		
222 6		279150	Classical swine fever virus 96TD	7		7			
222 7		279329	Influenza B virus (B/Shandong/7/97)	1	1	1	1		
222 8		279797	Influenza A virus (A/egret/Hong Kong/757.3/2002(H5N1))		3		3		
223 0		279889	Triticum spelta var. arduini		3		3		
223 1		280240	Nocardiosis baichengensis	1		1			
223 2		280463	Emiliania huxleyi CCMP1516		1		1		
223 3	X	280504	Leptospira borgpetersenii serovar Javanica			1		1	
223 4		280855	Seoul virus BjHD01	2		2			
223 5		281310	Haemophilus influenzae 86-028NP	13		13			
223 6		281689	Desulfuromonas acetoxidans DSM 684		1		1		
223 7		282124	Influenza A virus (A/mallard/Alberta/127/00(H3N8))		1		1		
223 8		282458	Staphylococcus aureus subsp. aureus MRSA252	86		86			
223 9		283166	Bartonella henselae str. Houston-1		1		1		
224 0		283643	Cryptococcus neoformans var. neoformans B-3501A	1	3	1	3		

224 1	283877	Leucobacter chromiireducens		1		1		
224 2	284168	Influenza A virus (A/Ck/HK/2133.1/2003(H5N1))		1		1		
224 3	284177	Influenza A virus (A/Ck/HK/YU22/2002(H5N1))	1		1			
224 4	284190	Influenza A virus (A/Ck/Viet Nam/36/2004(H5N1))	1		1			
224 5	284195	Influenza A virus (A/Ck/YN/115/2004(H5N1))		1		1		
224 6	284202	Influenza A virus (A/Dk/ST/5048/2001(H3N8))	1		1			
224 7	284218	Influenza A virus (A/Viet Nam/1203/2004(H5N1))	76	134	76	134		
224 8	284581	Priestia koreensis		1		1		
224 9	284672	SARS coronavirus TJF		14		14		
225 0	284813	Encephalitozoon cuniculi GB-M1		28		28		
225 1	285006	Saccharomyces cerevisiae RM11-1a		2		2		
225 2	286239	Influenza A virus (A/red knot/Delaware/2561/1987(H10N5))		1		1		
225 3	286542	Soochong virus-2		1		1		
225 4	286636	Streptococcus pyogenes MGAS10394		1		1		
225 5	286783	Salmonella enterica subsp. enterica serovar Indiana		1		1		
225 6	288118	Mandrillus leucophaeus foamy virus	1		1			
225 7	289338	Influenza B virus (B/Nanchang/12/98)		1		1		
225 8	289365	Human parvovirus 4		3		3		
225 9	290028	Human coronavirus HKU1	4851	6	4859	17	8	11
226 1	290512	Prosthecochloris aestuarii DSM 271		1		1		
226 3	291592	Influenza A virus (A/crow/Kyoto/53/2004(H5N1))	1		1			
226 4	292039	Influenza A virus (A/chicken/HongKong/SF1/03(H9N2))	1		1			
226 5	292045	Influenza A virus (A/chicken/HongKong/YU427/03(H9N2))	1		1			
226 6	292348	Canine calicivirus (strain 48)	2		2			
226 7	292349	Feline calicivirus (strain Urbana)	3		3			
226 8	292590	Influenza A virus (A/swine/Ontario/42729A/01(H3N3))	1		1			
226 9	292633	Dragon grouper nervous necrosis virus	20		20			
227 0	293054	Influenza A virus (A/turkey/Ontario/6213/66(H5N?))	1		1			
227 1	293055	Influenza A virus (A/mallard/Ohio/556/1987(H5N9))		1		1		
227 2	293090	Influenza A virus (A/Fujian/411/2002(H3N2))		1		1		
227 3	293614	Rickettsia akari str. Hartford		1		1		
227 4	294381	Entamoeba histolytica HM-1:IMSS		103		103		
227 5	295027	Human herpesvirus 5 strain Merlin	4	3	4	7		4
227 6	295319	Salmonella enterica subsp. enterica serovar Paratyphi A str. ATCC 9150		3		3		

227 7	295358	Mesomycoplasma hyopneumoniae 232	21		21			
227 8	296587	Micromonas commoda		1		1		
227 9	298339	Pan troglodytes foamy virus	1		1			
228 0	298653	Frankia sp. EAN1pec		2		2		
228 1	298713	Influenza A virus (A/chicken/Korea/SNU0091/00(H9N2))		1		1		
228 2	298716	Influenza A virus (A/chicken/Korea/SNU1035C/00(H9N2))		1		1		
228 3	299386	Equine arteritis virus Bucyrus	2		9		7	
228 4	299390	Influenza A virus (A/Viet Nam/HN/2004(H5N1))		6		6		
228 5	299730	Influenza A virus (A/chicken/Viet Nam/AG-010/2004(H5N1))		1		1		
228 7	300015	Lactate dehydrogenase elevating virus C	1		1			
228 8	300016	Lactate dehydrogenase elevating virus Plagemann	1		1			
228 9	300180	Mopeia Lassa virus reassortant 29		4		4		
229 0	300267	Shigella dysenteriae Sd197		5		5		
229 1	300268	Shigella boydii Sb227		3		3		
229 2	300269	Shigella sonnei Ss046		9		9		
229 3	300559	PRRSV VR2332	21	7	21	7		
229 4	300563	PRRSV HB-1(sh)/2002	1		1			
229 5	300748	Influenza A virus (A/swine/Korea/S190/2004(H9N2))		3		3		
229 6	300749	Influenza A virus (A/swine/Korea/S452/2004(H9N2))		1		1		
229 7	300750	Influenza A virus (A/chicken/Korea/S1/2003(H9N2))		1		1		
229 8	300852	Thermus thermophilus HB8		1		1		
229 9	301448	Streptococcus pyogenes serotype M3	1		1			
230 0	301449	Streptococcus pyogenes serotype M5	56	57	56	57		
230 1	301450	Streptococcus pyogenes serotype M6	9	7	9	7		
230 2	301451	Streptococcus pyogenes serotype M18	1		1			
230 3	301452	Streptococcus pyogenes serotype M49	1		1			
230 4	301478	Murray valley encephalitis virus (strain MVE-1-51)	11	14	11	14		
230 5	301964	CY1014 virus		1		1		
230 6	302272	Sendai virus (strain Ohita)	1		1			
230 7	303316	Influenza A virus (A/Hatay/2004/(H5N1))	3		3			
230 8	304883	Influenza A virus (A/Belfast/53582/2004(H3N?))		2		2		
230 9	304895	Catenulispora acidiphila		1		1		
231 0	305674	Deerpox virus W-848-83		2		2		
231 1	306061	Influenza A virus (A/swine/IDT/Bakum1832/2000(H1N2))		1		1		

231 2	306254	Campylobacter coli RM2228		2		2		
231 3	306263	Campylobacter lari RM2100		1		1		
231 4	306264	Campylobacter upsaliensis RM3195		1		1		
231 5	307044	West Nile virus strain 385-99	22	12	22	12		
231 6	307685	Merluccius australis polylepis	1		1			
231 7	307819	Influenza A virus (A/Jiangsu/38/2004(H3N2))		1		1		
231 8	308061	Hantavirus HPR/02-73		1		1		
231 9	308159	Araucaria virus		6		6		
232 0	310542	Chimpanzee adenovirus	2		2			
232 1	311169	Influenza A virus (A/duck/Korea/S10/03(H3N2))		1		1		
232 2	311339	Human herpesvirus 5 strain Toledo	9		9			
232 3	311410	Roseibium album		1		1		
232 4	311596	Influenza A virus (A/New York/18/2003(H3N2))		9		9		
232 5	311695	Influenza A virus (A/New York/124/2001(H3N2))		1		1		
232 7	311775	Influenza A virus (A/New York/205/2001(H1N1))		12		12		
232 8	313589	Janibacter sp. HTCC2649		1		1		
232 9	313596	Robiginitalea biformata HTCC2501		1		1		
233 0	313606	Microscilla marina ATCC 23134		1		1		
233 1	313627	Bacillus sp. NRRL B-14911		2		2		
233 2	314262	Roseobacter sp. MED193		1		1		
233 3	314275	Alteromonas mediterranea		1		1		
233 4	314278	Nitrococcus mobilis Nb-231		1		1		
233 5	314283	Reinekea blandensis MED297		1		1		
233 6	314285	Congregibacter litoralis KT71		1		1		
233 7	314288	Vibrio alginolyticus 12G01		3		3		
233 8	314291	Vibrio splendidus 12B01		2		2		
233 9	314292	Photobacterium angustum S14		1		1		
234 0	315277	Chlamydia trachomatis A/HAR-13	6	1	6	1		
234 1	315456	Rickettsia felis URRWXCal2		7		7		
234 2	315730	Bacillus mycoides KBAB4		2		2		
234 3	315749	Bacillus cytotoxicus NVH 391-98		1		1		
234 4	316056	Rhodopseudomonas palustris BisB18		1		1		
234 5	316068	Leadbetterella byssohila		1		1		
234 6	316287	Ceratogyrus marshalli	1		1			
234 7	316385	Escherichia coli str. K-12 substr. DH10B		3		3		
234 8	316401	Escherichia coli ETEC H10407	63	31	63	31		
234 9	316407	Escherichia coli str. K-12 substr. W3110		1		1		

235 0		317652	Influenza A virus (A/Hong Kong/2/68(H3N2))		4		4		
235 1		317655	Sphingopyxis alaskensis RB2256		1		1		
235 2		318586	Paracoccus denitrificans PD1222		2		2		
235 3		319224	Shewanella putrefaciens CN-32		1		1		
235 4		320306	Influenza A virus (A/crested eagle/Belgium/01/2004(H5N1))	2		2			
235 5		320372	Burkholderia pseudomallei 1710b		1		1		
235 6		320373	Burkholderia pseudomallei 668		9		9		
235 7		320374	Burkholderia pseudomallei S13		2		2		
235 8		320388	Burkholderia mallei SAVP1		29		29		
235 9		320390	Burkholderia mallei GB8 horse 4		1		1		
236 0		320483	Anaplasma marginale str. Florida	19	61	19	61		
236 1		320667	Trichosanthes dioica	1		1			
236 2		321314	Salmonella enterica subsp. enterica serovar Choleraesuis str. SC-B67		5		5		
236 3		322053	Konjac mosaic virus	1		1			
236 4		322564	Influenza A virus (A/tiger/Thailand/CU-T4/04(H5N1))		5		5		
236 5		322566	Influenza A virus (A/tiger/Thailand/CU-T6/04(H5N1))		1		1		
236 6		322710	Azotobacter vinelandii DJ		2		2		
236 8		325240	Shewanella baltica OS155		2		2		
236 9		327105	HIV-1 O_ANT70	1		1			
237 0		327205	Influenza A virus (A/New York/348/2003(H1N1))		150		150		
237 1	X	327255	Influenza A virus (A/Canada/720/2005(H2N2))			1		1	
237 2		327310	Influenza A virus (A/Kinmen/645/04(H3N2))		1		1		
237 3		327311	Influenza A virus (A/Yilan/515/03(H3N2))		1		1		
237 4		328670	Banana streak GF virus	4		4			
237 5		328671	Banana streak OL virus	4		4			
237 6		328812	Parabacteroides goldsteinii		4		5		1
237 7	X	328814	Alistipes shahii				1		1
237 9		329389	Hepatitis C virus (isolate Glasgow)	3	4	3	4		
238 0		329593	Influenza A virus (A/chicken/Fujian/25/00(H9N2))		6		6		
238 1	X	329606	Influenza A virus (A/chicken/Henan/43/02(H9N2))			1		1	
238 2		329854	Bacteroides intestinalis		1		1		
238 3		330879	Aspergillus fumigatus Af293		18		18		
238 4		331111	Escherichia coli O139:H28 str. E24377A		36		36		
238 5		331112	Escherichia coli HS		14		14		
238 6		331272	Burkholderia cenocepacia HI2424		1		1		
238 7		331635	Chlamydia pecorum E58	29		29			

238 8	331636	Chlamydia psittaci 6BC	1		1			
238 9	331978	Burkholderia pseudomallei Pasteur 52237		1		1		
239 0	332162	Candidatus Solibacter		1		1		
239 1	332193	Vaccinia Virus Acambis 3000 MVA		20		20		
239 3	332747	Influenza A virus (A/swine/Tennessee/25/1977(H1N1))	1		1			
239 4	332884	Norovirus Hu/GII.4/Sydney715D/04S/AU		1		1		
239 5	333278	H7N9 subtype	12	26	14	26	2	
239 6	333284	Hepatitis C virus (isolate Con1)	4	3	4	3		
239 7	333367	Enterocloster asparagiformis		1		1		
239 8	333668	Theileria parva strain Muguga	14	51	14	53		2
239 9	333751	Human papillomavirus type 2		7		7		
240 0	333754	Alphapapillomavirus 10	4		4			
240 1	333760	Human papillomavirus type 16	352	524	356	572	4	48
240 2	333761	human papillomavirus 18	28	104	28	112		8
240 3	333762	Human papillomavirus type 26		1		1		
240 4	333763	Human papillomavirus type 32	1	1	1	1		
240 5	333765	Human papillomavirus type 53		1		1		
240 6	333849	Enterococcus faecium DO		1		1		
240 7	333923	Human papillomavirus 5	1		1			
240 8	333990	Carnobacterium sp. AT7	1		1			
240 9	334203	Mupapillomavirus 1	1	38	1	38		
241 0	334380	Orientia tsutsugamushi str. Ikeda		1		1		
241 1	334426	Angiostrongylus costaricensis	1	1	1	1		
241 2	334440	Palythoa tuberculosa	1		1			
241 4	334590	Influenza A virus (A/Moscow/343/2003(H3N2))		1		1		
241 5	334802	Burkholderia mallei FMH		1		1		
241 6	334803	Burkholderia mallei JHU		2		2		
241 7	335333	Influenza A virus (A/New York/384/2005(H3N2))		64		64		
241 8	335337	Influenza A virus (A/New York/388/2005(H3N2))		14		14		
241 9	335358	Influenza A virus (A/New York/232/2004(H3N2))		110		110		
242 0	335812	Influenza B virus (B/Shanghai/361/2002)	1		1			
242 1	336092	Influenza A virus (A/black-headed goose/Qinghai/1/2005(H5N1))	2		2			
242 2	336205	Influenza A virus (A/swine/Bakum/5/95(H1N1))		2		2		
242 3	336207	Influenza A virus (A/swine/Bakum/1832/2000(H1N2))		3		3		
242 4	336238	Influenza A virus (A/Bar-headed Goose/Qinghai/61/05(H5N1))		8		8		
242 5	336306	Enterobacter cloacae subsp. cloacae		1		1		

242 6		336407	Rickettsia bellii RML369-C		4		4		
242 7		336794	Influenza A virus (A/equine/Massachusetts/213/2003(H3N8))		1		1		
242 8		336800	Influenza A virus (A/equine/New York/1/1999(H3N8))		3		3		
242 9		336982	Mycobacterium tuberculosis F11		24		24		
243 0		337042	Alphapapillomavirus 7	11		11			
243 1		337043	Alphapapillomavirus 4		1		1		
243 2		337052	Deltapapillomavirus 4	24		24			
243 3		337090	Chlorobium chlorochromatii		1		1		
243 5		338079	African green monkey simian foamy virus	3		3			
243 6		338188	Bacteroides fingoldii		1		1		
243 7	X	338220	Leptospira borgpetersenii serovar Sejroe			8		8	
243 8		338478	Macaque simian foamy virus	1		1			
243 9		338966	Pelobacter propionicus DSM 2379		3		3		
244 0		339670	Burkholderia ambifaria AMMD		4		4		
244 1		339854	Bacillus thuringiensis serovar israelensis ATCC 35646		4		4		
244 2		340177	Chlorobium chlorochromatii CaD3		1		1		
244 3		340184	Escherichia coli B7A		5		5		
244 4		340185	Escherichia coli E22		13		13		
244 5		340186	Escherichia coli E110019		2		2		
244 6		340197	Escherichia coli F11		12		12		
244 8		341663	Aspergillus terreus NIH2624	2		2			
244 9		341946	Woodchuck hepatitis virus 2	4		4			
245 0		341980	Human herpesvirus 3 strain Oka vaccine	4	276	4	276		
245 1		342023	Streptococcus pyogenes serotype M12	3	1	3	1		
245 2		342137	Influenza A virus (A/chicken/CA/375TR/2002(H6N2))		1		1		
245 3		342145	Influenza A virus (A/turkey/MN/735/79(H6N2))		2		2		
245 4		342207	Influenza A virus (A/shorebird/Delaware/141/2002(H9N9))		1		1		
245 5		342210	Influenza A virus (A/shorebird/Delaware/275/2001(H9N7))		1		1		
245 6		342221	Influenza A virus (A/chicken/Korea/S4/2003(H9N2))		1		1		
245 7		342396	Influenza A virus (A/turkey/Canada-Ontario/NS-01839-1/05(H3))		1		1		
245 8		342409	White spot syndrome virus	1		1			
245 9		342508	Influenza A virus (A/New York/444/2001(H1N1))		25		25		
246 1		342613	Streptococcus agalactiae 18RS21	2		2			
246 2		342614	Streptococcus agalactiae 515	1		1			

246 3		342615	Streptococcus agalactiae H36B	1		1		
246 4		342617	Streptococcus agalactiae CJB111	2		2		
246 5		343462	Human adenovirus 11p	7		7		
246 6		343463	Human adenovirus 11a	1		1		
246 7		344598	Influenza A virus (A/whooper swan/Mongolia/6/05(H5N1))		1		1	
246 8		344601	Escherichia coli B171		25		25	
246 9		344609	Shigella boydii CDC 3083-94		20		20	
247 0		344610	Escherichia coli 53638		1		1	
247 1		344702	SARS coronavirus ZJ0301		1		1	
247 2	X	344880	<i>Sphingomonas dokdonensis</i>				7	7
247 3		345072	Vibrio cholerae MO10		1		1	
247 4		345073	Vibrio cholerae O395		4		4	
247 5		345074	Vibrio cholerae RC385		2		2	
247 6		345075	Vibrio cholerae V51		3		3	
247 7		345076	Vibrio cholerae V52		32		32	
247 8		345840	Palaemon modestus	74		74		
247 9		347257	Mycoplasmopsis agalactiae PG2	3		3		
248 0		347495	Bacillus cereus F837/76		1		1	
248 1		347515	Leishmania major strain Friedlin	29	38	29	38	
248 2		348776	Mycobacterium tuberculosis C		177		177	
248 3		349101	Cereibacter sphaeroides ATCC 17029		1		1	
248 4		349163	Acidiphilium cryptum JF-5		1		1	
248 5		349741	Akkermansia muciniphila ATCC BAA-835		4		4	
248 6		349746	Yersinia pestis Angola		3		3	
248 7		349747	Yersinia pseudotuberculosis IP 31758		9		9	
248 8		349966	Yersinia frederiksenii ATCC 33641		2		2	
248 9		349967	Yersinia mollaretii ATCC 43969		3		3	
249 0		349968	Yersinia bercovieri ATCC 43970		5		5	
249 1		350054	Mycolicibacterium gilvum PYR-GCK		64		64	
249 2		350058	Mycolicibacterium vanbaalenii PYR-1		55		55	
249 3		350702	Burkholderia cenocepacia PC184		1		1	
249 4		350703	Pseudomonas aeruginosa 2192		2		2	
249 5		350704	Pseudomonas aeruginosa C3719		2		2	
249 6		351071	Newcastle disease virus AF2240	1		1		
249 7		351073	Mammalian orthoreovirus		1		1	
249 8	X	351091	<i>Oscillibacter valericigenes</i>				2	2
250 0		351277	Influenza A virus (A/Nanchang/0058/1994(H3N2))		1		1	

250 1	351581	Francisella tularensis subsp. holarctica FSC200		1		1		
250 2	351607	Acidothermus cellulolyticus 11B		2		2		
250 3	351627	Caldicellulosiruptor saccharolyticus DSM 8903		1		1		
250 4	351745	Shewanella sp. W3-18-1		1		1		
250 5	351746	Pseudomonas putida F1		5		5		
250 6	352034	Influenza A virus (A/duck/Potsdam/1402-6/1986(H5N2))	1		1			
250 7	352348	Influenza A virus (A/chicken/Hong Kong/17/1977(H6N1))		1		1		
250 8	352520	Influenza A virus (A/mallard/Alberta/35/1976(H1N1))	1	6	1	6		
250 9	352561	Influenza A virus (A/duck/Australia/341/1983(H15N8))		1		1		
251 0	352564	Influenza A virus (A/mallard/Astrakhan/263/1982(H14N5))	1		1			
251 1	352764	Influenza A virus (A/mallard/Alberta/202/1996(H2N5))	1		1			
251 2	352914	Plasmodium yoelii yoelii 17XNL	6	3	6	3		
251 3	352963	Influenza A virus (A/California/7/2004(H3N2))		1		1		
251 4	353152	Cryptosporidium parvum Iowa II		84		84		
251 5	353153	Trypanosoma cruzi strain CL Brener	2127	12	2127	12		
251 6	353154	Theileria annulata strain Ankara		4		4		
251 7	353253	Influenza A virus (A/duck/Novosibirsk/56/2005(H5N1))	6	2	6	2		
251 8	353466	Influenza A virus (A/Canterbury/236/2005(H3N2))		1		1		
251 9	354228	Influenza A virus (A/Gf/HK/SSP607/2003(H9N2))		1		1		
252 0	354235	Influenza A virus (A/Ck/HK/CSW161/2003(H9N2))		1		1		
252 1	354242	Campylobacter jejuni subsp. jejuni 81-176		4		4		
252 2	355315	Streptococcus agalactiae serogroup Ia	1		1			
252 4	356114	Hepatitis C virus genotype 3	121	20	121	20		
252 5	356386	Hepatitis C virus (isolate India)		1		1		
252 6	356391	Hepatitis C virus (isolate 6a33)		11		11		
252 7	356410	Hepatitis C virus (isolate HC-G9)	1	1	1	1		
252 8	356411	Hepatitis C virus JFH-1	8	5	8	5		
252 9	356413	Hepatitis C virus (isolate BEBE1)		14		14		
253 0	356415	Hepatitis C virus (isolate NZL1)	1	3	1	3		
253 1	356416	Hepatitis C virus (isolate HCV-K3a/650)		21		21		
253 2	356417	Hepatitis C virus (isolate JK049)		6		6		
253 3	356418	Hepatitis C virus ED43	3	1	3	1		
253 4	356419	Hepatitis C virus (isolate EUH1480)		5		5		
253 5	356421	Hepatitis C virus (isolate Th580)		7		7		

253 6		356424	Hepatitis C virus (isolate VN004)		1		1		
253 7		356426	Hepatitis C virus subtype 3a	12	105	12	105		
253 8		357205	Influenza A virus (A/chicken/Hong Kong/3123.1/2002(H5N1))		1		1		
253 9		357208	Influenza A virus (A/Vietnam/CL26/2004(H5N1))		79		79		
254 0		357210	Influenza A virus (A/Vietnam/CL100/2004(H5N1))	1		1			
254 1		357244	Orientia tsutsugamushi str. Boryong	2	2	2	2		
254 2	X	357276	<i>Phocaeicola dorei</i>				5		5
254 3		357348	Burkholderia pseudomallei 1106a		2		2		
254 4		357355	Hepatitis C virus (isolate Tr Kj)		3		3		
254 5		358708	Shigella dysenteriae 1012		1		1		
254 6		358709	Escherichia coli 101-1		5		5		
254 7		358769	Classical swine fever virus - Alfort/187	70		70			
254 8		358805	Classical swine fever virus - Alfort/Tuebingen	5		5			
254 9		358812	Classical swine fever virus - C	2		2			
255 0		359391	Brucella abortus 2308		31		31		
255 1		359787	Staphylococcus aureus subsp. aureus JH1	1	1	1	1		
255 2		359921	Influenza A virus (A/Australian shelduck/Western Australia/1756/1983(H15N2))	1		1			
255 3		360095	Bartonella bacilliformis KC583		1		1		
255 4		360102	Yersinia pestis Antiqua		36		36		
255 5		360112	Campylobacter jejuni subsp. jejuni HB93-13		2		2		
255 6		360115	Coxiella burnetii RSA 331		8		8		
255 7		360116	Coxiella burnetii 'MSU Goat Q177'		7		7		
255 8		360117	Coxiella burnetii Q321		14		14		
255 9		360118	Burkholderia pseudomallei 406e		2		2		
256 0		360549	Themiste hennahi	1		1			
256 1		362242	Mycobacterium ulcerans Agy99		15		15		
256 2		362651	Human immunodeficiency virus type 1 (isolate YU2)	16	21	16	21		
256 3		362663	Escherichia coli 536		65		65		
256 4		363020	Avian leukosis virus HPRS103	1		1			
256 6		364106	Escherichia coli UT189		2		2		
256 7		364132	Influenza A virus (A/New York/504/1998(H3N2))		1		1		
256 8		365044	Polaromonas naphthalenivorans CJ2		2		2		
257 0		365078	Influenza A virus (A/duck/Guangxi/1586/2004(H5N1))		1		1		
257 1		365080	Influenza A virus (A/duck/Guangxi/1793/2004(H5N1))		1		1		
257 2		365129	Influenza A virus (A/chicken/Vietnam/DT171/2004(H5N1))		1		1		

257 5	367120	Influenza A virus (A/Fort Monmouth/1/1947(H1N1))	1		1			
257 6	367400	Neembucu hantavirus		5		5		
257 7	367737	Aliarcobacter butzleri RM4018	1		1			
257 8	367830	Staphylococcus aureus subsp. aureus USA300	2		2			
257 9	368445	Crocodilepox virus		2		2		
258 0	370127	Influenza A virus (A/Victoria/1968(H3N2))		1		1		
258 1	370128	Influenza A virus (A/Northern Territory/60/1968(H3N2))		3		3		
258 2	370354	Entamoeba dispar SAW760		19		19		
258 3	370810	Influenza A virus (A/Anhui/1/2005(H5N1))	7		7			
258 4	370811	Influenza A virus (A/Anhui/2/2005(H5N1))	1		1			
258 5	370813	Influenza A virus (A/duck/IT/701/2005(H10N7))		1		1		
258 6	370830	Hantaan virus Q32		4		4		
258 7	371082	Influenza A virus (A/Taiwan/2/2006(H1N1))		7		7		
258 8	371094	Chikungunya virus strain S27-African prototype	24		24			
259 0	371214	Influenza A virus (A/chicken/Taiwan/0329/01(H6N1))	5		5			
259 1	371601	Bacteroides xylanisolvens	1		1	1		1
259 2	373098	Gambierdiscus toxicus	7		7			
259 4	373153	Streptococcus pneumoniae D39		60		60		
259 5	373384	Shigella flexneri 5 str. 8401		23		23		
259 6	373665	Yersinia pestis biovar Orientalis str. IP275		1		1		
259 7	374405	Influenza C virus (C/Yamagata/15/2004)	2		2			
259 8	374507	Human rotavirus G9 isolate F45	7		7			
259 9	374927	Haemophilus influenzae 22.1-21		3		3		
260 0	374928	Haemophilus influenzae PittAA		1		1		
260 1	374931	Haemophilus influenzae PittGG		1		1		
260 2	374932	Haemophilus influenzae PittHH	1		1			
260 3	374933	Haemophilus influenzae PittII		3		3		
260 4	375177	Haemophilus influenzae 3655	6	1	6	1		
260 5	375432	Haemophilus influenzae R3021		2		2		
260 6	375450	Yersinia pestis FV-1		2		2		
260 7	376619	Francisella tularensis subsp. holarctica LVS	10	301	10	301		
260 8	376721	Influenza A virus (A/chicken/Kurgan/05/2005(H5N1))	1		1			
260 9	377628	Yersinia pestis Nepal516		135		135		
261 0	378131	Influenza A virus (A/Memphis/13/1978(H1N1))		1		1		
261 1	378139	Influenza A virus (A/Memphis/1/1978(H1N1))		7		7		
261 2	378809	Ravn virus - Ravn, Kenya, 1987	2	32	2	32		

261 3		378830	Lake Victoria marburgvirus - Angola2005	8	18	8	18		
261 5		380210	Influenza A virus (A/England/333/1980(H1N1))	1	1	1	1		
261 6		380213	Influenza A virus (A/Taiwan/1/1986(H1N1))		1		1		
261 7		380217	Influenza A virus (A/swine/Hong Kong/4/1976(H3N2))		1		1		
261 8		380282	Influenza A virus (A/Fort Monmouth/1/1947-mouse adapted(H1N1))		47		47		
261 9		380284	Influenza A virus (A/England/268/1996(H7N7))	1	1	1	1		
262 0		380285	Influenza A virus (A/turkey/Ireland/1378/1983(H5N8))		1		1		
262 1		380299	Influenza A virus (A/turkey/Wisconsin/1968(H5N9))		1		1		
262 2		380301	Influenza A virus (A/turkey/Ontario/7732/1966(H5N9))	5	8	5	8		
262 3		380330	Influenza A virus (A/goose/Hong Kong/8/1976(H1N1))		2		2		
262 4		380332	Influenza A virus (A/swine/Hong Kong/273/1994(H1N1))		1		1		
262 5		380337	Influenza A virus (A/equine/Prague/1/1956(H7N7))		1		1		
262 6		380339	Influenza A virus (A/equine/Tennessee/5/1986(H3N8))		1		1		
262 7		380340	Influenza A virus (A/equine/London/1416/1973(H7N7))	1	1	1	1		
262 8		380342	Influenza A virus (A/swine/Iowa/15/1930(H1N1))	1		1			
262 9		380343	Influenza A virus (A/ruddy turnstone/New Jersey/47/1985(H4N6))		2		2		
263 0		380346	Influenza A virus (A/Wisconsin/3523/1988(H1N1))	2		2			
263 2		380836	Influenza A virus (A/chicken/Vietnam/5/2003(H5N1))		1		1		
263 3		380842	Influenza A virus (A/duck/Vietnam/15/2003(H5N1))		3		3		
263 4		380905	Influenza A virus (A/X-47(H3N2))	3	2	3	2		
263 5		380950	Influenza A virus (A/Beijing/32/1992(H3N2))		54		54		
263 6		380951	Influenza A virus (A/Hebei/19/1995(H3N2))	1		1			
263 7		380960	Influenza A virus (A/Tokyo/3/1967(H2N2))	2		2			
263 8		380964	Influenza A virus (A/Texas/36/1991(H1N1))		1		1		
263 9		380978	Influenza A virus (A/Wisconsin/4755/1994(H1N1))		5		5		
264 0		380979	Influenza A virus (A/Wisconsin/4754/1994(H1N1))		1		1		
264 1		380982	Influenza A virus (A/WSN(H1N1))		1		1		
264 2		380984	Influenza A virus (A/Leningrad/134/17/1957(H2N2))		3		3		
264 3		380985	Influenza A virus (A/Chile/1/1983(H1N1))		4		4		
264 4		381512	Influenza A virus (A/New Caledonia/20/1999(H1N1))	40	247	43	247	3	
264 5		381513	Influenza A virus (A/Panama/2007/1999(H3N2))	10	16	10	16		
264 6		381516	Influenza A virus (A/USSR/90/1977(H1N1))	12		12			

264 7	381517	Influenza A virus (A/Udorn/307/1972(H3N2))		5		5		
264 8	381518	Influenza A virus (A/Wilson-Smith/1933(H1N1))	1	158	1	158		
264 9	381666	Cupriavidus necator H16		1		1		
265 0	382781	Influenza A virus (A/Singapore/1/1957(H2N2))	1	32	1	32		
265 1	382813	Influenza A virus (A/Japan/305+/1957(H2N2))	2		2			
265 2	382825	Influenza A virus (A/Philippines/2/1982(H3N2))	1	1	1	1		
265 3	382828	Influenza A virus (A/RI/5-/1957(H2N2))		1		1		
265 4	382832	Influenza A virus (A/VM113-V1(H1N1))	1		1			
265 5	382835	Influenza A virus (A/WSN/1933(H1N1))	13	3	13	3		
265 6	382842	Influenza A virus (A/swine/29/1937(H1N1))		2		2		
265 7	382848	Influenza A virus (A/swine/Hong Kong/126/1982(H3N2))		1		1		
265 8	383028	Influenza A virus (A/New York/687/1995(H3N2))	1		1			
265 9	383121	Influenza A virus (A/Waikato/7/2000(H3N2))		2		2		
266 0	383152	Influenza A virus (A/Wellington/22/2001(H3N2))		1		1		
266 1	383231	Influenza A virus (A/Wellington/8/2004(H3N2))		2		2		
266 2	383281	Influenza A virus (A/Otago/5/2005(H1N1))		1		1		
266 3	383379	Toxoplasma gondii RH	27	23	27	23		
266 4	383531	Influenza A virus (A/swine/Italy/2/1979(H1N1))		5		5		
266 5	383537	Influenza A virus (A/swine/Iowa/1946(H1N1))		1		1		
266 6	383550	Influenza A virus (A/duck/England/1/1956(H11N6))		2		2		
266 8	383561	Influenza A virus (A/swine/Italy/1850/1977(H3N2))		1		1		
266 9	383566	Influenza A virus (A/swine/Italy/425/1976(H1N1))		1		1		
267 0	383568	Influenza A virus (A/Shanghai/11/1987(H3N2))		1		1		
267 2	383571	Influenza A virus (A/Memphis/6/1986(H3N2))	4	1	4	1		
267 3	383574	Influenza A virus (A/Memphis/2/1985(H3N2))	1		1			
267 4	383577	Influenza A virus (A/Memphis/1/1990(H3N2))		1		1		
267 5	383578	Influenza A virus (A/Memphis/4/1980(H3N2))		1		1		
267 6	383586	Influenza A virus (A/Memphis/1/1971(H3N2))	19	11	19	11		
267 7	383602	Influenza A virus (A/Anas acuta/Primorje/695/1976(H2N3))		17		17		
267 8	383603	Influenza A virus (A/turkey/Minnesota/833/1980(H4N2))		1		1		
268 0	384483	Influenza A virus (A/swine/Hong Kong/127/1982(H3N2))		1		1		
268 1	384484	Influenza A virus (A/swine/Hong Kong/81/1978(H3N2))		12		12		

268 2	384487	Influenza A virus (A/Swine/Indiana/1726/1988(H1N1))		2		2		
268 3	384493	Influenza A virus (A/seal/Mass/1/1980(H7N7))	1	1	1	1		
268 4	384495	Influenza A virus (A/Kiev/59/1979(H1N1))		5		5		
268 5	384498	Influenza A virus (A/Ann Arbor/6/1960(H2N2))		271		271		
268 6	384499	Influenza A virus (A/gull/Maryland/704/1977(H13N6))		1		1		
268 7	384500	Influenza A virus (A/Beijing/11/1956(H1N1))		30		30		
268 9	384505	Influenza A virus (A/nt/60/1968(H3N2))	1	70	1	70		
269 0	384509	Influenza A virus (A/tern/Australia/G70C/1975(H11N9))	9		9			
269 1	384510	Influenza A virus (A/tern/South Africa/1961(H5N3))	1		1			
269 2	384527	Influenza A virus (A/Cameron/1946(H1N1))		1		1		
269 3	384676	Pseudomonas entomophila L48		1		1		
269 4	384946	Influenza A virus (A/duck/Hungary/1/1970(H6N2))		1		1		
269 5	385576	Influenza A virus (A/Alaska/6/1977(H3N2))		2		2		
269 6	385580	Influenza A virus (A/duck/Ukraine/1/1963(H3N8))	3		3			
269 7	385583	Influenza A virus (A/Denver/1957(H1N1))		2		2		
269 8	385585	Influenza A virus (A/equine/Jilin/1/1989(H3N8))	1		1			
269 9	385586	Influenza A virus (A/chicken/Pennsylvania/1/1983(H5N2))		1		1		
270 0	385587	Influenza A virus (A/budgerigar/Hokkaido/1/1977(H4N6))		10		10		
270 1	385599	Influenza A virus (A/udorn/1972(H3N2))		1		1		
270 2	385600	Influenza A virus (A/Puerto Rico/8/1934(Cambridge)(H1N1))	1		1			
270 3	385605	Influenza A virus (A/equine/Kentucky/2/1986(H3N8))		3		3		
270 4	385616	Influenza A virus (A/ruddy turnstone/NJ/60/1985(N9))		1		1		
270 5	385617	Influenza A virus (A/chicken/Pennsylvania/1370/1983(H5N2))	1		1			
270 6	385624	Influenza A virus (A/Port Chalmers/1/1973(H3N2))	7	1	7	1		
270 7	385630	Influenza A virus (A/Bangkok/1/1979(H3N2))	1	7	1	7		
270 8	385640	Influenza A virus (A/Memphis/102/1972(H3N2))		8		8		
270 9	385644	Influenza A virus (A/swine/Ukkel/1/1984(H3N2))		1		1		
271 0	386032	Reston ebolavirus - Reston (1989)		4		4		
271 1	386033	Sudan ebolavirus - Uganda (2000)	1	6	1	6		
271 2	386043	Listeria welshimeri serovar 6b str. SLCC5334		3		3		
271 3	386056	Paraburkholderia ferrariae		1		1		
271 4	386415	Clostridium novyi NT		2		2		

271 5		386585	Escherichia coli O157:H7 str. Sakai		17		17		
271 6		386656	Yersinia pestis Pestoides F		2		2		
271 7		387090	Phocaeicola coprophilus		2		2		
271 8		387093	Sulfurovum sp. NBC37-1		1		1		
271 9		387139	Influenza A virus (A/Aichi/2/1968(H3N2))	12	11	13	11	1	
272 0		387147	Influenza A virus (A/England/878/1969(H3N2))	1	2	1	2		
272 1		387156	Influenza A virus (A/Harbin/1/1988(H1N2))		17		17		
272 2		387161	Influenza A virus (A/Japan/305/1957(H2N2))	5	29	5	29		
272 4		387191	Influenza A virus (A/camel/Mongolia/1982(H1N1))		8		8		
272 5		387203	Influenza A virus (A/duck/Bavaria/2/1977(H1N1))		1		1		
272 6		387207	Influenza A virus (A/duck/Hokkaido/8/1980(H3N8))		6		6		
272 8		387213	Influenza A virus (A/equine/Alaska/1/1991(H3N8))		1		1		
272 9		387223	Influenza A virus (A/equine/Miami/1963(H3N8))	5		5			
273 0		387239	Influenza A virus (A/gull/Delaware/475/1986(H2N2))		1		1		
273 1		387243	Influenza A virus (A/herring gull/DE/677/1988(H2N8))		1		1		
273 2	X	387661	Parabacteroides johnsonii				1		1
273 3		388041	Influenza A virus (A/bar-headed goose/Qinghai/1A/2005(H5N1))	1		1			
273 4		388272	Pseudomonas aeruginosa PACS2		12		12		
273 5		388634	Bombyx mandarina nuclear polyhedrosis virus		1		1		
273 6		388799	HIV-1 group O	1		1			
273 7		389259	Influenza A virus (A/Nagasaki/N03/2005(H3N2))		1		1		
273 8		390157	Senecavirus A	8		9		1	
273 9		390236	Borrelia afzelii PKo	2		2			
274 0		390333	Lactobacillus delbrueckii subsp. bulgaricus ATCC 11842 = JCM 1002	1		1			
274 1		391646	Hepatitis B virus ayw3	1		1			
274 2		391647	Hepatitis B virus ayw2	8		8			
274 3		391650	Hepatitis B virus ayw4	1		1			
274 4		391896	Rickettsia bellii OSU 85-389		3		3		
274 5		392021	Rickettsia rickettsii str. 'Sheila Smith'	1	9	1	9		
274 6	X	392172	Norovirus Hu/GII.4/MD-2004/2004/US			5		5	
274 7		392734	Terriglobus roseus		1		1		
274 8		392809	Influenza A virus (A/Victoria/3/1975(H3N2))	45	13	45	13		
274 9		392810	Influenza A virus (A/chicken/Rostock/8/1934(H7N1))	2	4	2	4		
275 0		392811	Influenza A virus (A/Udorn/8/1972(H3N2))	1		1			
275 1		393011	Francisella tularensis subsp. holarctica OSU18		6		6		

275 2	393115	Francisella tularensis subsp. tularensis FSC198		85		85		
275 3	393117	Listeria monocytogenes FSL J1-194		2		2		
275 4	393124	Listeria monocytogenes FSL N3-165		2		2		
275 5	393125	Listeria monocytogenes FSL R2-503		1		1		
275 6	393126	Listeria monocytogenes FSL R2-561		1		1		
275 7	393128	Listeria monocytogenes F6900		1		1		
275 8	393130	Listeria monocytogenes J0161		6		6		
275 9	393133	Listeria monocytogenes 10403S		4		4		
276 0	393305	Yersinia enterocolitica subsp. enterocolitica 8081		15		15		
276 2	393548	Influenza A virus (A/chicken/Tula/Russia/Oct-5/2005(H5N1))		1		1		
276 3	393557	Influenza A virus (A/Leningrad/1954/1(H1N1))		8		8		
276 4	393697	Influenza A virus (A/Liaoning/25/2000(H1N1))		1		1		
276 5	393902	Influenza A virus (A/Wisconsin/67/2005(H3N2))	1	5	1	53		48
276 6	395095	Mycobacterium tuberculosis str. Haarlem		127		127		
276 7	395494	Gallionella capsiferriformans ES-2		1		1		
276 8	395841	Influenza A virus (A/duck/Hokkaido/1130/01(H1N1))		1		1		
276 9	395842	Influenza A virus (A/tern/Australia/1/04(H2N5))		1		1		
277 0	397342	Horsepox virus		6		6		
277 2	398031	Toxoplasma gondii type III	6	1	6	1		
277 3	398812	TGEV virulent Purdue	4		4			
277 4	400331	Influenza A virus (A/New South Wales/15/1999(H3N2))		1		1		
277 5	400667	Acinetobacter baumannii ATCC 17978	13	15	13	15		
277 6	400788	Influenza A virus (A/Indonesia/5/2005(H5N1))	1	3	1	3		
277 7	400810	Influenza A virus (A/Indonesia/CDC594/2006(H5N1))	1		1			
277 8	400828	Influenza A virus (A/Indonesia/CDC669/2006(H5N1))	5		5			
277 9	400830	Influenza A virus (A/Indonesia/CDC699/2006(H5N1))	2		2			
278 1	401650	Listeria monocytogenes HPB2262		3		3		
278 2	401671	HIV-1 M:B_89.6		1		1		
278 3	402194	Influenza A virus (A/Istanbul/CTF/10/2004(H3N2))		1		1		
278 4	402464	Influenza A virus (A/gray teal/Australia/2/1979(H4N4))		1		1		
278 5	402489	Influenza A virus (A/turkey/Minnesota/1200/1980(H7N3))		1		1		
278 6	402518	Influenza A virus (A/Canada/rv504/2004(H7N3))	1		1			
278 7	402527	Influenza A virus (A/chicken/Scotland/1959(H5N1))		1		1		
278 8	402648	Streptomyces hainanensis		1		1		

278 9		402938	Influenza A virus (A/Michigan/2/2003(H1N2))	1		1		
279 0		404137	Influenza A virus (A/chicken/Beijing/nl/2002(H9N2))		1	1		
279 2		404150	Influenza A virus (A/chicken/Hubei/wk/1997(H5N1))		2	2		
279 4		404155	Influenza A virus (A/chicken/Hunan/fg/2004(H5N1))		2	2		
279 5		404163	Influenza A virus (A/chicken/Jilin/hg/2002(H5N1))		1	1		
279 6		404330	Streptococcus pyogenes serotype M2	2		2		
279 7		404331	Streptococcus pyogenes serotype M4	2		2		
279 8		404408	Influenza A virus (A/chicken/Shanxi/2/2006(H5N1))	1		1		
279 9		404573	Influenza A virus (A/chicken/Netherlands/03010132/03(H7N7))	1		1		
280 0		404974	Vibrio cholerae AM-19226		1	1		
280 1		405050	Influenza A virus (A/tern/Australia/1402/2004(H2N5))		1	1		
280 2		405531	Bacillus cereus G9842		3	3		
280 3		405532	Bacillus cereus B4264		1	1		
280 4		405533	Bacillus cereus AH1134		3	3		
280 5		405536	Bacillus anthracis str. Tsiankovskii-I		1	1		
280 6		405917	Bacillus cereus W		1	1		
280 7		405955	Escherichia coli APEC O1		8	8		
280 8		406269	West Nile virus strain PTRoxo		12	12		
280 9		407134	Yellow fever virus strain Ghana/Asibi/1927		5	5		
281 0		407141	Yellow fever virus isolate Ethiopia/Couma/1961		2	2		
281 1		407148	Campylobacter jejuni subsp. jejuni 81116		14	14		
281 2		407418	Influenza A virus (A/chicken/Guiyang/2147/2005(H5N1))		1	1		
281 3	X	408598	Human rotavirus G1P[8]			1		1
281 4		408599	Human rotavirus G9P[8]	1		1		
281 5	X	408657	Dyadobacter koreensis				1	1
281 6		408685	Dengue virus 1 Brazil/97-11/1997		1058	1058		
281 7		408686	Dengue virus 4 Philippines/H241/1956	4	947	4	947	
281 8		408687	Dengue virus 4 Singapore/8976/1995		5	5		
281 9		408688	Dengue virus 4 Thailand/0348/1991		15	15		
282 0		408689	Dengue virus 4 Thailand/0476/1997		3	3		
282 1		408690	Dengue virus 3 China/80-2/1980		84	84		
282 2		408691	Dengue virus 3 Martinique/1243/1999		889	889		
282 3		408692	Dengue virus 3 Sri Lanka/1266/2000		66	66		
282 4		408693	Dengue virus 3 Singapore/8120/1995		69	69		

2825	408694	Dengue virus 2 Peru/IQT2913/1996		87		87		
2826	408870	Dengue virus 3 Philippines/H87/1956	8	13	8	13		
2827	408871	Dengue virus 4 Dominica/814669/1981	1	256	8	256	7	
2828	410069	Streptococcus pyogenes serotype M80	1		1			
2829	410072	Coprococcus comes	1		1			
2830	410078	Human respiratory syncytial virus S2	1		1			
2831	410289	Mycobacterium tuberculosis variant bovis BCG str. Pasteur 1173P2	1	230	1	230		
2832	411157	Influenza A virus (A/Thailand/271/2005(H1N1))	1		1			
2833	411167	Influenza A virus (A/Queensland/11/2001(H3N2))		1		1		
2834	411662	Influenza A virus (A/Hanoi/TN405/2005(H3N2))		1		1		
2835	412022	Burkholderia mallei NCTC 10229		9		9		
2836	412419	Borrelia duttonii Ly	12		12			
2837	412420	Yersinia pestis CA88-4125		21		21		
2838	412614	Vibrio cholerae 2740-80		44		44		
2839	412694	Bacillus thuringiensis str. Al Hakam		7		7		
2840	412883	Vibrio cholerae MZO-3		1		1		
2841	412966	Vibrio cholerae 1587		3		3		
2842	412967	Vibrio cholerae MAK 757		9		9		
2843	413502	Cronobacter turicensis	1		1			
2844	413999	Clostridium botulinum A str. ATCC 3502		3		3		
2845	414044	Influenza A virus (A/Thailand/1(KAN-1A)/2004(H5N1))	1		1			
2846	415166	Influenza A virus(A/swine/Henan/2/2004(H9N2))		2		2		
2847	415167	Influenza A virus(A/swine/Henan/3/2004(H9N2))		1		1		
2848	415169	Influenza A virus(A/swine/Henan/5/2004(H9N2))		1		1		
2849	415172	Influenza A virus(A/swine/Henan/8/2004(H9N2))		2		2		
2850	415176	Influenza A virus(A/swine/Shandong/fNY/2003(H9N2))		3		3		
2851	415177	Influenza A virus(A/swine/Shandong/fZC/2003(H9N2))		7		7		
2852	416035	Norovirus Hu/GII.4/DenHaag89/2006/NL	1		3		2	
2854	416276	Rickettsia massiliae MTU5		2		2		
2855	416674	Influenza B virus (B/Yamagata/16/1988)	5	1	5	1		
2856	416730	Influenza A virus (A/Memphis/10/1996(H1N1))		1		1		
2857	417398	Vibrio cholerae MZO-2		2		2		
2858	417399	Vibrio cholerae NCTC 8457		1		1		
2859	417400	Vibrio cholerae B33		5		5		

286 0	417758	Influenza A virus (A/quail/Shantou/1475/2004(H9N2))	1		1			
286 1	418136	Francisella tularensis subsp. tularensis WY96-3418		2		2		
286 2	418868	Influenza A virus (A/Egypt/14724-NAMRU3/2006(H5N1))	1		1			
286 3	419109	Vibrio parahaemolyticus AQ3810		38		38		
286 4	419612	Camelus ferus		1		1		
286 5	419947	Mycobacterium tuberculosis H37Ra		130		130		
286 6	420174	Hepatitis C virus isolate HC-J4	23	180	23	180		
286 7	420245	Leishmania braziliensis MHOM/BR/75/M2904	14	18	14	18		
286 8	420246	Geobacillus thermodenitrificans NG80-2	2		2			
286 9	420662	Methylibium petroleiphilum PM1		1		1		
287 0	421472	Influenza A virus (A/Indonesia/CDC1032N/2007(H5N1))	3		3			
287 1	421877	Hepatitis C virus isolate HC-J1	3	26	3	26		
287 2	421901	Influenza A virus (A/Egypt/0636-NAMRU3/2007(H5N1))	2	1	2	1		
287 3	423368	Salmonella enterica subsp. enterica serovar Newport str. SL254		1		1		
287 4	424362	Bat coronavirus HKU4-4		1		1		
287 5	424717	Shigella flexneri 3a	4		4			
287 6	424718	Shigella flexneri 5a	2		2			
287 7	424720	Shigella flexneri Y	8		8			
287 8	425067	Burkholderia pseudomallei 305		2		2		
287 9	425088	Tanganya virus		1		1		
288 0	425551	Influenza A virus (A/AA/Huston/1945(H1N1))	1		1			
288 1	425557	Influenza A virus (A/California/10/1978(H1N1))		1		1		
288 2	426430	Staphylococcus aureus subsp. aureus str. Newman	7		7			
288 3	426738	Influenza A virus (A/swine/Miyagi/5/2003(H1N2))		2		2		
288 4	427773	Influenza A virus (A/Thailand/SP83/2004(H5N1))	1		1			
288 5	427826	Influenza B virus (B/Hong Kong/8/1973)		1		1		
288 6	427890	Influenza A virus (A/Viet Nam/JP4207/2005(H5N1))	1		1			
288 8	430066	Brucella abortus S19		15		15		
288 9	430417	Influenza A virus (A/Wyoming/3e5/2003(H3))	1		1			
289 0	430511	Juquitiba virus		2		2		
289 1	430557	Francisella tularensis subsp. tularensis FSC033		27		27		
289 2	431944	Magnetospirillum gryphiswaldense MSR-1		1		1		
289 3	431947	Porphyromonas gingivalis ATCC 33277	20	4	20	4		
289 4	432070	Influenza A virus (A/Hong Kong/213/2003(H5N1))	1	4	1	4		

289 5	432094	Influenza A virus (A/Wyoming/3/2003-X-147(H3N2))		1		1		
289 6	432359	Toxoplasma gondii VEG	14		14			
289 7	434922	Coxiella burnetii Dugway 5J108-111		8		8		
289 8	434923	Coxiella burnetii CbuG_Q212		1		1		
289 9	434924	Coxiella burnetii CbuK_Q154		1		1		
290 0	435258	Leishmania infantum JPCM5	28	19	28	19		
290 1	435359	Influenza A virus (A/chicken/Hebei/706/2005(H5N1))		2		2		
290 2	436004	Influenza A virus (A/kunke/1/71(H3N2))		1		1		
290 3	439184	Escherichia coli RS218	1		1			
290 4	439220	Streptococcus caballi		1		1		
290 5	439235	Desulfatibacillum alkenivorans AK-01		1		1		
290 6	439843	Salmonella enterica subsp. enterica serovar Schwarzengrund str. CVM19633		1		1		
290 7	439847	Salmonella enterica subsp. enterica serovar Saintpaul str. SARA29		1		1		
290 8	440534	Salmonella enterica subsp. enterica serovar 4,[5],12:i:- str. CVM23701		1		1		
290 9	441158	Burkholderia pseudomallei 9		1		1		
291 0	441159	Burkholderia pseudomallei 91		2		2		
291 1	441160	Burkholderia pseudomallei 14		1		1		
291 2	441769	Bacillus coahuilensis m4-4		1		1		
291 3	441771	Clostridium botulinum A str. Hall	45		45			
291 4	441962	Influenza A virus (A/duck/Mongolia/47/2001(H7N1))		3		3		
291 5	443149	Mycobacterium tuberculosis CCDC5079		3		3		
291 6	443239	Human coronavirus HKU1 (isolate N1)	1		1	1		1
291 7	443240	Human coronavirus HKU1 (isolate N2)	1		1			
291 8	443241	Human coronavirus HKU1 (isolate N5)	1		2	1	1	1
291 9	444178	Brucella ovis ATCC 25840		8		8		
292 0	444185	Simian rotavirus A strain RRV	10	1	10	1		
292 1	444186	Simian rotavirus A strain TUCH		5		5		
292 2	444318	Influenza A virus (A/Texas/1/1977(H3N2))	1	15	1	15		
292 3	444793	Influenza A virus (A/Auckland/590/2000(H1N1))		1		1		
292 4	444922	Psychromonas ossibalaenae		1		1		
292 5	445027	Influenza A virus (A/Beijing/01/2003(H5N1))	1		1			
292 6	445335	Clostridium botulinum NCTC 2916		2		2		
292 7	445338	Clostridium botulinum str. Iwanei E	6		6			
292 8	445983	Borrelia burgdorferi 156a	1		1			
292 9	445985	Borrelia burgdorferi ZS7	3	45	3	45		

2930		446023	Influenza A virus (A/Hong Kong/68(H3N2))	1		1			
2931		446679	Nostoc sphaeroides		1		1		
2933		447095	Blastomyces dermatitidis ATCC 26199		1		1		
2934		447106	Leptospira licerasiae		1		1		
2935		447133	Influenza A virus (A/chicken/Japan/1925(H7N7))	1		1			
2936		447188	Influenza A virus (A/duck/Tasmania/277/2007(H7N2))	1		1			
2937		448058	Influenza A virus (A/swine/Guangdong/01/2005(H3N2))		1		1		
2938		449278	West Nile virus SPU116/89		1		1		
2939		449656	Influenza A virus (A/chicken/Taiwan/2838V/00(H6N1))	15		15			
2940		451515	Staphylococcus aureus subsp. aureus USA300_FPR3757		1		1		
2941		451516	Staphylococcus aureus subsp. aureus USA300_TCH1516	1		1			
2942		451707	Bacillus cereus NVH0597-99		7		7		
2943		451708	Bacillus cereus H3081.97		12		12		
2944		451709	Bacillus cereus 03BB108		8		8		
2945		451804	Aspergillus fumigatus A1163		13		13		
2946		452646	Neogale vison		1		1		
2947		452652	Kitasatospora setae KM-6054		1		1		
2948		452659	Rickettsia rickettsii str. Iowa		1		1		
2949		453057	Influenza A virus (A/Indonesia/CDC540/2006(H5N1))	1		1			
2950		453927	Juniperus formosana	1		1			
2951	X	454951	Feline coronavirus UU3			1		1	
2952		456999	Rhizoctonia solani		1		1		
2953		458234	Francisella tularensis subsp. holarctica FTNF002-00		1		1		
2954		458678	Hantaanvirus CGRn93P8		2		2		
2955		461739	Influenza B virus (B/Florida/4/2006)	4		4			
2956		461772	Influenza A virus (A/Wisconsin/43/2006(H3N2))	1		1			
2957		462695	Influenza A virus (A/Hiroshima/52/2005(H3))	2		2			
2958		463676	Rhinovirus C		14		14		
2959		463722	Murine norovirus GV/CR6/2005/USA		1		1		
2960		464314	Influenza A virus (A/Anhui/T2/2006(H5N1))	1		1			
2961		464417	Influenza B virus (B/Malaysia/2506/2004)	1		1	238		238
2962		464622	Influenza A virus (A/Solomon Islands/3/2006 (Egg passage)(H1N1))	1		1			
2963		464623	Influenza A virus (A/Solomon Islands/3/2006(H1N1))	18		19		1	
2964		465541	Streptomyces sp. Mg1		1		1		

2965		467144	Modified Vaccinia Ankara virus		23		23		
2966		468968	Influenza A virus (A/Indonesia/560H/2006(H5N1))	1		1			
2967	X	469009	'Brassica napus' phytoplasma			1		1	
2969		470137	Brucella suis ATCC 23445	1	5	1	5		
2970		471223	Geobacillus sp. WCH70		2		2		
2971		471721	HIV-1 CRF01_AE	1		1			
2973		472511	Influenza A virus (A/Indonesia/TLL007/2006(H5N1))	1		1			
2974		475478	Influenza A virus (A/equine/New Market/1/1979(H3N8))	1		1			
2975		475493	Influenza A virus (A/equine/California/8560/2002(H3N8))		2		2		
2976		476176	Moraea pallida	1		1			
2978		476294	Influenza A virus (A/Brisbane/10/2007(H3N2))	5		5			
2979		476303	Influenza A virus (A/California/04/2007(H1N1))		1		1		
2980		478005	Escherichia coli O157:H7 str. EC4486		1		1		
2981		478006	Escherichia coli O157:H7 str. EC4501		2		2		
2982		478008	Escherichia coli O157:H7 str. EC869		13		13		
2983		479022	Influenza A virus (A/chicken/Korea/IS/2006(H5N1))	1		1			
2984	X	479063	Norovirus Hu/Cumberland/2004/USA			1		1	
2985		479935	Gluconobacter morbifer		1		1		
2986		480019	Influenza A virus (A/Moscow/10/1999(H3N2))	1	2	1	2		
2987		480024	Influenza A virus (A/Wyoming/03/2003(H3N2))		8		8		
2988		480116	Hepatitis B virus adw2/Rutter/1979		1		1		
2989		481805	Escherichia coli ATCC 8739		1		1		
2990		482134	Hepatitis B virus adw/Japan/Nishioka/1983		1		1		
2991		482628	Campylobacter jejuni subsp. jejuni BH-01-0142		1		1		
2992		482957	Burkholderia lata		2		2		
2993		483179	Brucella canis ATCC 23365		9		9		
2994		484021	Klebsiella pneumoniae subsp. pneumoniae NTUH-K2044		1		1		
2995		486280	Torque teno virus VT416		3		3		
2996		486619	Bacillus anthracis str. A0193		3		3		
2997		486620	Bacillus anthracis str. A0465		5		5		
2998		486621	Bacillus anthracis str. A0442		1		1		
2999		486623	Bacillus anthracis str. A0389		5		5		
3000		486624	Bacillus anthracis str. A0488		91		91		
3001		487067	Torque teno virus Human/Ghana/GH1/1996		3		3		
3002		487317	Aequorivita capsosiphonis		1		1		

3003		487521	Mycobacterium intracellulare ATCC 13950	1		1			
3004		488233	Influenza A virus (A/Iran/1/1957(H2N2))		1		1		
3005		488241	Influenza A virus (A/Korea/426/1968(H2N2))		2		2		
3006		489455	hepatitis B virus genotype A	1	1	1	9		8
3007		489460	Hepatitis B virus genotype B	1	18	1	23		5
3008		489469	Hepatitis B virus ayw/China/Tibet127/2002	1		1			
3009		489821	Norovirus GII.4	3	16	3	16		
3010		489822	Norovirus Hu/Houston/TCH186/2002/US	2		2			
3011		490039	Norovirus GII.2	1		1			
3012		490041	Norovirus GII.3	1		1			
3013		490133	Hepatitis B virus ayw/France/Tiollais/1979		4		4		
3014		493803	Merkel cell polyomavirus	2	30	2	30		
3015		496036	Influenza A virus (A/duck/Vietnam/37/2007(H5N1))		1		1		
3016		497976	Salmonella enterica subsp. enterica serovar Typhi str. E00-7866		1		1		
3017		497977	Salmonella enterica subsp. enterica serovar Typhi str. 404ty		1		1		
3019		499191	Norovirus GII.1	1		1			
3020		499286	Influenza A virus (A/Hong Kong/CUHK13527/2003(H3N2))		2		2		
3021	X	502049	<i>Thalassospira profundimaris</i>			1			1
3022		502057	Vaccinia virus GLV-1h68		2		2		
3023	X	502101	<i>Feline coronavirus UU2</i>			1			1
3024		502780	Paracoccidioides brasiliensis Pb18	1	1	1	1		
3025		502790	Aeromonas diversa		1		1		
3026		503386	Influenza B virus (B/Kobe/3/2004)	1		1			
3027		504660	Influenza B virus (B/Kobe/67/2005)	1		1			
3028		504662	Influenza B virus (B/Kobe/39/2005)	1		1			
3029		504664	Influenza B virus (B/Kobe/113/2005)	1		1			
3030		504666	Influenza B virus (B/Kobe/115/2005-T1)	1		1			
3031		504697	Influenza A virus (A/Cygnus olor/Czech Republic/10732/07(H5N1))		1		1		
3032		504904	Influenza A virus (A/Brisbane/59/2007(H1N1))	26	14	26	14		
3033		504910	Influenza A virus (A/Wellington/75/2006(H1N1))		1		1		
3034		505184	HIV-1 M:A	9	1	9	1		
3035		505185	HIV-1 M:B	8	25	8	25		
3036		505186	HIV-1 M:C	11		11			
3037		505228	HIV-1 M:G	4		4			
3038		506282	Influenza A virus (A/unknown/New York/11646-5/2005(H7N2))	1		1			

303 9		506350	Influenza A virus (A/Hong Kong/1/1968(H3N2))	22	3	22	3		
304 1		506380	Influenza A virus (A/Hong Kong/1-4-MA21-1/1968(H3N2))	1		1			
304 2		507601	Toxoplasma gondii GT1		205		205		
304 4		508767	Clostridium botulinum E3 str. Alaska E43		1		1		
304 5		508771	Toxoplasma gondii ME49	21	89	21	89		
304 6		508775	Norovirus GII.9		1		1		
304 7		509173	Acinetobacter baumannii AYE		1		1		
304 8		509628	Hepatitis E virus type 3	1	1	1	1		
304 9		510222	Influenza A virus (A/duck/Vietnam/NCVD-9/2007(H5N1))	1		1			
305 0		510670	Hepatitis delta virus dTk6		1		1		
305 1		511145	Escherichia coli str. K-12 substr. MG1655		31		31		
305 2		511839	Influenza A virus (A/turkey/England/N28/1973(H5N2))	1		1			
305 3		514118	Influenza A virus (A/BRISBANE/9/2006(H3N2))		1		1		
305 4		514193	Influenza A virus (A/CHU/2-524/2005(H3N2))		2		2		
305 5		515581	Influenza A virus (A/swine/Ohio/24366/2007(H1N1))		2		2		
305 7		515817	Amphioctopus fangsiao	1		1			
305 8		518922	Influenza A virus (A/Beijing/262/1995(H1N1))	5		6		1	
305 9		518968	Influenza A virus (A/X-31B(Puerto Rico/8/1934-Aichi/2/1968)(H3N2))	2		2			
306 0		518987	Influenza B virus (B/Lee/1940)	5	3	5	3		
306 1		519082	Influenza A virus (A/New York/107/2003(H7N2))	1		1			
306 2		519424	Alkalihalobacillus trypoxylicola		1		1		
306 4		520450	Brucella abortus bv. 2 str. 86/8/59		1		1		
306 5		520464	Brucella melitensis bv. 1 str. Rev.1	1		1			
306 6		520963	Norovirus GII.11	1		1			
306 7		520964	Norovirus GII.18	1		1			
306 8		521004	Haemophilus influenzae 6P18H1		1		1		
306 9		521005	Haemophilus influenzae 7P49H1		1		1		
307 0		521007	Borrelia burgdorferi N40	4	3	4	3		
307 1	X	521009	Borrelia burgdorferi 297			7		7	
307 2		522740	Influenza A virus (A/swine/Hannover/1/1981(H1N1))		12		12		
307 3		524364	Norovirus Hu/1968/US	6	1	6	1		
307 4		524651	Muromegalovirus WP15B		1		1		
307 5		525281	Escherichia coli 83972		3		3		
307 6		525368	Mycobacterium parascrofulaceum ATCC BAA-614		35		35		
307 7		525374	Staphylococcus epidermidis BCM-HMP0060		1		1		

307 8		526974	Bacillus cereus BDRD-ST24		1		1		
307 9		526977	Bacillus cereus ATCC 4342		1		1		
308 1		527024	Bacillus thuringiensis serovar tochiensis BGSC 4Y1		1		1		
308 2		527026	Bacillus thuringiensis serovar sotto str. T04001		1		1		
308 4		528354	Neisseria gonorrhoeae MS11	8		8			
308 5		529058	Influenza A virus (A/Uruguay/716/2007(H3N2))	1		1			
308 6		529507	Proteus mirabilis HI4320	1		1			
308 7		533026	Influenza A virus (A/turkey/Ohio/313053/2004(H3N2))	1	2	1	2		
308 8		535026	Bacillus subtilis subsp. subtilis NCIB 3610 = ATCC 6051 = DSM 10		1		1		
308 9		535852	Influenza A virus (A/California/06/2008(H1N1))		1		1		
309 0		538123	Mammalian orthoreovirus 3		1		1		
309 1		538555	Influenza A virus (A/swine/Korea/Hongsong2/2004(H1N2))		2		2		
309 2		543153	Mycobacterium phage Predator		1		1		
309 3		543824	Synechocystis aquatilis		1		1		
309 4		545501	Oceanobacillus oncorhynchi		1		1		
309 5		546980	Norovirus Hu/GII-4/Saga1/2006/JP	1	1	1	1		
309 6		548470	Staphylococcus aureus subsp. aureus MN8	11		11			
309 7		548473	Staphylococcus aureus subsp. aureus TCH60	47		47			
309 8		549169	Pseudogulbenkiania ferrooxidans		1		1		
310 0		552536	Listeria monocytogenes HCC23		2		2		
310 1		552592	Norovirus GII.17	1		2		1	
310 2		553345	Influenza A virus (A/Aichi/72/2007(H3N2))	1		1			
310 3		553583	Staphylococcus aureus A9635	1		1			
310 4		554290	Salmonella enterica subsp. enterica serovar Paratyphi A str. AKU_12601		3		3		
310 5		556277	Influenza A virus (A/swine/Guangdong/02/2005(H3N2))	1		1			
310 6		557433	Limosilactobacillus reuteri subsp. reuteri JCM 1112	3		3			
310 7		557599	Mycobacterium kansasii ATCC 12478	1		1			
310 8	X	557723	Glaesserella parasuis SH0165			1		1	
310 9		558177	Influenza A virus (A/mallard/Bavaria/10/2007(H5N1))		1		1		
311 0		558553	Crassostrea angulata	15		31		16	
311 1		559292	Saccharomyces cerevisiae S288C	16	2	16	2		
311 2		559561	Aureobasidium namibiae		1		1		
311 3		560463	Influenza A virus (A/Taiwan/VGHYM0325-06/2002(H3N2))		1		1		
311 6		562988	Influenza A virus (A/Egypt/2289-NAMRU3/2008(H5N1))	1		1			

311 7		562992	Influenza A virus (A/Egypt/3300-NAMRU3/2008(H5N1))	1		1		
311 8		563041	Helicobacter pylori G27	2		2		
311 9		563466	Scedosporium apiospermum		1		1	
312 0		563773	Vibrio parahaemolyticus AN-5034		4		4	
312 1		565995	Bundibugyo ebolavirus	17		17		
312 3		566466	gamma proteobacterium NOR5-3		1		1	
312 4		567106	Campylobacter jejuni subsp. jejuni IA3902		1		1	
312 5		568206	Bacillus anthracis str. CDC 684		3		3	
312 6		568819	Listeria monocytogenes serotype 4b str. CLIP 80459		6		6	
312 7		568833	Influenza A virus (A/chicken/Jiangsu/7/2002(H9N2))		1		1	
312 8		571800	Psychrobacter sp. G	1		1		
312 9		572264	Bacillus cereus 03BB102		2		2	
313 0		572307	Neospora caninum Liverpool		1		1	
313 1	X	573199	Murine adenovirus 3			1		1
313 2		573729	Thermothelomyces thermophilus ATCC 42464	1		1		
313 3		573797	Influenza A virus (A/England/654/2007(H1N1))		1		1	
313 4		573977	Human bocavirus 2	4		4		
313 5		574556	Anaplasma centrale str. Israel		2		2	
313 6		575591	Brucella abortus NCTC 8038		1		1	
313 7		575788	Vibrio atlanticus LGP32		1		1	
313 9		577543	Influenza A virus (A/Xinjiang/1/2006(H5N1))	4		4		
314 0		578079	Influenza A virus (A/ferret/Maryland/P10-UMD/2008(H9N2))		1		1	
314 1		578831	Rotavirus A human/USA/D/1974 G1P1A[8]		23		23	
314 2		580047	Chlamydia trachomatis A2497	3		3		
314 3		580165	Bacillus cytotoxicus		2		2	
314 4		585034	Escherichia coli IA1	3		3		
314 5		586029	Fowl adenovirus 8b	2		2		
314 6		587200	Variola virus human/India/Ind3/1967		4		4	
314 7		587884	Influenza A virus (A/Sydney/5/1997(H3N2))	7		7		
314 8		588533	Norovirus Hu/GII/GZ-1/2008/CHN	1		1		
314 9		588596	Rhizophagus irregularis		1		1	
315 0		591020	Shigella flexneri 2002017		1		1	
315 1		592021	Bacillus anthracis str. A0248		1		1	
315 2		592313	Vibrio cholerae 12129(1)		1		1	
315 4		595609	Recombinant Hepatitis C virus HK6a/JFH-1	8		8		
315 5		598745	Giardia intestinalis ATCC 50581		7		7	

315 6	604436	Influenza B virus (B/Brisbane/60/2008)	14		14			
315 7	614035	Influenza A virus (A/swine/Belgium/1/1998(H1N1))	1		1			
315 8	616922	Influenza A virus (A/chicken/Anhui/1/1998(H9N2))		1		1		
315 9	617102	Yellow fever virus 17D/Tiantan	4	5	4	6		1
316 0	619693	Prevotella sp. oral taxon 472 str. F0295	1		1			
316 2	627442	SARS coronavirus P2	1		1			
316 3	627611	Vibrio parahaemolyticus K5030		22		22		
316 4	630277	Tarsius lariang		1		1		
316 5	632134	Influenza A virus (A/chicken/Pakistan/UDL-01/2008(H9N2))	7		7			
316 6	632955	Acinetobacter rudis		1		1		
316 7	633137	SARS coronavirus MA15	1		1			
316 8	633557	Infectious bronchitis virus CK/CH/LHLJ/04V	1		1			
316 9	634464	Chlamydia trachomatis Sweden2	1		1			
317 0	637383	Helicobacter pylori NCTC 11639		8		8		
317 1	638313	Human bocavirus 3	6		6			
317 2	641140	Brucella abortus str. 2308 A		1		1		
317 3	641501	Influenza A virus (A/California/04/2009(H1N1))	128	128	138	128	10	
317 4	641809	Influenza A virus (A/California/07/2009(H1N1))	36	68	46	68	10	
317 5	641812	Influenza A virus (A/Texas/05/2009(H1N1))	1		1			
317 6	642261	Influenza A virus (A/California/08/2009(H1N1))	8		8			
317 7	642794	Influenza A virus (A/Auckland/1/2009(H1N1))		1		1		
317 8	643212	Influenza A virus (A/Netherlands/602/2009(H1N1))	7	14	11	14	4	
317 9	643680	Saccharomyces cerevisiae EC1118		1		1		
318 0	644289	Influenza A virus (A/Korea/01/2009(H1N1))	4		4			
318 1	644788	Influenza A virus (A/Vietnam/1194/2004(H5N1))	14	10	14	10		
318 2	645463	Clostridioides difficile R20291	25		25			
318 3	645724	Influenza A virus (A/Hong Kong/01/2009(H1N1))	3		3			
318 4	645771	Influenza A virus (A/Catalonia/63/2009(H1N1))		1		1		
318 5	647096	Influenza A virus (A/Beijing/01/2009(H1N1))		35		35		
318 6	647343	Norovirus Hu/GII.4/Hiroshima/48/2004/JPN		1		1		
318 7	647514	Norovirus GI.1	1		1			
318 8	647515	Norovirus GI.2	1		1			
318 9	647516	Norovirus GI.3		1		1		
319 0	647519	Norovirus GI.5	1		1			
319 1	647521	Norovirus GI.7	1		1			

319 2		647923	Influenza A virus (A/reassortant/NYMC X-179A(California/07/2009 x NYMC X-157)(H1N1))	16	6	16	6		
319 3		648194	Neisseria meningitidis serogroup Y	3		3			
319 4		648856	Influenza A virus (A/Beijing/501/2009(H1N1))	1		1			
319 6	X	650130	Human rhinovirus 89 ATCC VR-1199			2		2	
319 7		650131	Equine rhinitis B virus 1 strain P1436/71	6		6			
319 8		651183	Aurantimonas manganooxydans		1		1		
319 9		652616	Mycobacterium tuberculosis str. Erdman = ATCC 35801	15	19	15	19		
320 0	X	652961	Crimean-Congo hemorrhagic fever virus strain IbAr10200			25		25	
320 2		654811	Influenza A virus (A/Perth/16/2009(H3N2))	6		7		1	
320 3		655288	Influenza A virus (A/mallard/Quebec/10969/2006(H2N3))		1		1		
320 4		655827	Metarhizium acridum CQMa 102	1		1			
320 5		655863	Grosmannia clavigera kw1407		1		1		
320 6		657165	Influenza A virus (A/Hong Kong/213/03)	2	2	2	2		
320 7		658578	Influenza A virus (A/duck/Beijing/MG0617/2005(H9N2))		1		1		
320 8		658655	Lachnospiraceae bacterium 1_4_56FAA		1		1		
320 9		658858	Giardia lamblia P15		1		1		
321 0	X	660469	Cryphonectria parasitica EP155			1		1	
321 1		660658	Norovirus Hu/GII.4/CHDC4108/1987/US		2		2		
321 3		661367	Legionella longbeachae NSW150		1		1		
321 5		667015	Phocaeicola salanitronis DSM 18170		1		1		
321 6		672161	Chlamydia trachomatis B/TZ1A828/OT	3		3			
321 7		673446	Influenza A virus (A/reassortant/NYMC X-181(California/07/2009 x NYMC X-157)(H1N1))	2		2			
321 8	X	674529	Bacteroides faecis				1	1	
321 9		679206	Escherichia coli MS 119-7		1		1		
322 0		679716	Trypanosoma brucei gambiense DAL972		1		1		
322 1		680689	Influenza A virus (A/England/AV877/1996(H7N7))		1		1		
322 2		680690	Influenza A virus (A/Hong Kong/1074/1999(H9N2))	2	1	2	1		
322 3		680692	Influenza A virus (A/Hong Kong/485/1997(H5N1))	1	1	1	1		
322 4		680693	Influenza A virus (A/Netherlands/219/2003(H7N7))	14		14			
322 5		680729	Influenza A virus (A/chicken/Italy/1067/1999(H7N1))		20		20		
322 6		680739	Influenza A virus (A/chicken/Mexico/37821-771/1996(H5N2))		1		1		

322 7		680767	Influenza A virus (A/duck/Hong Kong/140/1998(H5N1))		1		1		
322 8		680785	Influenza A virus (A/environment/Hong Kong/156/1997(H5N1))	1		1			
322 9		680789	Influenza A virus (A/environment/Viet Nam/1203/2004(H5N1))	1		1			
323 0		680799	Influenza A virus (A/goose/Hong Kong/437-6/1999(H5N1))		1		1		
323 1		680803	Influenza A virus (A/goose/Hong Kong/739.2/2002(H5N1))		1		1		
323 2		682728	Influenza A virus (A/Quebec/144147/2009(H1N1))	10		10			
323 3		683780	Influenza A virus (A/wild bird/Korea/A81/2009(H5N2))	1		1			
323 4	X	686984	Sapelovirus A			1		1	
323 5		687340	Torque teno virus 1		3		3		
323 6		687341	Torque teno virus 2		4		4		
323 7		687342	Torque teno virus 3		4		4		
323 8		687343	Torque teno virus 4		6		6		
323 9		687345	Torque teno virus 6		2		2		
324 0		687346	Torque teno virus 7		3		3		
324 1		687347	Torque teno virus 8		6		6		
324 2		687350	Torque teno virus 11		1		1		
324 3		687353	Torque teno virus 14		5		5		
324 4		687354	Torque teno virus 15		3		3		
324 5		687355	Torque teno virus 16		4		4		
324 6		687358	Torque teno virus 19		5		5		
324 7		687359	Torque teno virus 20		2		2		
324 8		687360	Torque teno virus 21		2		2		
324 9		687362	Torque teno virus 23		2		2		
325 0		687363	Torque teno virus 24		3		3		
325 1		687364	Torque teno virus 25		3		3		
325 2		687365	Torque teno virus 26		5		5		
325 3		687366	Torque teno virus 27		5		5		
325 4		687367	Torque teno virus 28		3		3		
325 5		687368	Torque teno virus 29		4		4		
325 6		687369	Torque teno mini virus 1		1		1		
325 7		687371	Torque teno mini virus 3		2		2		
325 8		687375	Torque teno mini virus 7		2		2		
325 9		687376	Torque teno mini virus 8		2		2		
326 0		687382	Torque teno tamarin virus		2		2		
326 1		687383	Torque teno douroucouli virus		5		5		
326 2		687384	Torque teno felis virus		1		1		
326 3		687385	Torque teno canis virus		1		1		

326 4		687386	Torque teno sus virus 1a	12	1	12	1		
326 5		689403	Human bocavirus 1	8		8			
326 9		693999	Scotophilus bat coronavirus 512		1		1		
327 0		694003	Betacoronavirus 1	260		260			
327 2		694007	Tylonycteris bat coronavirus HKU4	2		2			
327 3		694008	Pipistrellus bat coronavirus HKU5	1	1	1	1		
327 4		696863	Sprivirus cyprinus		1		1		
327 5		696871	Vaccinia virus Western Reserve		31		31		
327 6		697298	Giant panda rotavirus A		1		1		
327 7		697610	Influenza A virus (A/India/GWMH05/2009(H1N1))		1		1		
327 8		703352	Brucella melitensis M5-90	5	26	5	26		
327 9		707424	Norovirus Hu/GII-4/Ehime4/2007/JP		1		1		
328 1		707488	Norovirus Hu/GII-4/Nagano3/2007/JP		4		4		
328 2		707882	Duck astrovirus 1	1		1			
328 3		708187	Colletotrichum chlorophyti		1		1		
328 4		714978	Human adenovirus 55	4		4			
328 5		715091	Influenza A virus (A/Roma/ISS1941/2009(H1N1))		1		1		
328 6		715494	Influenza A virus (A/Hong Kong/33982/2009(H9N2))		3		3		
328 7		722438	Mycoplasmoides pneumoniae FH	3		3			
328 8		742305	Influenza A virus (A/slaty-backed gull/Japan/6KS0185/2006(H4N8))		1		1		
328 9		742503	Tokudaia muenninki		1		1		
329 0		743722	Sphingobacterium sp. 21	1		1			
329 1		743973	Taylorella equigenitalis ATCC 35865	1		1			
329 2		746128	Aspergillus fumigatus	136	88	136	88		
329 3		747305	Norovirus GII.10	7		7			
329 4		754027	Treponema phagedenis F0421		1		1		
329 5		754366	Influenza A virus (A/Ontario/309862/2009(H1N1))	1		1			
329 6		754503	Mesomycoplasma hyopneumoniae 7422	1		1			
329 7		759272	Thermochaetoides thermophila DSM 1495	1		1			
329 8		759363	Chlamydia trachomatis D-EC	3		3			
329 9		759851	Sporosarcina newyorkensis		2		2		
330 0	X	760063	Influenza A virus (A/Puerto Rico/8-MC/1934(H1N1))				1		1
330 1		760591	Influenza A virus (A/Egypt/N03072/2010(H5N1))	1		1			
330 2		761193	Runella slithyformis DSM 19594		1		1		
330 3		761251	Influenza A virus (A/Mexico City/WR1704T/2009(H1N1))		1		1		
330 4		762377	Influenza A virus (A/bar-headed goose/Qinghai/1-HVRI/2006(H5N1))		5		5		

330 5		762378	Influenza A virus (A/bar-headed goose/Qinghai/3/2005(H5N1))		1		1		
330 7		763552	Mus musculus papillomavirus type 1		4		4		
330 8	X	766147	<i>Shigella flexneri</i> K-227			1		1	
331 0		768646	Influenza A virus (A/chicken/Uchal/8293/2006(H9N2))		1		1		
331 1		768647	Influenza A virus (A/chicken/Uchal/8286/2006(H9N2))		1		1		
331 2		857099	Streptococcus mutans OMZ175	2	1	2	1		
331 3		857571	Moraxella catarrhalis O35E	27		27			
331 4	X	861244	<i>Pueraria candollei</i> var. <i>mirifica</i>			1		1	
331 5		862909	Trichodysplasia spinulosa-associated polyomavirus	2		2			
331 6		863370	Meiothermus granaticus		1		1		
331 7		866344	Helicobacter pylori F16	2		2			
332 0		870484	Nonlabens agnitus		1		1		
332 1	X	871325	<i>Bacteroides faecichinchillae</i>				1		1
332 2		871575	Ogataea parapolyomorpha DL-1		1		1		
332 3		874269	African swine fever virus Georgia 2007/1	15	1	15	5		4
332 4		875328	Mycolicibacter sinensis		10		10		
332 6		885311	Entamoeba histolytica KU27		1		1		
332 7		887118	Influenza A virus (A/Mexico/UASLP-012/2008(H3N2))		1		1		
332 8		908203	Influenza A virus (A/Thailand/CU-H126/2009(H1N1))		1		1		
332 9		909420	Neisseria meningitidis H44/76	2		2			
333 0		913028	Yersinia enterocolitica W22703		1		1		
333 1	X	914127	<i>Vibrio vulnificus</i> MO6-24/O			1		1	
333 2		914129	Streptococcus pneumoniae 2061376		1		1		
333 3		915014	Influenza A virus (A/swine/Heilongjiang/44/2009(H1N1))	1		1			
333 4		928301	Fowlpox virus strain NVSL		3		3		
333 5		928303	Hendra virus horse/Australia/Hendra/1994	3		3			
333 6		928313	Tacaribe virus strain Franze-Fernandez		6		6		
333 7		929439	Leishmania mexicana MHOM/GT/2001/U1103		1		1		
333 8		932622	Influenza A virus (A/chicken/Huadong/4/2008(H5N1))	1		1			
333 9		935198	Clostridium botulinum B str. Eklund 17B (NRP)		1		1		
334 0		936115	Influenza A virus (A/Netherlands/18/1994(H3N2))		1		1		
334 1		940614	Granulicella mallensis		1		1		
334 2		945782	Synechocystis salina LEGE 06155		1		1		
334 3		947939	Influenza A virus (A/aquatic bird/Korea/w44/2005(H7N3))		1		1		
334 4		981087	Leishmania donovani BPK282A1		9		9		

3345		981676	Bovine viral diarrhea virus 1b		28		28		
3347	X	983929	Chaerephon bat coronavirus/Kenya/KY22/2006				1		1
3348		986174	Influenza A virus (A/swine/Korea/4382/2009(H1N2))		1		1		
3349		991335	Influenza A virus (A/swine/Hong Kong/71/2009(H1N1))		1		1		
3350		992121	Helicobacter pylori Hp M5	1		1			
3351		996581	Influenza A virus (A/chicken/Shanghai/Q0808-1/2008(H9N2))		1		1		
3352	X	1002900	Baiyangdian virus				12		12
3353	X	1002921	Porcine sapelovirus 1			1		1	
3354		1003835	Severe fever with thrombocytopenia syndrome virus	1		1			
3355		1004253	Cyberlindnera mrakii	1	1	1	1		
3356		1005048	Collimonas fungivorans Ter331		1		1		
3357		1005962	Ogataea parapolyomorpha		1		1		
3358		1006061	Duck hepatitis A virus 1	1		1			
3359		1006581	Mycoplasma gallisepticum S6	1		1			
3360		1009714	Pseudomonas aeruginosa PAK	7	3	7	3		
3361		1016852	Banana streak CA virus	5		5			
3362		1016853	Banana streak IM virus	10		10			
3363		1016998	Salmonella enterica subsp. enterica serovar Paratyphi B str. SPB7		11		11		
3364		1035514	Plasmodium vivax North Korean	1		1			
3365		1036723	Plasmodium falciparum Vietnam Oak-Knoll (FVO)		2		2		
3366		1041522	Mycobacterium colombiense CECT 3035		20		20		
3367		1041990	Influenza A virus (A/Guangdong/15/2007(H3N2))	3		3			
3368		1045010	Escherichia coli O157	1		1			
3369		1045775	Paenibacillus catalpae	1		1			
3370		1046049	Norovirus Hu/GII.12/HS206/2010/USA		1		1		
3371		1047168	Zymoseptoria brevis		1		1		
3372		1050221	Chlamydia psittaci NJ1	1		1			
3373		1053141	Recombinant Hepatitis C virus J6/JFH1-J6	2		2			
3374		1055687	Trypanosoma vivax Y486	1		1			
3375		1068705	Influenza A virus (A/swine/Guangdong/221/2009(H1N1))		1		1		
3378		1071379	Tetrapisispora blattae		1		1		
3379		1077587	Influenza B virus (B/Victoria/2/1987)	2	4	2	4		
3380		1081253	Influenza A virus (A/Indiana/08/2011(H3N2))		6		6		
3381		1082519	Influenza A virus (A/chicken/Taixing/10/2010(H9N2))	2		2			
3382		1086859	Influenza A virus (A/Bilthoven/4791/1981(H3N2))		3		3		

338 4		1091045	Cutibacterium acnes subsp. defensens ATCC 11828	5		5			
338 5		1093545	Influenza A virus (A/shorebird/Delaware Bay/277/2000(H9N7))		1		1		
338 7		1112244	Chlamydia psittaci 06-1683	3		3			
338 8		1112253	Chlamydia psittaci 02DC14	1		1			
338 9		1112254	Chlamydia psittaci 02DC15	19		19			
339 0		1112258	Chlamydia psittaci 02DC22	2		2			
339 1		1112262	Chlamydia psittaci 03DC29	1		1			
339 2		1112269	Chlamydia psittaci 09DC78	1		1			
339 3		1117322	Glaesserella parasuis str. Nagasaki	1		1			
339 4		1118369	Wobbly possum disease virus		14		14		
339 5		1122279	Influenza A virus (A/equine/Guangxi/3/2011(H9N2))		1		1		
339 6		1125722	Porphyromonas gingivalis W50		36		36		
339 7		1126212	Macrophomina phaseolina MS6		1		1		
339 8		1129345	Influenza A virus (A/little yellow-shouldered bat/Guatemala/153/2009(H17N10))		3		3		
340 0		1133319	Bacteroides reticulotermitis		1		1		
340 1		1133363	Schmallenberg virus	6		6			
340 2		1133968	Babesia microti strain RI	3		4		1	
340 3	X	1134687	Klebsiella michiganensis				1		1
340 4		1144025	Influenza A virus (A/Shandong/9/1993(H3N2))	13		13			
340 5		1146883	Blastococcus saxosidens DD2		1		1		
340 6		1156983	Norovirus Hu/GII.4/Minerva/CS1258/2006/USA	1		1			
340 7	X	1159556	Ustilaginoidea virens			7		7	
340 8	X	1159904	Night heron coronavirus HKU19				1		1
340 9	X	1159907	White-eye coronavirus HKU16				1		1
341 0	X	1159908	Wigeon coronavirus HKU20				1		1
341 2		1163697	Influenza A virus (A/Oklahoma/309/2006(H3N2))	4	3	4	3		
341 3		1169127	Clostridioides difficile genomovar ribotype_087	1		1			
341 4		1171336	Influenza A virus (A/swine/Nebraska/A01116984/2011(H3N2))		1		1		
341 5		1172206	Neisseria meningitidis NMB	1		1			
341 6		1173018	Phlebovirus WCH/97/HN/China/2011	19		19			
341 7		1173701	Colletotrichum sublineola		1		1		
341 8		1176042	Influenza A virus (A/swine/Italy/1513-1/1998(H1N1))		1		1		
341 9		1178016	Encephalitozoon romaleae SJ-2008		1		1		
342 0		1179777	Mycoplasma feriruminatoris		1		1		
342 1		1182143	Norovirus Hu/GII.4/Farmington Hills/2004/USA	2		2			

342 2		1182144	Norovirus Hu/GII.4/Minerva/2006/USA	2		2		
342 3		1184252	Streptococcus suis S735	2		2		
342 5		1184593	Influenza A virus (A/Victoria/210/2009(H3N2))	16		16		
342 6	X	1184594	Influenza A virus (A/Wisconsin/15/2009(H3N2))			1		1
342 7		1186058	Trichosporon asahii var. asahii CBS 2479		1		1	
342 8		1193095	Paucilactobacillus hokkaidonensis		1		1	
342 9		1193576	Staphylococcus aureus subsp. aureus CN1		3	1	3	1
343 0		1197131	Escherichia coli F576	1		1		
343 1		1198627	Mycobacteroides abscessus subsp. massiliense str. GO 06		1		1	
343 2		1198676	Streptococcus mutans GS-5		14		14	
343 3		1207470	Streptococcus pyogenes M1 476	1		1		
343 4		1209523	Toxoplasma gondii type II	11	2	11	2	
343 5		1209525	Toxoplasma gondii type I	10	2	10	2	
343 6		1209926	Colletotrichum orchidophilum		1		1	
343 7		1214154	Streptococcus suis 2651	1		1		
343 8		1214159	Streptococcus suis 13730	2		2		
343 9		1214225	Acidocella sp. MX-AZ02	1		1		
344 0		1218488	Torque teno sus virus k2	11		11		
344 1		1219377	rhinovirus C3		52		52	
344 2	X	1223514	Corynebacterium humireducens				11	11
344 3		1225860	Porcine torque teno virus 1	2		2		
344 4		1229831	Chlamydia avium 10DC88	2		2		
344 5		1235801	Ligilactobacillus murinus ASF361		2		2	
344 6		1235802	Eubacterium plexicaudatum ASF492		2		2	
344 7		1235996	Human betacoronavirus 2c EMC/2012	11	26	31	26	20
344 8		1236180	Halopelagius longus		1		1	
344 9		1236978	Staphylococcus epidermidis JCM 2414	1		1		
345 0		1239567	Mamastrovirus 3	2		2		
345 1		1241973	Norovirus Hu/GII.4/Sydney/NSW0514/2012/AU	8	31	8	31	
345 2		1254407	Norovirus Hu/GII.4/N3/2008/HuZhou		1		1	
345 4		1263102	Segatella copri CAG:164		1		1	
345 5		1263720	Betacoronavirus England 1	13		13		
345 6		1268303	Rhodococcus sp. AW25M09	1	1	1	1	
345 7		1268360	Influenza A virus (A/Victoria/361/2011(H3N2))	8		8		
345 8		1278073	Myxococcus stipitatus DSM 14675		1		1	
346 0		1281454	Rodent hepacivirus		12		12	

346 1		1285028	Influenza A virus (A/American black duck/New Brunswick/00464/2010(H4N6))	1		1			
346 2		1286616	Norovirus Hu/GII.3/693/425/2008/AU	6		6			
346 3		1286635	Desulfotignum phosphitoxidans DSM 13687		1		1		
346 4		1288825	Shigella flexneri 2b	2		2			
346 5		1289562	Influenza A virus (A/swine/Spain/SF11131/2007(H1N1))		3		3		
346 6		1291821	Norovirus Hu/GII/20144/2009/VNM		1		1		
346 7		1291870	Norovirus Hu/GII/20457/2010/VNM		1		1		
346 9		1303514	Diaporthe toxica	1		1			
347 0		1310532	Influenza A virus (A/chicken/West Java/Sbg-29/2007(H5N1))	7		7			
347 1		1314758	Influenza A virus (A/Hangzhou/1/2013(H7N9))		1		1		
347 2		1318439	Influenza A virus (A/Washington/05/2011(H1N1))	1		1			
347 3		1318464	Bovine Schmallenberg virus BH80/Germany/2011	2		2			
347 4	X	1320096	Norovirus Hu/GII.4/Huzhou121/2012/CHN			4		4	
347 5		1321009	Influenza A virus (A/Texas/50/2012(H3N2))	6		10		4	
347 6		1321999	Norovirus Hu/GII.6/Ehime120246/2012/JP	2		2			
347 8		1330520	Enterovirus F	1		1			
347 9		1331831	Influenza A virus (A/Taiwan/1/2013(H7N9))	2		2			
348 0		1332244	Influenza A virus (A/Shanghai/02/2013(H7N9))	66		67		1	
348 2		1335626	Middle East respiratory syndrome-related coronavirus	1563 6	26	1564 2	29	6	3
348 3		1337063	Puumala virus bank vole/CG1820/Russia/1984		1		1		
348 4		1337393	Helicobacter pylori PZ5056	1		1			
348 5		1342397	Influenza A virus (A/chicken/Hong Kong/TP38/2003(H9N2))		3		3		
348 6		1342434	Influenza A virus (A/turkey/Minnesota/511/1978(H9N2))		1		1		
348 7		1345266	HIV-1 M:CRF01_AE	1		1			
348 8		1346336	Influenza A virus (A/Fort Monmouth/1-JY2/1947(H1N1))		1		1		
348 9	X	1349999	Goose astrovirus			4		4	
349 0		1350216	Norovirus Hu/GII.12/Texas/E13842/USA/2000		1		1		
349 1		1352357	Helicobacter pylori SouthAfrica50	1		1			
349 2		1355477	Bradyrhizobium diazoefficiens		1		1		
349 3		1380386	Mycobacterium sp. URHB0044		1		1		
349 5		1384672	Mumps virus genotype G		83		83		
349 7		1389959	Mycobacterium tuberculosis variant bovis AN5	9	28	9	28		
349 8		1391655	Influenza A virus (A/Aichi/2-1/1968(H3N2))	1		1			

349 9		1391998	Mycobacterium avium subsp. paratuberculosis 08-8281	1		1			
350 0		1392869	Escherichia coli K1		1		1		
350 2		1395982	Influenza A virus (A/tree sparrow/Shanghai/01/2013(H7N9))	1		1			
350 3		1398154	Sporothrix brasiliensis 5110		4		4		
350 4		1399582	Duck Tembusu virus	8		8			
350 5		1401444	Avian leukosis virus ev/J	6		6			
350 6		1403335	Porphyromonas gingivalis 381	71	55	71	55		
350 7		1403514	Influenza A virus (A/chicken/Jiangsu/x1/2004(H9N2))	8		8			
350 8		1405296	Chlamydia suis MD56	1		1			
350 9		1406148	Norovirus Hu/GII.3/Jingzhou/2013402/CHN	2		2			
351 1		1408475	Taylorella asinigenitalis ATCC 700933	1		1			
351 2		1413188	Influenza A virus (A/Anhui/PA-1/2013(H7N9))	1	3	1	3		
351 3	X	1415851	Bat SARS-like coronavirus RsSHC014			1		1	
351 4	X	1415852	Bat SARS-like coronavirus WIV1			1		1	
351 5		1416333	Trypanosoma cruzi Dm28c	2		2			
351 6		1420840	Influenza A virus (A/Luxembourg/46/2009(H1N1))		1		1		
351 7		1423732	Lactiseibacillus casei DSM 20011 = JCM 1134 = ATCC 393	2		2			
351 8		1427371	Chlamydia pecorum VR629	5		5			
351 9		1432052	Eisenbergiella tayi		3		3		
352 0		1432555	Escherichia coli ISC7		1		1		
352 1		1440122	Murid herpesvirus 68		2		2		
352 2		1447813	Influenza A virus (A/Shanghai/5190T/2013(H7N9))		6		6		
352 3		1451093	Influenza A virus (A/chicken/Taiwan/67/2013(H6N1))		5		5		
352 4		1457141	Chlamydia avium	1		1			
352 5		1457153	Chlamydia gallinacea	3		3			
352 6		1458279	Staphylococcus aureus USA300-ISMM51	1		1			
352 7		1458555	Influenza A virus (A/Jiangxi/IPB13/2013(H10N8))	1		1			
352 8		1463841	Streptomyces sp. NRRL F-2580		1		1		
352 9		1464048	Micromonospora parva		1		1		
353 0		1470614	Influenza A virus (A/swine/Thailand/CU-S3673N/2012(H3N2))		1		1		
353 1	X	1472417	Butyrlicimonas paravirosa				1		1
353 2		1472713	Norovirus Hu/GII/Amsterdam/1994		1		1		
353 3		1476909	Banana streak MY virus	39		39			
353 4		1481964	Influenza A virus (A/Anhui/1-DEWH730/2013(H7N9))	1	35	1	35		
353 5		1481966	Influenza A virus (A/Anhui/1-JCVI1_RG2/2013(H7N9))	3		3			

353 6		1481987	Influenza A virus (A/Anhui/1-BALF_RG44/2013(H7N9))	1		1			
353 8		1498499	Legionella norrandica		1		1		
353 9		1498657	Norovirus Hu/GII.4/Chiba5/2010/JP		2		2		
354 0		1501332	Oribacterium asaccharolyticum		1		1		
354 1		1507381	Influenza A virus (A/Minnesota/11/2010(H3N2))	19		19			
354 3		1507390	Influenza A virus (A/reassortant/NYMC X-217A(A/Puerto Rico/8/1934 x A/Victoria/361/2011)(H3N2))	1		1			
354 5		1508227	Bat SARS-like coronavirus	646		646			
354 8		1562038	Norway rat hepacivirus 1		5		5		
355 0		1566298	Paraiso Escondido virus		1		1		
355 1		1570291	Ebola virus	3	142	3	142		
355 2	X	1577715	Norovirus GVI.1/Ca/JPN/2012/M49-1			3		3	
355 3		1581419	Amblyomma sculptum	1		1			
355 4		1586324	Porcine deltacoronavirus	3		5		2	
355 5		1590370	Betacoronavirus HKU24		3		3		
355 6		1631247	Mesorhizobium delmotii		1		1		
355 7		1634342	Norovirus Hu/GII/JP/2015/GII.P17_GII.17/Kawasaki308	1		1			
355 8	X	1654135	Influenza A virus (A/swine/England/1353/2009(H1N1))				3		3
355 9		1663864	Influenza A virus (A/Anhui/1-YK_RG134/2013(H7N9))		1		1		
356 0		1663934	Influenza A virus (A/Anhui/1-YK_RG64/2013(H7N9))		2		2		
356 1		1663996	Influenza A virus (A/Anhui/1-YK_RG01/2013(H7N9))	1		1			
356 2		1665645	Norovirus GI/Hu/NL/2012/GI.6/Groningen		1		1		
356 3		1671798	Human papillomavirus type 54		1		1		
356 4		1678143	Paslahepevirus balayani	153	91	153	91		
356 5		1678144	Avihepevirus magniecur	20		20			
356 6		1727172	Influenza A virus (A/duck/Bavaria/1/1977(H1N1))		2		2		
356 7		1737357	Rhizobium wenxiniae		1		1		
356 8	X	1739614	229E-related bat coronavirus				1		1
356 9		1763596	Guertu virus	32		32			
357 0		1777792	Influenza A virus (A/Michigan/45/2015(H1N1))	1		6		5	
357 1		1783360	Comamonas sediminis		1		1		
357 3	X	1796646	Muribaculum intestinale				1		1
357 4	X	1803956	Parechovirus A			1		1	
357 5	X	1812858	Anaerosacchariphilus polymeriproducens				1		1
357 6		1835656	Rotavirus A RVA/Cow-tc/USA/B223/1983/G10P[11]	2		2			

357 7	X	1849603	Enterobacteriaceae bacterium				1		1
357 8		1855372	bacterium JGI 053		1		1		
357 9		1868221	Porcine circovirus 3	4		4			
358 0		1868482	Carlito syrichta	1		1			
358 1		1885248	Variiegated squirrel bornavirus 1		1		1		
358 2		1891187	Zaire ebolavirus Makona	55	57	59	57	4	
358 4		1891729	Alphapolyomavirus mauratus	11		11			
358 5		1891730	Alphapolyomavirus muris	4	37	4	37		
358 6		1891762	Betapolyomavirus hominis	4	81	4	81		
358 7		1891767	Betapolyomavirus macacae	16	42	16	42		
358 8		1898104	Bacteroidota bacterium	1	1	1	1		
358 9		1898112	Rhodospirillaceae bacterium		1		1		
359 0		1898203	Lachnospiraceae bacterium		2		2		
359 1	X	1913988	Alphaproteobacteria bacterium				1		1
359 2		1914447	Atypical porcine pestivirus	3		3			
359 3	X	1920748	NL63-related bat coronavirus				1		1
359 6		1965067	Porcine reproductive and respiratory syndrome virus 2		41		41		
359 7		1968303	Influenza A virus (A/chicken/Guangdong/GD15/2016(H7N9))	2		2			
359 9		1972699	Norovirus Hu/GII.3/3-34/2015/HNZZ/CHN	11		11			
360 0		1977402	Inovirus M13		1		1		
360 1		1978231	Acidobacteriota bacterium		1		1		
360 2		1980442	Orthohantavirus		13		13		
360 3		1980456	Orthohantavirus andesense	2	79	2	79		
360 4	X	2015173	Ooceraea biroi			1		1	
360 5		2016460	Giant panda anellovirus		1		1		
360 6		2016461	Giant panda associated gemycircularvirus		1		1		
360 7		2016463	Giant panda polyomavirus		1		1		
360 9		2019571	Influenza A virus (A/reassortant/IDCDC-RG56B(Hong Kong/125/2017 X Puerto Rico/8/1934)(H7N9))	2		2			
361 0		2024894	Acidimicrobiaceae bacterium		1		1		
361 2		2026033	Influenza A virus (A/chicken/Hunan/S12753/2016(H7N9))	2		2			
361 3	X	2026800	Verrucomicrobiaceae bacterium				1		1
361 4		2029108	Bacillus sp. UMB0899	1		1			
361 5		2030927	Bacteroidales bacterium		1		8		7
361 6	X	2032731	Swine acute diarrhea syndrome coronavirus			2		2	

361 7		2043570	Zika virus ZIKV/H. sapiens/FrenchPolynesia/10087PF/2013	26	75	26	75		
361 8		2044587	Schaederella arabinosiphila		5		5		
361 9		2044939	Clostridia bacterium		2		3		1
362 0		2049047	Prevotellaceae bacterium		1		2		1
362 2		2066070	Mesorhizobium japonicum		1		1		
362 3		2070132	Influenza A virus (A/chicken/Jiangsu/W1-8/2015(H7N9))	6		6			
362 5		2115988	Swine acute diarrhea syndrome related coronavirus	1		1			
362 6		2116544	Synechococcus lacustris	1		1			
362 7	X	2136182	Acinetobacter cumulans				4		4
362 8		2169971	Visna-maedi virus	5	7	5	7		
362 9		2169986	Barley yellow dwarf virus PAV	1		1			
363 0		2169991	Mammarenavirus juninense	8	873	8	873		
363 1		2170197	Guenon simian foamy virus	1		1			
363 2		2170200	Spider monkey simian foamy virus	1		1			
363 3		2170201	Squirrel monkey simian foamy virus	1		1			
363 4		2170206	Yellow-breasted capuchin simian foamy virus	1		1			
363 5	X	2212467	Bacteroidaceae bacterium				1		1
363 6	X	2233538	Duck adenovirus 3			6		6	
363 7		2282107	Pyrrhoderma noxium		1		1		
363 8		2316109	Zika virus ZIKV/Human/Cambodia/FSS13025/2010		2		2		
363 9		2478486	Holophagales bacterium		1		1		
364 0	X	2485925	Oscillospiraceae bacterium				1		1
364 1		2487134	Clostridium sp. E02		1		1		
364 2		2492960	Bacillus yapensis		1		1		
364 3		2501420	Yak coronavirus		1		1		
364 4		2502994	Fusarium odoratissimum		1		1		
364 5		2560319	Avian orthoavulavirus 1	26	5	26	5		
364 6		2560525	Human orthorubulavirus 2	10		10			
364 7		2560580	Mammalian orthorubulavirus 5	1	1	1	1		
364 8		2560602	Mumps orthorubulavirus	3	22	3	23		1
364 9	X	2584949	Enterobacter sp. DE0047				1		1
365 0		2593991	Peste des petits ruminants virus		1		1		
365 1		2594813	Fusarium fasciculatum		1		1		
365 2		2599941	Oligoflexia bacterium		1		1		
365 3		2678873	Trimerodytes annularis	1		1			
365 4		2697049	Severe acute respiratory syndrome coronavirus 2	7631	3166	8589	8883	958	5717

365 5		2707005	Paenibacillus lutimimalis		1		1		
365 6		2708335	Pangolin coronavirus	1782	1	1782	1		
365 7		2709072	Bat coronavirus RaTG13	687		688		1	
365 8		2758382	Bos taurus papillomavirus 2	15		15			
365 9		2764122	hepatitis B virus genotype C	21	29	21	31		2
366 0	X	2770637	Aestuariatibaculum sediminum				1		1
366 2		2847087	GB virus-B		4		4		
366 3		2847137	hepatitis B virus genotype D	1	9	1	12		3
366 4		2847144	hepatitis C virus genotype 1a	216	398	220	398	4	
366 7	X	2900548	Actinomycetota bacterium				1		1
366 8		2978683	Burkholderia orbicola		7		7		
366 9		3042615	Paraglaciecola sp. T6c		1		1		
367 0		3052223	Henipavirus hendraense	9	2	12	2	3	
367 1		3052225	Henipavirus nipahense	7	11	8	11	1	
367 2		3052230	Hepacivirus hominis	720	1155	721	1156	1	1
367 3		3052296	Mammarenavirus amapariense		3		3		
367 4		3052299	Mammarenavirus braziliense		879		879		
367 5		3052300	Mammarenavirus caliense		11		11		
367 6		3052303	Mammarenavirus choriomeningitidis	5	655	5	655		
367 7		3052304	Mammarenavirus cupixiense		1		1		
367 8		3052307	Mammarenavirus guanaritoense		842		842		
367 9		3052310	Mammarenavirus lassaense	15	707	15	707		
368 0		3052317	Mammarenavirus machupoense	1	801	1	801		
368 1		3052320	Mammarenavirus mopeiaense		14		14		
368 2		3052322	Mammarenavirus oliverosense		2		2		
368 3		3052323	Mammarenavirus paranaense		1		1		
368 4		3052325	Mammarenavirus praomyidis	1		1			
368 5		3052328	Mammarenavirus tacaribeense		106		106		
368 6		3052331	Mammarenavirus whitewaterense		349		349		
368 7		3052342	Morbillivirus canis	14	14	14	14		
368 8		3052343	Morbillivirus caprinae	13	31	13	31		
368 9		3052470	Orthohantavirus bayoui	1		1			
369 0		3052476	Orthohantavirus delgaditoense		1		1		
369 1		3052477	Orthohantavirus dobravaense		25		25		
369 2		3052480	Orthohantavirus hantanense	4	69	4	69		
369 3		3052483	Orthohantavirus khabarovskense		1		1		
369 4		3052486	Orthohantavirus montanoense	3		3			
369 5		3052487	Orthohantavirus moroense		2		2		
369 6		3052489	Orthohantavirus negraense		12		12		

369 7	3052492	Orthohantavirus prospectense	1	2	1	2		
369 8	3052493	Orthohantavirus puumalaense	33	20	33	20		
369 9	3052496	Orthohantavirus sangassouense		6		6		
370 0	3052498	Orthohantavirus seoulense		5		5		
370 1	3052499	Orthohantavirus sinnombreense	1	35	1	35		
370 2	3052503	Orthohantavirus tulaense	1	14	1	14		
370 3	3052505	Orthomareburgvirus marburgense	6	104	6	104		
370 4	3052518	Orthonairovirus haemorrhagiae	95	18	95	18		
370 5	3052562	Orthorubulavirus suis	3		3			
370 6	3052571	Orthospovirus citrullomaculosi	3		3			
370 7	3052585	Orthospovirus tomatomaculae	1		1			
370 8	3052729	Respirovirus bovis	3	1	3	1		
370 9	3052731	Respirovirus muris	63	10	63	10		
371 0	10000000	Mus musculus BALB/c		3		3		
371 1	10000055	Mus musculus DBA/2	1		1			
371 2	10000067	Mus musculus C57BL/6	1	381	1	4004		3623
371 3	10000187	Sus scrofa Landrace X Large White	1		1			
371 4	10000225	Mus musculus C57BL/6N		1		1		
371 5	10000291	Bacillus anthracis str. Sterne 34F2	1		1			
371 6	10000293	Clostridium botulinum A 1	1		1			
371 7	10000294	Clostridium botulinum A 2	2		2			
371 8	10000295	Clostridium botulinum B 111	2		2			
371 9	10000296	Clostridium botulinum D 1873	2		2			
372 0	10000297	Clostridium botulinum C 92-13	5		5			
372 1	10000299	Clostridium botulinum E Beluga	2		2			
372 2	10000301	Clostridium botulinum A str. Hall hyper	3		3			
372 3	10000302	Clostridium botulinum A Kyoto-F	1		1			
372 4	10000303	Clostridium botulinum B Lammana	1		1			
372 5	10000304	Clostridium botulinum F NCTC 10281	1		1			
372 6	10000305	Clostridium botulinum B Okra	3		3			
372 7	10000306	Clostridium botulinum C Stockholm	2		2			
372 8	10000307	Clostridium difficile BART'S W1	1		1			
372 9	10000308	Listeria monocytogenes ATCC 35967		1		1		
373 0	10000309	Listeria monocytogenes ATCC 43251		3		3		
373 1	10000313	Mycobacterium avium serovar 1	1		1			
373 2	10000314	Mycobacterium avium serovar 2	1		1			
373 3	10000316	Mycobacterium avium serovar 25	1		1			
373 4	10000317	Mycobacterium avium serovar 26	1		1			

373 5	10000318	Mycobacterium avium serovar 4	2		2			
373 6	10000319	Mycobacterium avium serovar 8	1	1	1	1		
373 7	10000320	Mycobacterium avium serovar 9	1		1			
373 8	10000323	Mycobacterium bovis T/91/1378		6		6		
373 9	10000328	Mycobacterium avium subsp. paratuberculosis Strain Ben		1		1		
374 0	10000329	Mycobacterium tuberculosis 103	1		1			
374 1	10000331	Mycobacterium gastris W471	1		1			
374 2	10000335	Candida albicans A-9 (serotype B)	1		1			
374 3	10000337	Candida albicans KIT 1113	1		1			
374 4	10000339	Candida albicans serotype A	1		1			
374 5	10000341	Leishmania donovani donovani 1S2D	1		1			
374 6	10000345	Leishmania infantum LEM 75	14		14			
374 7	10000348	Trypanosoma cruzi G	1		1			
374 8	10000351	Trypanosoma cruzi Y	1		1			
374 9	10000352	Entamoeba histolytica YS-27	1		1			
375 0	10000353	Toxoplasma gondii 76K	5	5	5	5		
375 1	10000354	Toxoplasma gondii BK	1	1	1	1		
375 2	10000355	Toxoplasma gondii Prugniaud		1		1		
375 3	10000356	Plasmodium berghei NK65	2	1	2	1		
375 4	10000357	Plasmodium chabaudi adami DS	2	37	2	37		
375 5	10000358	Plasmodium falciparum 366		1		1		
375 9	10000363	Plasmodium falciparum Brazil-608	1	1	1	1		
376 0	10000366	Plasmodium falciparum FCB-2	1		1			
376 1	10000369	Plasmodium falciparum FVO		5		5		
376 2	10000370	Plasmodium falciparum GAM5		1		1		
376 3	10000371	Plasmodium falciparum Indochina I/CDC	32		32			
376 4	10000373	Plasmodium falciparum ItG2G1		1		1		
376 5	10000375	Plasmodium falciparum RO71	1	1	1	1		
376 6	10000376	Plasmodium falciparum UF-5	2		2			
376 7	10000378	Plasmodium vivax NK		2		2		
376 8	10000381	Plasmodium yoelii yoelii YM	1	5	1	5		
376 9	10000382	Babesia bovis Mexico	3	4	3	4		
377 0	10000383	Babesia bovis Mexico Mo7	1	33	1	33		
377 1	10000385	Schistosoma mansoni Puerto Rico	20	7	20	7		
377 2	10000386	Monkeypox virus USA_2003_039		1		1		
377 3	10000388	Vaccinia virus NYCBH - Dryvax		29		29		
377 4	10000390	Variola major virus India-1967		1		1		
377 5	10000391	Equid herpesvirus 2 ER32	1		1			

377 6	10000394	Human herpesvirus 1 103/65	1		1			
377 7	10000396	Human herpesvirus 1 McIntyre		1		1		
377 8	10000398	Human herpesvirus 1 NS	1		1			
377 9	10000404	Bovine herpesvirus 1 Lam	1		1			
378 0	10000405	Equid herpesvirus 4 TH20	2		2			
378 1	10000406	Human herpesvirus 3 H-551	26	15	26	15		
378 2	10000408	Human herpesvirus 5 TB40		1		1		
378 3	10000411	Murid herpesvirus 1 deltaMS94.5		2		2		
378 4	10000412	Murid herpesvirus 1 Isolate G4		1		1		
378 5	10000413	Murid herpesvirus 1 Isolate K6		1		1		
378 7	10000420	Human herpesvirus 4 BL74		1		1		
378 8	10000421	Human herpesvirus 4 CKL		10		10		
378 9	10000424	Human herpesvirus 4 GD1	4	1	4	1		
379 0	10000427	Human herpesvirus 4 type A		5		5		
379 2	10000431	Hepatitis B virus ay/ad	4		4			
379 3	10000432	Hepatitis B virus genotype A	1		1			
379 4	10000436	Hepatitis B virus subtype AY	7		7			
379 5	10000437	Hepatitis B virus subtype AYR		1		1		
379 6	10000438	Human parvovirus B19 genotype 1	1		1			
379 7	10000439	Eastern equine encephalitis virus SV	8		8			
379 8	10000440	Dengue virus type 1 Hawaii	29	22	29	22		
379 9	10000442	Dengue virus type 3 CH53489		7		7		
380 0	10000444	Japanese encephalitis virus CH2195LA	1		1			
380 1	10000445	Japanese encephalitis virus JaOH0566	1		1			
380 2	10000447	West Nile virus 3000.0259	2		2			
380 3	10000449	Tick-borne encephalitis virus (WESTERN SUBTYPE) - Neudoerfl	10		10			
380 4	10000451	Classical swine fever virus Shimen	19		21		2	
380 5	10000453	Hepatitis C virus subtype 1a (isolate Gla)	5		5			
380 6	10000455	Hepatitis C virus subtype 1a 1/910		18		18		
380 7	10000456	Hepatitis C virus subtype 1b AD78	71		71			
380 8	10000457	Hepatitis C virus subtype 1a Chiron Corp.	1	1	1	1		
381 0	10000460	Hepatitis C virus subtype 1b JK1	13		13			
381 1	10000462	Measles virus CAM/RB		1		1		
381 2	10000463	Phocine distemper virus 2558/Han 88	5		5			
381 3	10000465	Rinderpest virus LATC	2		2			
381 4	10000467	Rabies virus CVS	14		14			
381 5	10000470	Rabies virus Flury LEP		1		1		

3816	10000471	Rabies virus RC-HL	4		4			
3817	10000472	Bovine ephemeral fever virus BB7721	2		2			
3818	10000474	Influenza A virus H3N2 (A/Resvir-9 (H3N2))		10		10		
3819	10000477	H1N1 subtype Influenza A virus (A/Netherlands/306/00 (H1N1))		1		1		
3820	10000482	Rift Valley fever virus ZH501	2	42	2	42		
3821	10000483	Puumala virus CG18-20	4		4			
3822	10000484	Puumala virus Kazan	26	7	26	7		
3823	10000485	Puumala virus (STRAIN HALLNAS B1) Vranica/Hallnas	4		4			
3825	10000487	Lymphocytic choriomeningitis virus (strain Armstrong) (clone 4)	1		1			
3826	10000488	Lymphocytic choriomeningitis virus (strain Armstrong) (clone 53b)		61		61		
3827	10000490	Lymphocytic choriomeningitis virus A22.2b		1		1		
3828	10000491	Lymphocytic choriomeningitis virus Docile		2		2		
3829	10000494	Lymphocytic choriomeningitis virus (strain WE) variant 8.7		1		1		
3830	10000495	Lymphocytic choriomeningitis virus (strain WE) WE CL1.2		1		1		
3831	10000496	Lymphocytic choriomeningitis virus (strain Armstrong) (clone 3)		1		1		
3832	10000497	Lymphocytic choriomeningitis virus (strain Armstrong) (clone 5)	1		1			
3833	10000499	Equine infectious anemia virus PV		2		2		
3834	10000500	Human immunodeficiency virus 1 IIIB	1	2	1	2		
3835	10000501	Simian immunodeficiency virus - mac - mac 239	1	474	1	474		
3836	10000502	Simian immunodeficiency virus - mac - mac 32H		9		9		
3837	10000503	Simian immunodeficiency virus - mac - mac BK28		4		4		
3838	10000504	Simian immunodeficiency virus - mac - mac F965		2		2		
3840	10000506	Simian immunodeficiency virus - sm - sm PT573		3		3		
3841	10000507	Rabbit hemorrhagic disease virus Olot/89	1		1			
3842	10000513	Foot-and-mouth disease virus (strain O1) (O1 Kaufbeuren)	43	8	43	8		
3843	10000514	Foot-and-mouth disease virus (strain O1) (O1BFS 1860)	9		9			
3844	10000515	Foot-and-mouth disease virus (strain O1) (O1BFS)	5		5			
3845	10000516	Foot-and-mouth disease virus (strain O1) (O1 Campos)	18	19	18	19		
3846	10000518	Borna disease virus Giessen strain He/80	6		6			
3847	10000519	Hepatitis E virus China Xinjiang	1		1			
3848	10000520	Hepatitis E virus SAR-55	2	6	2	6		
3849	10000523	Hepatitis delta virus TW2667		5		5		
3850	10000524	Equid herpesvirus 2 691	1		1			
3851	10000525	Equid herpesvirus 2 16V	1		1			

385 2	10000526	Equid herpesvirus 2 5FN	1		1			
385 3	10000527	Equid herpesvirus 2 FIN60	1		1			
385 4	10000528	Porcine respiratory and reproductive syndrome virus BJ-4	2		2			
385 5	10000529	Porcine respiratory and reproductive syndrome virus CH-1a	7	1	7	1		
385 6	10000530	Borrelia garinii IP90	3		3			
385 7	10000532	Peste-des-petits-ruminants virus Sungri-96		11		11		
385 8	10000535	Human herpesvirus 6B HST	1		1			
385 9	10000536	Murid herpesvirus 4 G2.4		3		3		
386 0	10000537	Murid herpesvirus 4 WUMS		19		19		
386 1	10000542	Streptococcus pyogenes serotype M3 D58	4		4			
386 2	10000544	Sin Nombre virus NM H10	4		4			
386 3	10000547	Human enterovirus 71 Subgenogroup B5	12		12			
386 4	10000548	Human enterovirus 71 Subgenogroup C2	19		19			
386 5	10000549	Human enterovirus 71 Subgenogroup C4	12		12			
386 6	10000550	Influenza A virus H3N2 (A/Netherlands/9/03 (H3N2))		1		1		
386 7	10000553	Andes virus CHI-7913	53		53			
386 8	10000554	Salmonella enterica subsp. enterica serovar Minnesota R595	2		2			
386 9	10000555	Plasmodium yoelii yoelii 265BY		1		1		
387 0	10000556	Foot-and-mouth disease virus (strain O1) Kaufbeuren	44	12	44	12		
387 1	10000559	Chlamydomyxa abortus B-577	11		11			
387 2	10000560	Norovirus genogroup 3 Bo/Jena/1980/DE	1		1			
387 3	10000566	Paracoccidioides brasiliensis B339		8		8		
387 4	10000567	Vibrio cholerae O1 serotype Inaba	1		1			
387 5	10000568	Vibrio cholerae O1 serotype Ogawa	6		6			
387 6	10000571	Cowpox virus (Brighton Red) White-pock		1		1		
387 7	10000632	Mus musculus NOD		578		578		
387 8	10000662	Rattus norvegicus Lewis		1		1		
387 9	10000675	Borrelia burgdorferi CA12		6		6		
388 0	10000679	Campylobacter jejuni subsp. jejuni 16971.94GSH (O:41)	2		2			
388 1	10000718	Helicobacter pylori J223	2		2			
388 2	10000720	Helicobacter pylori UA948	2		2			
388 3	10000721	Helicobacter pylori UA955	1		1			
388 4	10000723	Pseudomonas aeruginosa Immunotype 4	4		4			
388 5	10000727	Escherichia coli 180/C3	1		1			
388 6	10000728	Escherichia coli B B/r CM6		1		1		
388 7	10000733	Escherichia coli O5:K4:H4	1		1			

388 8	10000734	Escherichia coli O65:K:-H-	1		1			
388 9	10000738	Salmonella typhimurium PL5 (O9,12)	1		1			
389 0	10000739	Salmonella typhi 620Ty	1		1			
389 1	10000740	Salmonella typhi Ty21a		6		6		
389 2	10000742	Salmonella typhimurium SH 4809	4		4			
389 3	10000743	Salmonella typhimurium SL3261		4		4		
389 4	10000748	Shigella dysenteriae serotype 1	3		3			
389 5	10000749	Shigella dysenteriae serotype 1 114Sd	1		1			
389 6	10000752	Shigella flexneri 5b	1		1			
389 7	10000754	Shigella flexneri X	1		1			
389 8	10000756	Yersinia pestis KIM 5	21		21			
389 9	10000757	Yersinia pestis 195/P	5		5			
390 0	10000759	Pasteurella multocida X-73	2		2			
390 1	10000760	Anaplasma marginale South Idaho		2		2		
390 2	10000761	Orientia tsutsugamushi Karp	1		1			
390 3	10000763	Chlamydia trachomatis Serovar C	16	1	16	1		
390 4	10000764	Chlamydia trachomatis Serovar E	19	2	19	2		
390 5	10000765	Chlamydia trachomatis Serovar H	7		7			
390 6	10000766	Chlamydia trachomatis Serovar I	10		10			
390 7	10000767	Chlamydia trachomatis serovar K	14		14			
390 8	10000768	Chlamydia trachomatis Serovar L2	15	12	15	12		
390 9	10000769	Chlamydia trachomatis Serovar L3	2		2			
391 0	10000773	Streptococcus mutans MT 8148	40	12	40	12		
391 1	10000775	Streptococcus pyogenes 156	1		1			
391 2	10000776	Streptococcus pyogenes 88/25	1		1			
391 3	10000777	Streptococcus pyogenes 88/30	1		1			
391 4	10000778	Streptococcus pyogenes 88/544	1		1			
391 5	10000779	Streptococcus pyogenes 90/85	1		1			
391 6	10000780	Streptococcus pyogenes serotype M12 A374	1		1			
391 7	10000781	Streptococcus pyogenes BSA10	3		3			
391 8	10000782	Streptococcus pyogenes NS1	1		1			
391 9	10000783	Streptococcus pyogenes NS14	1		1			
392 0	10000784	Streptococcus pyogenes NS27	1		1			
392 1	10000785	Streptococcus pyogenes NS5	1		1			
392 2	10000786	Streptococcus pyogenes serotype M11	2		2			
392 3	10000787	Streptococcus pyogenes serotype M13	1		1			
392 4	10000788	Streptococcus pyogenes serotype M22	1		1			

3925	10000789	Streptococcus pyogenes serotype M24	6	2	6	2		
3926	10000790	Streptococcus pyogenes serotype M30	1		1			
3927	10000791	Streptococcus pyogenes serotype M41	1		1			
3928	10000792	Streptococcus pyogenes serotype M52	1		1			
3929	10000793	Streptococcus pyogenes serotype M54	1		1			
3930	10000794	Streptococcus pyogenes serotype M55	1		1			
3931	10000795	Streptococcus pyogenes serotype M57	1		1			
3932	10000796	Streptococcus pyogenes serotype M60	1		1			
3933	10000797	Streptococcus pyogenes serotype M75	1		1			
3934	10000798	Streptococcus pyogenes serotype M8	1		1			
3935	10000801	Foot-and-mouth disease virus - type O (O/SKR/2002)	1		1			
3936	10000802	Taenia crassiceps Strain ORF	3	3	3	3		
3937	10000804	Chlamydia trachomatis Serovar B	47	5	47	5		
3938	10000806	Foot-and-mouth disease virus (strain O1) (O1 Manisa)	6	1	6	1		
3939	10000807	Haemophilus influenzae NTHi UC19	2	2	2	2		
3940	10000809	Swine vesicular disease virus ITL/1/66	1		1			
3941	10000814	Leptospira interrogans serovar Lai str. HY-1	1		1			
3942	10000815	Pseudomonas aeruginosa PAO	2	2	2	2		
3943	10000816	Plasmodium falciparum T9/96	1	1	1	1		
3944	10000817	Pseudomonas aeruginosa KB7	2		2			
3945	10000818	Pseudomonas aeruginosa P1	1		1			
3946	10000820	Foot-and-mouth disease virus (strain O1) (O/Taiwan/1/97)	1		1			
3947	10000822	Pseudomonas aeruginosa Immunotype 3	1		1			
3948	10000824	Streptococcus pyogenes serotype M6 strain D471	4		4			
3949	10000825	Infectious bronchitis virus Avian strain D207	11		11			
3950	10000828	Ajellomyces dermatitidis ATCC 60636		2		2		
3951	10000829	Norovirus genogroup 2 Mexico type strain 36	3		3			
3952	10000832	Norovirus genogroup 2 Camberwell 1890	1		1			
3953	10000833	Haemophilus influenzae NTHi 1128	6		6			
3954	10000835	Equine rhinitis A virus 393/76	4		4			
3955	10000836	Foot-and-mouth disease virus C1 CS8	23	24	23	24		
3956	10000840	Foot-and-mouth disease virus C1 Brescia It/64	2		2			
3957	10000842	Swine vesicular disease virus NET/1/92	1		1			
3958	10000843	Neisseria meningitidis serogroup B H44/76	74	22	74	22		

3959	10000845	Chlamydia trachomatis Serovar F	3		3		
3960	10000848	Theileria sergenti Type B1		1		1	
3961	10000849	Theileria sergenti Type B2		2		2	
3962	10000850	Theileria sergenti Type C		5		5	
3963	10000851	Theileria sergenti Type I		1		1	
3964	10000852	Chlamydia pneumoniae Kajaani 6		7		7	
3965	10000853	Chlamydia trachomatis Serovar L1	21	3	21	3	
3966	10000855	Theiler's murine encephalomyelitis virus (strain BeAn 8386) (variant M2)		1		1	
3967	10000856	Foot-and-mouth disease virus - type O isolate O/UKG/35/2001		14		14	
3969	10000858	Chlamydia trachomatis B/Jali-20/OT	2		2		
3970	10000859	Porcine respiratory and reproductive syndrome virus JA142	1		1		
3971	10000860	Haemophilus influenzae Serotype B	18	9	18	9	
3972	10000861	Haemophilus influenzae Variant d1	18		18		
3973	10000862	Plasmodium vivax VK247	1		2		1
3974	10000863	Neisseria gonorrhoeae 4505	1		1		
3975	10000864	Porcine respiratory and reproductive syndrome virus MD-001	2		2		
3976	10000865	Influenza A virus H3N2 (A/Kiev/301/94)	3		3		
3977	10000888	Porcine respiratory and reproductive syndrome virus Olot/91	3	5	3	5	
3978	10000890	Porcine respiratory and reproductive syndrome virus SD92-23983		33		33	
3979	10000960	Human respiratory syncytial virus A Mon/3/88	6		6		
3980	10000961	Dengue virus type 1 strain 16007	15		15		
3981	10000964	Foot-and-mouth disease virus (strain O1) (O1 Brugge)	2		2		
3982	10000965	Dengue virus type 1 FGA/89	4		4		
3983	10000968	Hepatitis C virus subtype 1b isolate BE-11	3		3		
3984	10000969	Porcine respiratory and reproductive syndrome virus 111/92	9		9		
3985	10000971	West Nile virus NY-99	15	21	15	21	
3986	10000972	Neisseria meningitidis serogroup B Strain B16.B6	1		1		
3987	10000979	Neisseria meningitidis serogroup B Strain 8047	4		4		
3988	10000980	Arcanobacterium pyogenes Strain 42	4		4		
3989	10000986	Foot-and-mouth disease virus C1 CS30	1		1		
3990	10000987	Human rhinovirus 2 Vienna	5		5		
3991	10000989	Foot-and-mouth disease virus - type A (strain A22)	10	6	10	6	
3992	10000991	Foot-and-mouth disease virus - type A (strain A22 Iraq)	5	1	5	1	
3993	10000992	Foot-and-mouth disease virus - type SAT 1 (Strain Bot 1/68)	1		1		
3994	10000993	Foot-and-mouth disease virus - type SAT 2 (Strain Ken 3/57)	1		1		

399 5	1000994	Foot-and-mouth disease virus - type SAT 3 (Strain Bec 1/65)	1		1			
399 6	1000995	Foot-and-mouth disease virus - type Asia 1 Pakistan	1		1			
399 8	10001000	Foot-and-mouth disease virus - type SAT 2 (strain Rho 1/48)	4		4			
399 9	10001002	Swine vesicular disease virus SPA/1/93	16		16			
400 0	10001003	Neisseria meningitidis serogroup B CU385	2		2			
400 1	10001004	Human T-cell lymphotropic virus type 1 (Caribbean isolate) (Strain HS35)	1		1			
400 2	10001006	Proteus mirabilis CFT322	1		1			
400 3	10001008	Chlamydia trachomatis Serovar J	6		6			
400 4	10001019	Mus musculus B10.D2		1		1		
400 5	10001020	Peste-des-petits-ruminants virus (strain Nigeria 75/1)		1		1		
400 6	10001021	Treponema pallidum subsp. pallidum (strain Chicago)	19		19			
400 7	10001025	Classical swine fever virus Glentorf		26		26		
400 8	10001027	Vaccinia virus Connaught	1		1			
400 9	10001028	Human poliovirus 2 (strain MEF-1)	1		1			
401 0	10001032	Newcastle disease virus (strain La Sota)	17		17			
401 1	10001039	Foot-and-mouth disease virus C3 (strain Resendne-Br/55)	1		1			
401 2	10001040	Human poliovirus 2 (strain Sabin)	2	7	2	7		
401 3	10001042	Haemophilus influenzae NTHi 1479	12	4	12	4		
401 4	10001044	Neisseria meningitidis serogroup C MC51	1		1			
401 5	10001047	West Nile virus strain 2741	32		32			
401 6	10001049	Neisseria meningitidis serogroup B Strain S3446	1		1			
401 7	10001050	Neisseria meningitidis serogroup B Strain 7967	1		1			
401 8	10001051	Neisseria meningitidis serogroup A Strain 8659	1		1			
401 9	10001053	Haemophilus influenzae MinnA	9		9			
402 0	10001055	Haemophilus influenzae 6U	2		2			
402 1	10001056	Haemophilus influenzae ATCC 9795	2		2			
402 2	10001057	Pseudomonas aeruginosa CD4	1		1			
402 3	10001058	Pseudomonas aeruginosa K122-4	1		1			
402 4	10001077	Neisseria gonorrhoeae SU89	1		1			
402 5	10001078	Neisseria gonorrhoeae SU96	3		3			
402 6	10001081	Porphyromonas gingivalis HG66	2		2			
402 7	10001085	Chlamydia trachomatis Serovar D	3		3			
402 8	10001091	Borrelia burgdorferi BEP4	1		1			
402 9	10001097	Sus scrofa Yorkshire	1		1			
403 0	10001100	Plum pox virus (strain W)	6		6			

403 1		10001102	Foot-and-mouth disease virus (strain O1) O/UKG/11/2001	1	3	1	3		
403 2		10001116	Norovirus genogroup 1 isolates 96-908	4		4			
403 3		10001118	Foot-and-mouth disease virus - type O (strain HKN/14/82)	2		2			
403 4		10001120	Cryptococcus neoformans var. neoformans Serotype D	1		1			
403 5		10001121	Chlamydia trachomatis Serovar Da	1		1			
403 6		10001130	Feline leukemia virus subtype A	2		2			
403 7		10001132	Cryptococcus neoformans var. neoformans Serotype A	1		1			
403 8		10001133	Rattus norvegicus DA		3		3		
403 9		10001138	Newcastle disease virus (strain Eaves)	1		1			
404 0		10001140	Newcastle disease virus (strain WA2116)	1		1			
404 1		10001141	Chlamydia trachomatis Serovar A	22	8	22	8		
404 2		10001145	Escherichia coli 1471	2		2			
404 3		10001149	Haemophilus influenzae Subtype 1H	23		23			
404 4		10001150	Haemophilus influenzae Strain Eagan	58		58			
404 5		10001154	Transmissible gastroenteritis virus MAD88	8	1	8	1		
404 6		10001155	Haemophilus influenzae strain 1479	2		2			
404 7		10001156	Hepatitis B virus subtype AD	3	2	3	2		
404 8		10001157	H3N2 subtype Influenza A virus (A/Hong Kong(H3N2))		15		15		
404 9		10001161	Neisseria meningitidis serogroup B Strain 2996	1		1			
405 0		10001162	Neisseria meningitidis serogroup B Strain M1239	4		4			
405 1		10001207	Murine leukemia virus LP-BM5		1		1		
405 2		10001209	Tobacco mosaic virus (strain PM5)	1		1			
405 3		10001210	Tobacco mosaic virus (strain Ni568)	1		1			
405 4		10001213	Human coxsackievirus B3 (strain RK)	1		1			
405 5		10001215	Human rotavirus G9 WI61	1		1			
405 6		10001216	Rotavirus G3 strain RV-3	1		1			
405 7		10001223	Porphyromonas gingivalis OMZ 409	23		23			
405 8		10001225	H1N1 subtype Influenza A/Oklahoma/7485/01		5		5		
405 9	X	10001235	Human metapneumovirus A1 NL/1/00			3		3	
406 0		10001238	Puumala virus (strain Umea/hu)	1		1			
406 1		10001243	Theileria parva strain Marikebuni		1		1		
406 2		10001307	Small ruminant lentivirus strain It-561	1		1			
406 3		10001308	Small ruminant lentivirus strain It-P1	1		1			
406 4		10001315	H1N1 swine influenza virus (A/swine/Korea/S10/2004(H1N1))		3		3		
406 5		10001316	H1N1 swine influenza virus (A/swine/Korea/S175/2004(H1N1))		1		1		

4066	10001317	H9N2 subtype Influenza A virus (A/swine/Korea/S81/2004(H9N2))		1		1		
4067	10001318	H9N2 subtype Influenza A virus (A/swine/Korea/S83/2004(H9N2))		2		2		
4068	10001335	Japanese encephalitis virus Vellore P20778	1	1	1	1		
4069	10001392	Human adenovirus B strain Harbin04B	5		5			
4070	10001396	Murine cytomegalovirus (strain Smith) MW97.01		5		5		
4071	10001412	Rattus norvegicus Wistar-Furth		11		11		
4072	10001424	Brucella abortus W99	1		1			
4073	10001431	Infectious bronchitis virus CK/CH/LDL/971	1		1			
4074	10001439	Streptococcus sanguinis strain BD113-20		6		6		
4075	10001456	Human respiratory syncytial virus A strain RGH	1		1			
4076	10001459	Babesia bovis Argentina R1A	4		4			
4077	10001489	Proteus vulgaris Strain 5/43	1		1			
4078	10001490	Proteus mirabilis O24	1		1			
4079	10001491	Proteus mirabilis O29	1		1			
4080	10001492	Salmonella enterica subsp. enterica serovar Enteritidis SH1262	3		3			
4082	10001498	Proteus mirabilis O23	1		1			
4083	10001499	Proteus mirabilis O6	1		1			
4084	10001500	Proteus mirabilis O43	1		1			
4085	10001501	Proteus penneri ATCC 33519	1		1			
4086	10001502	Escherichia coli J-5	1		1			
4087	10001503	Acinetobacter lwoffii F78	1		1			
4088	10001504	Yokenella regensburgei PCM 2476	1		1			
4089	10001505	Yokenella regensburgei PCM 2477	1		1			
4090	10001511	Salmonella 'group A'	2		2			
4091	10001514	Providencia stuartii O33	1		1			
4092	10001519	Pseudomonas aeruginosa serotype O11	1		1			
4093	10001520	Haemophilus influenzae strain RM7004	1		1			
4094	10001523	Haemophilus influenzae strain I-69 Rd-/b+	4		4			
4095	10001524	Neisseria meningitidis strain 3006	1		1			
4096	10001526	Salmonella thompson C1 strain IS40	1		1			
4097	10001528	Escherichia coli F515	1		1			
4098	10001530	Acinetobacter haemolyticus strain 57	1		1			
4099	10001531	Acinetobacter haemolyticus strain 61	1		1			
4100	10001556	Moraxella catarrhalis 26404	1		1			
4101	10001558	Neisseria gonorrhoeae 15253	1		1			
4102	10001560	Neisseria meningitidis M982B	5		5			

410 3	10001569	Salmonella enterica subsp. enterica serovar Gallinarum/pullorum str. 11	1		1			
410 4	10001570	Salmonella enterica subsp. enterica serovar Gallinarum/pullorum str. 77	1		1			
410 5	10001573	Anatid herpesvirus 1 Clone-03	1		1			
410 6	10001578	Classical swine fever virus LPC/AHRI	5		5			
410 7	10001581	Proteus penneri 8 (O67)	1		1			
410 8	10001582	Dengue virus 1 Mochizuki	4		4			
410 9	10001583	Dengue virus 2 New Guinea C	1	38	13	38	12	
411 0	10001586	Porcine reproductive and respiratory syndrome virus 07V063	3		3			
411 1	10001587	Porcine reproductive and respiratory syndrome virus 08V204	2		2			
411 2	10001588	Providencia stuartii O4	1		1			
411 4	10001606	Norovirus genogroup 2 Hu/NoV/Farmington Hills/2002/USA	1	17	1	17		
411 5	10001614	Junin virus strain MC2		1		1		
411 6	10001615	Machupo virus strain Carvallo		1		1		
411 7	10001616	Guanarito virus strain INH-95551		1		1		
411 8	10001617	Whitewater Arroyo virus strain AV9310135		1		1		
411 9	10001618	Pichinde virus strain Munchique		1		1		
412 0	10001619	Dengue virus 3 strain 16652	16		16			
412 1	10001624	Feline infectious peritonitis virus (strain KU-2)	22	43	22	43		
412 2	10001627	Human herpesvirus 5 (strain RV798)		6		6		
412 3	10001628	Aggregatibacter actinomycetemcomitans serotype b str. Y4	1		1			
412 4	10001633	Escherichia coli 055:B5	3		3			
412 5	10001635	Dengue virus 2 PL046		7		7		
412 6	10001636	Dengue virus 2 S221		42		42		
412 7	10001638	Streptococcus pneumoniae type 27	2		2			
412 8	10001641	Streptococcus dysgalactiae subsp. equisimilis D181	1		1			
412 9	10001642	Foot-and-mouth disease virus - type O Wuppertal/FRG/82		1		1		
413 0	10001644	Streptococcus pneumoniae CCUG 1378	1		1			
413 1	10001645	Streptococcus pneumoniae CSR-SCS-2	1		1			
413 2	10001646	Streptococcus pneumoniae type 37	2		2			
413 3	10001648	Japanese encephalitis virus strain SA-14 -14-2	2		2			
413 4	10001649	West Nile virus strain 956	1		1			
413 5	10001651	Streptococcus pneumoniae type 14	26		26			
413 6	10001652	Candida albicans NIH B-792 (serotype B)	1		1			
413 7	10001659	Escherichia coli O125	1		1			
413 8	10001662	Staphylococcus aureus Cowan 1	2		2			

413 9		10001663	Streptococcus anginosus K214-2K	4		4		
414 0		10001667	Helicobacter pylori 487	1		1		
414 1		10001676	Streptococcus dysgalactiae subsp. equisimilis ATCC 12388	2		2		
414 2	X	10001680	Influenza A virus (A/X-31(H3N2)) A/X-31 X HK				176	176
414 3		10001686	Streptococcus pneumoniae type 6A	2		3		1
414 4		10001687	Streptococcus pneumoniae type 6B	5		6		1
414 5		10001688	Pseudomonas syringae pv. atrofaciens str. IMV 7836	1		1		
414 6		10001689	Pseudomonas syringae pv. atrofaciens str. IMV K-1025	1		1		
414 7		10001690	Pseudomonas syringae pv. atrofaciens str. IMV 4394	1		1		
414 8		10001693	Pseudomonas syringae pv. tagetis str. ICMP 6370	1		1		
414 9		10001697	Pseudomonas syringae pv. phaseolicola str. IMV 120a	1		1		
415 0		10001698	Pseudomonas syringae pv. morsprunorum str. GSPB 883	1		1		
415 1		10001700	Pseudomonas syringae pv. morsprunorum str. CFBP 1650	1		1		
415 2		10001705	Dengue virus 2 strain 43	2		2		
415 3		10001713	Neisseria meningitidis serogroup W-135	1		1		
415 4		10001722	Salmonella 'group D'	3		3		
415 5		10001730	Streptococcus pneumoniae type 6C	1		1		
415 6		10001731	Streptococcus pneumoniae type 2	8		8		
415 8		10001740	Candida glabrata IFO 0622	1		1		
415 9		10001747	Candida parapsilosis M1015	1		1		
416 0		10001761	BK polyomavirus strain Dunlop		1		1	
416 1		10001762	JC polyomavirus strain MAD1		1		1	
416 2		10001763	Simian virus 40 strain 776		1		1	
416 3		10001764	Classical swine fever virus Margarita (AJ704817)	4	1	4	1	
416 4		10001768	Dengue virus 2 S-16803	1		1		
416 5		10001771	Porcine reproductive and respiratory syndrome virus HuN4-F112	4	5	4	5	
416 6		10001783	Neisseria meningitidis serogroup C strain C11	1	1	1	1	
416 7		10001792	Streptococcus pneumoniae type 3	6		6		
416 8		10001794	Helicobacter pylori B128 Gerbil-adapted variant 7.13	1		1		
417 0		10001797	Neisseria meningitidis serogroup C strain MC19	1		1		
417 1		10001798	Porcine transmissible gastroenteritis coronavirus strain Pur46-MAD	4		4		
417 2		10001800	Porcine reproductive and respiratory syndrome virus HuN4	4	4	4	4	
417 3		10001801	Streptococcus pneumoniae type 8	10		10		
417 4		10001831	Moraxella catarrhalis serotype B	1		1		
417 5		10001833	Streptococcus pneumoniae type 19F	1		1		

417 6	10001834	Streptococcus pneumoniae type 19A	2		2			
417 7	10001835	Proteus penneri 7 (O61)	2		2			
417 8	10001836	Proteus penneri 14 (O59)	2		2			
417 9	10001837	Proteus penneri 15 (O52)	2		2			
418 0	10001847	Clostridium difficile BI / NAP1/ 027	1		1			
418 1	10001849	Moraxella catarrhalis serotype A	3		3			
418 2	10001851	Moraxella catarrhalis serotype C	1		1			
418 3	10001860	Neisseria meningitidis serogroup X	4		4			
418 4	10001879	West Nile virus strain 68856	2		2			
418 5	10001881	Porcine circovirus 2 -A	5		5			
418 7	10001886	Brucella ovis O20	2		2			
418 8	10001887	Brucella ovis Reo 198	1		1			
418 9	10001888	Brucella ovis 63/290	1		1			
419 1	10001895	Yersinia pseudotuberculosis str. 32777		1		1		
419 2	10001900	Streptococcus agalactiae serogroup III M781	1	1	1	1		
419 3	10001902	Human enterovirus 71 NUH0083/SIN/08	4		4			
419 4	10001913	Lactobacillus johnsonii JCM 1022	3		3			
419 5	10001918	Muscovy duck reovirus S12	4		4			
419 6	10001929	Equine rhinitis B virus 2 strain 313/75	5		5			
419 7	10001930	Equine rhinitis B virus 3 isolate 2225AS	5		5			
419 8	10001934	Chikungunya virus Singapore/11/2008	8		8			
419 9	10001943	Dengue virus 4 Burma/63632/1976	3		3			
420 0	10001944	Dengue virus 3 Thailand/PaH881/1988	2		2			
420 1	10001945	Dengue virus 1 PVP159	2		2			
420 2	10001946	Foot-and-mouth disease virus - type Asia 1 YS/CHA/05	4		4			
420 3	10001964	Porcine circovirus 1 strain 1/G	7		7			
420 4	10001965	Avian leukosis virus strain NX0101	1		1			
420 5	10001967	Avian leukosis virus isolate CAUHM01	1		1			
420 6	10001973	Vaccinia virus Acambis 2000	8		8			
420 7	10001975	Streptococcus salivarius JCM 5707	1		1			
420 8	10001997	Chikungunya virus strain LR2006_OPY1 IMT/Reunion Island/2006	1		1			
420 9	10001999	Reticuloendotheliosis virus strain HLJ071	1		1			
421 0	10002000	Porcine epidemic diarrhea virus CH/SHH/06	1		1			
421 1	10002003	Campylobacter jejuni subsp. jejuni serotype HS:15	1		1			
421 2	10002006	Bluetongue virus 16 BN96/16	2		2			
421 3	10002007	Plasmodium falciparum isolate UAS22	4		4			

421 4	10002008	Plasmodium falciparum isolate UAS31	1		1		
421 5	10002009	Plasmodium falciparum isolate UAS29	2		2		
421 6	10002010	Dengue virus 2 D2/SG/05K4155DK1/2005		49		49	
421 7	10002011	Plasmodium falciparum R29/IT4	1		1		
421 8	10002015	Human coxsackievirus A16 shzh05-1	92		92		
421 9	10002018	Bovine coronavirus Kakegawa	1		1		
422 0	10002020	Foot-and-mouth disease virus - type Asia 1 Nepal 29/97	4		4		
422 1	10002022	Escherichia coli O6 O6:K15:H31		1		1	
422 2	10002026	Murine norovirus 1 Mu/NoV/GV/MNV1/2002/USA		1		1	
422 3	10002027	Bluetongue virus 16 Kumamoto/1985	2		2		
422 4	10002028	Bluetongue virus 16 Beatrice Hill/1987	2		2		
422 5	10002034	Fish	4		4		
422 6	10002052	Toxoplasma gondii Gansu Jingtai	27		27		
422 7	10002056	Enterovirus A71 TW/2086/98	1		1		
422 8	10002069	Bordetella pertussis 509		7		7	
422 9	10002079	Clostridium difficile VPI 10463	2		2		
423 0	10002082	Rhesus rotavirus MMU 18006		3		3	
423 1	10002089	Porcine reproductive and respiratory syndrome virus SY0608	17		17		
423 2	10002090	Influenza A virus (A/swine/Denmark/101310-1/2011(H1N1))		7		7	
423 3	10002111	Dengue virus 4 TVP-376	1		1		
423 4	10002112	Dengue virus 4 1036	1		1		
423 5	10002120	Human metapneumovirus A2 TN/94-49		7		7	
423 6	10002121	Staphylococcus aureus Reynolds	2		2		
423 7	10002122	Staphylococcus aureus Becker	2		2		
423 8	10002123	Peste-des-petits-ruminants virus China/Tibet/Geg/07-30	53		53		
423 9	10002127	Escherichia coli O86:B7	1		1		
424 0	10002129	Staphylococcus aureus RN4850	1		1		
424 1	10002131	Foot-and-mouth disease virus A/HuBWH/CHA/2009	1		2		1
424 2	10002132	Duck hepatitis A virus 1 HP-1	2		2		
424 3	10002144	Chikungunya virus CHIKV/SL/2006 SL15649		26		26	
424 4	10002148	Dengue virus 4 Mexico/BC287/1997	1		1		
424 5	10002151	Duck hepatitis A virus 1 LY0801	1		1		
424 6	10002152	Duck hepatitis A virus 3 SD1201	1		1		
424 7	10002161	Streptococcus pneumoniae type 1	5		5		
424 8	10002165	Avian infectious bronchitis virus (strain CHINA/Sczy3/09)	2		2		

424 9	10002169	Avian infectious bronchitis virus (strain Holte)		2		2		
425 1	10002180	ZIKV/H. sapiens/China/SZ01/2016		40		40		
425 3	10002183	ZIKV/H. sapiens/French Polynesia/PF13/2013	4		4			
425 4	10002188	ZIKV/H. sapiens/PR1/PRVABC59/2015		12		12		
425 5	10002190	ZIKV/H. sapiens/Uganda/MR-766/1947	5	1	5	1		
425 6	10002192	Streptococcus suis 1178027	2		2			
425 8	10002196	Hepatitis E virus type 4 JAK-Sai	1		1			
425 9	10002197	Chikungunya virus MY/08/065	15		15			
426 1	10002238	Influenza A virus (A/swan/Poland/305-135V08/2006(H5N1))	1		1			
426 2	10002244	Influenza A virus (A/Taiwan/2/2013(H6N1))	1		1			
426 3	10002270	Foot-and-mouth disease virus O/Tibet/CHA/99	3		3			
426 4	10002272	Streptococcus pneumoniae 7F	5		5			
426 5	10002273	Influenza A virus (A/Hong Kong/4801/2014(H3N2))		1		1		
426 6	10002285	Yellow fever virus CNYF01/2016	1		3		2	
426 7	10002287	Salmonella enterica subsp. enterica serovar Typhi str. Quailles		1		1		
426 8	10002289	Enterobacteria phage PRD1	2		2			
426 9	10002301	ZIKV/H. sapiens/Suriname/Z1106033/2015	1		1			
427 0	10002308	West Nile virus goshawk/Austria/2008		61		61		
427 1	10002309	Andes orthohantavirus CHI-9717869	11		11			
427 2	10002310	Canine Distemper Virus strain PS	2		2			
427 3	10002313	Porcine epidemic diarrhea virus strain HeB/TS/2016/325b	1		1			
427 4	10002314	Streptococcus pneumoniae type 10A	1		1			
427 5	10002316	SARS-CoV1	1059 5	509	1060 3	930	8	421
427 6	10002317	ZIKV/H. sapiens/Brazil/BEH823339/2015	1		1			
427 7	10002321	Severe acute respiratory syndrome coronavirus 2 Wuhan/IVDC-HB-01/2019	2		2			
428 0	10002329	Severe acute respiratory syndrome coronavirus 2 USA-WA1/2020	10	27	14	27	4	
428 1	10002330	Severe acute respiratory syndrome coronavirus 2 Australia/VIC01/2020	4		9		5	
428 2	10002332	Severe acute respiratory syndrome coronavirus 2 Victoria/01/2020		22		22		
428 4	10002337	Anogeissus	1		1			
428 5	10002345	Influenza B virus (B/Phuket/3073/2013)	3		3			
428 6	10002359	Influenza A virus (A/Serbia/NS-601/2014 (H1N1))	1		1			
428 7	10002360	Influenza A virus (A/chicken/Anhui/BRI99/2016(H9N2))	1		1			
428 8	10002361	Eastern equine encephalitis virus (strain FL93-939)	10		10			
428 9	10002365	Nipah henipavirus strain NiV Malaysia	2		2			

4290		10002370	Severe acute respiratory syndrome coronavirus 2 Wuhan/Hu-1/2019	57	33	1302	53	1245	20
4291		10002371	Dengue virus 2 East Timor/ET300/2000	1		1			
4293		10002380	Severe acute respiratory syndrome coronavirus 2 ITA/INMI1/2020	4		4			
4294	X	10002382	Severe acute respiratory syndrome coronavirus 2 Germany/BavPat1/2020			3		3	
4295		10002384	Influenza A virus (A/Switzerland/9715293/2013(H3N2))	3		3			
4296		10002394	Severe acute respiratory syndrome coronavirus 2 England/MIG457/2020	3		3			
4297		10002407	Foot-and-mouth disease virus O/HN/CHA/93	9		9			
4298		10002408	Foot-and-mouth disease virus A/GDMM/CHA/2013	2		2			
4299	X	10002410	Gallid alphaherpesvirus 2 strain CVI988 (Gallid alphaherpesvirus 2 (strain CVI988))				4		4
4300		10002414	Avian leukosis virus strain HB2018003	1		1			
4301		10002422	Bordetella pertussis 186	2		2			
4302		10002442	Severe acute respiratory syndrome coronavirus 2 DEU/Vero E6 cells/2021	8		8			
4303		10002444	Porcine reproductive and respiratory syndrome virus JXA1-R	6		6			
4305		10002450	African swine fever virus pig/China/HLJ/18	5		8		3	
4307		10002484	Porcine astrovirus 1 Guangxi/Nanning/PAstV-1/2013	2		2			
4309	X	10002508	Foot and mouth disease virus A/AF72			1		1	
4310	X	10002524	SARS-CoV2 Alpha			9	150	9	150
4311	X	10002525	SARS-CoV2 Beta			55		55	
4312	X	10002526	SARS-CoV2 Epsilon			2		2	
4313	X	10002528	SARS-CoV2 Gamma			2		2	
4315	X	10002530	SARS-CoV2 Delta			11		11	
4317	X	10002532	SARS-CoV2 Mu			1		1	
4318	X	10002533	SARS-CoV2 Omicron			23	82	23	82
4319	X	10002534	SARS-CoV2 Kappa			3		3	
4320	X	10002557	Vibrio vulnificus BO62316			1		1	
4321	X	10002558	Plasmodium vivax VK210			3		3	
4322	X	10002561	Influenza A virus (A/chicken/Anhui/LH99/AH17/2017(H9N2))			2		2	
4325	X	10002570	African swine fever virus China/2018/AnhuiXCGQ			1		1	
4326	X	10002579	Senecavirus A China/SSV/LNSY01/2017			1		1	
4327	X	10002580	Influenza A virus (A/chicken/Jiangsu/JT157/2016(H7N9))			3		3	
4328	X	10002583	Severe acute respiratory syndrome coronavirus 2 South Africa/NHLS-UCT-GS-0683/2020			1		1	
4330	X	10002598	Hu/GII.4/RockvilleD1/US/2012			10		10	

433 1	X	10002600	Norovirus Hu/GII.4/Arizona/379/96019984/AZ/USA/1996			1		1	
433 2	X	10002603	Norovirus Hu/GII.4/NewOrleans/2010SP0168020210VA/Virginia/USA/2010			2		2	
433 3	X	10002611	Severe acute respiratory syndrome coronavirus 2 USA/EHC_C19_2811C/2021			1		1	
433 5	X	10002633	Human coronavirus OC43 USA/ATCC VR-759/1967			9		9	
433 6	X	10002644	Severe acute respiratory syndrome coronavirus 2 South Africa/C.1.2/2021			1		1	
433 7	X	10002652	Human metapneumovirus B2 NL/1/94			1		1	
433 8	X	10002653	Severe acute respiratory syndrome coronavirus 2 Recombinant/40589-V08H26/2022			1		1	
433 9	X	10002654	Severe acute respiratory syndrome coronavirus 2 South Africa/CERI-KRISP-K032307/2021			1		1	
434 0	X	10002655	Severe acute respiratory syndrome coronavirus 2 Botswana/R40B60_BHP_3321001247/2021			13		13	
434 1	X	10002657	Influenza A virus (A/chicken/Shandong/LY1/2017(H9N2))			1		1	
434 2	X	10002661	Severe acute respiratory syndrome coronavirus 2 Botswana/R40B59_BHP_3321001248/2021			3		3	
434 3	X	10002665	Human Coronavirus 229E USA/Seattle/SC1073/2016			1		1	
434 4	X	10002667	Human enterovirus A71 GD10-12/2010			1		1	
434 5	X	10002671	Lassa virus Pinneo (Lassa virus (strain Pinneo))			2		2	
434 6	X	10002672	Severe acute respiratory syndrome coronavirus 2 England/MILK-2D6B000/2021			1		1	
435 1	X	10002682	Porcine Deltacoronavirus ZJ17HZ0102/2017			1		1	
435 2	X	10002683	Guertu virus strain DXM			24		24	
435 3	X	10002686	Human coronavirus OC43 USA/2017			1		1	
435 4	X	10002687	Human coronavirus NL63 UF-2/2015			1		1	
435 5	X	10002691	Porcine epidemic diarrhea virus SD2014			67		67	
435 7	X	10002695	Chikungunya virus TR206/H804187			3		3	
435 9	X	10002697	Severe acute respiratory syndrome coronavirus 2 Hong Kong/VM21044713-1/2021			2		2	
436 0	X	10002707	Infectious bursal disease virus Gx			2		2	
436 1	X	10002709	Severe acute respiratory syndrome coronavirus 2 Germany/BY-MVP-0283/2020				4		4
436 8	X	10002721	Severe acute respiratory syndrome coronavirus 2 CHN/Yunnan-01/2020			1		1	
436 9	X	10002722	Severe acute respiratory syndrome coronavirus 2 IND/166/2020			1		1	

4370	X	10002723	Severe acute respiratory syndrome coronavirus 2 IND/29/2020			21		21	
4373	X	10002726	Severe acute respiratory syndrome coronavirus 2 Japan/TKYE6182/2020			1		1	
4376	X	10002729	Severe acute respiratory syndrome coronavirus 2 USA/CA-CDC-03037551-001/2020			20		20	
4384	X	10002737	Severe acute respiratory syndrome coronavirus 2 USA/Cruise-CDC-03052614-001/2020			1		1	
4386	X	10002739	Severe acute respiratory syndrome coronavirus 2 USA/Cruise-CDC-03056442-001/2020			35		35	
4388	X	10002741	Severe acute respiratory syndrome coronavirus 2 USA/Cruise-CDC-03057008-001/2020			1		1	
4391	X	10002745	Severe acute respiratory syndrome coronavirus 2 USA/Cruise-CDC-3057405-001/2020			1		1	
4396	X	10002750	Severe acute respiratory syndrome coronavirus 2 USA/MN3-MDH3/2020			2		2	
4402	X	10002756	Severe acute respiratory syndrome coronavirus 2 USA/WI-CDC-03041142-001/2020			1		1	
4408	X	10002774	Severe acute respiratory syndrome coronavirus 2 USA/NJ-CDC-LC0471426/2022				1		1
4409	X	10002775	Severe acute respiratory syndrome coronavirus 2 Canada/QC-L00478241001/2022				1		1
4410	X	10002785	Rabbit Hemorrhagic Disease Virus N11			1		1	
4411	X	10002794	Severe acute respiratory syndrome coronavirus 2 USA/AZ-CDC-LC0521423/2022			2		2	
4442	X	10002948	Bat mastadenovirus G				1		1

2 Website Features

In 2022, the IEDB continued the agile, continuous deployment methodology, whereby weekly, iterative updates were deployed to the IEDB external website (4.4). Changes of note include:

- Released new 3D viewer (iCn3D) and modified ImmunomeBrowser to use new 3D viewer on right pane where applicable. Enabled the iCn3D menu in all 3D viewers (Epitopes and Assays Results tabs, ImmunomeBrowser).
- Continued development of new Excel exports with dynamically selectable columns (pending future release).
- Integrated new NCBI SARS-CoV-2 variant python script into weekly build to display mutations on the spike glycoprotein ImmunomeBrowser.
- Added new data to API tables as requested by users.

In addition to feature updates and improvements, many outstanding bugs and defects were also addressed throughout the contract year.

Furthermore, there was one major releases (Release 2.27) of the Analysis Resource since the 2021 Annual Compendium plus numerous smaller updates made throughout the year. Changes of note include:

- New tool: Peptide eXpression annotator (pepX) helps to estimate tissue-specific peptide abundance from public RNA-Seq data.
- NetMHCIIpan 4.1 has been incorporated into the MHC II binding prediction tool and has become the 'IEDB Recommended' method.
- TCRMatch updated to version 1.1.1 to allow for TRUST4-formatted input.
- AxelF updated to allow for more flexible input and to support the output of pepX as input for the tool.
- Additional bug fixes and performance improvements

The IEDB website functionality can be divided into five categories – Home Page (Section 2.1), Specialized Searches (Section 2.2.2) however this was retired in 2021, Analysis Resource (Section 2.3), Help (Section 2.4), and More IEDB (Section 2.5), which largely correspond to the pull-down menus at the top of the home page. The subsections that follow describe the website features within these categories.

The reader will find it helpful to keep in mind how data are stored in the IEDB. Each item contained in the IEDB consists of a reference (article or submission) containing information about one or more epitopes and associated binding or response information. The same epitope can exist in multiple references.

2.1 Home Page

The IEDB Home Page (www.iedb.org) is the default screen displayed when users enter the IEDB system. Besides providing a general description of the IEDB project, the home page displays system level status and notification of scheduled updates or maintenance. As one can see in Figure 2.1, the page is divided into three columns that contain introductory information, Summary Metrics of the data, a basic data search capability, and links to the epitope prediction and analysis tools in the Analysis Resource. As users browse the IEDB system, they can return to the home page anytime by clicking *Home* on the far left of the main menu bar or by clicking on the IEDB logo in the upper left-hand corner of the page.

The top box in the left column contains a welcome statement that briefly describes the content of the database. It also has a “Learn More” link that brings the user to a web page containing more information about the support and user help features available, about the data itself, and about the IEDB program. This page is described in detail in Section 2.6. The Summary Metrics are displayed in the bottom box of this column. These numbers are intended to be a gauge of the volume of data available in the system. The Summary Metrics provide the number of epitopes (peptidic and non-peptidic), assays (T cell, B cell, and MHC ligand assays), epitope source organisms, restricting MHC alleles, and references. A middle box may appear (as shown in Figure 2.10) when there are special messages to convey to users. The text will appear in red. The middle column contains the initial search interface that is described in detail in Section 2.2.1. This section includes several finders on the search interface, thus enabling more complete queries without the need for further filtering of the results. The rightmost column has links to the most frequently used tools in the Analysis Resource.

The IEDB has just launched its updated 3D viewers! Learn more via our help article [here](#).

Welcome

The Immune Epitope Database (IEDB) is a freely available resource funded by NIAID. It catalogs experimental data on antibody and T cell epitopes studied in humans, non-human primates, and other animal species in the context of infectious disease, allergy, autoimmunity and transplantation. The IEDB also hosts tools to assist in the prediction and analysis of epitopes.

[Learn More](#)

Upcoming Events & News

[Virtual User Workshop](#) Oct 26-28
* view recordings [here](#)

Antibody Engineering & Therapeutics Exhibitor Booth Dec 4-8

[IEDB SARS-CoV-2 Epitope Analysis Videos](#)

Summary Metrics

Peptidic Epitopes	1,546,008
Non-Peptidic Epitopes	3,150
T Cell Assays	448,582
B Cell Assays	1,335,511
MHC Ligand Assays	4,691,036
Epitope Source Organisms	4,241
Restricting MHC Alleles	971
References	23,372

START YOUR SEARCH HERE

Epitope

Any
 Linear peptide
 Discontinuous
 Non-peptidic

Exact IV Ex: SIINFEKL

Assay

T Cell
 B Cell
 MHC Ligand

Ex: neutralization

Outcome: Positive Negative

Epitope Source

Organism

Antigen

MHC Restriction

Any
 Class I
 Class II
 Non-classical

Ex: HLA-A*02:01

Host

Any
 Human
 Mouse
 Non-human primate

Ex: dog, camel

Disease

Any
 Infectious
 Allergic
 Autoimmune

Ex: asthma

Epitope Analysis Resource

T Cell Epitope Prediction

Scan an antigen sequence for amino acid patterns indicative of:

- MHC I Binding
- MHC II Binding
- MHC I Processing (Proteasome, TAP)
- MHC I Immunogenicity

B Cell Epitope Prediction

Predict linear B cell epitopes using:

- Antigen Sequence Properties

Predict discontinuous B cell epitopes using antigen structure via:

- Discotope
- ElliPro

Epitope Analysis Tools

Analyze epitope sets of:

- Population Coverage
- Conservation Across Antigens
- Clusters with Similar Sequences

[Provide Feedback](#) | [Help/Request](#) | [Solutions Center](#) | [Tool Licensing Information](#)

Supported by a contract from the [National Institute of Allergy and Infectious Diseases](#), a component of the National Institutes of Health in the Department of Health and Human Services.

Last Updated: November 06, 2022

Figure 2.10 IEDB 4.4 Home page

2.2 Query


There are several ways users can find information in the IEDB. A simplified search that is especially designed for the needs of most immunologists appears on the home page. This document will refer to this particular query as the Simple Search or Home Page Search. A family of advanced query-by-example searches could previously be accessed on the Search pull-down menu and consisted of “Epitope Details”, “T Cell Assay Details”, “B Cell Assay Details”, “MHC Assay Details”, “Identifier Search”, and “Browse by 3D Structure”. This group of queries are referred to as Specialized Searches, however most of these features were retired in 2021 due to low usability. Now only “Identifier Search” and “Browse by 3D Structure” can be accessed from the Specialized Searches menu.

2.2.1 Perform a Home Page Search

The home page search feature available in the center of the home page (Figure 2.11) is designed to simplify the search process for many commonly asked queries by immunologists. It is the most frequently used query mode. The approach it takes is analogous to many travel and shopping websites. IEDB users can start their search by specifying a number of fundamental and common search parameters on the home page. On the subsequent results page, users can then apply additional and more specific filters. The epitope structure choices include linear epitopes, discontinuous epitopes, and non-peptidic epitopes. For the linear epitope search, the user can prescribe the type of search desired - an exact match to the entered sequence, a substring search (described below), or a homologous peptide sequence at identity levels of 70%, 80%, or 90% as determined by BLAST. The IEDB categorizes all assays as T cell, B cell, or MHC ligand. The MHC ligand includes binding and elution assays. Users can also select the MHC restriction for non-B cell searches. Finally, users can select the host species and the disease.


The results of the query are displayed in five tabs – Epitopes, Antigens, Assays, Receptors, and References. The Epitopes tab lists the epitope description, such as the amino acid sequence for linear epitopes, its associated antigen and source organism, the number of references in which it appears, and the number of assays in which it is used. The Antigen tab lists the antigen, its source organism, and the number of related epitopes, assays, and references. The Assays tab contains three tabs, one each for T cell, B cell, and MHC ligand assays. The Receptors tab details the receptor types alongside the sequences of each chain component, and is divided into two separate tabs for T Cell Receptors (TCRs) and B Cell Receptors (BCRs). The References tab lists the IEDB-specific reference identifier, the authors, title, abstract, and year of publication or submission. A user can cross-query specific criteria from any of these five tabs; for example, filtering on a specific assay would refresh the results to display data across all tabs which is only relevant to that assay. Additionally, a user can click on the IEDB-specific identifier link to drill down to get additional information about the epitope, assay, receptor or reference.

START YOUR SEARCH HERE

Epitope ? 

Any
 Linear peptide
 Discontinuous
 Non-peptidic


Exact Iv

Assay ? 

T Cell
 B Cell
 MHC Ligand


Ex: neutralization

Outcome: Positive Negative

Epitope Source ? 


Organism

Antigen

MHC Restriction ? 


Any
 Class I
 Class II
 Non-classical

Ex: HLA-A*02:01

Host ? 

Any
 Human
 Mouse
 Non-human primate

Ex: dog, camel

Disease ? 

Any
 Infectious
 Allergic
 Autoimmune

Ex: asthma

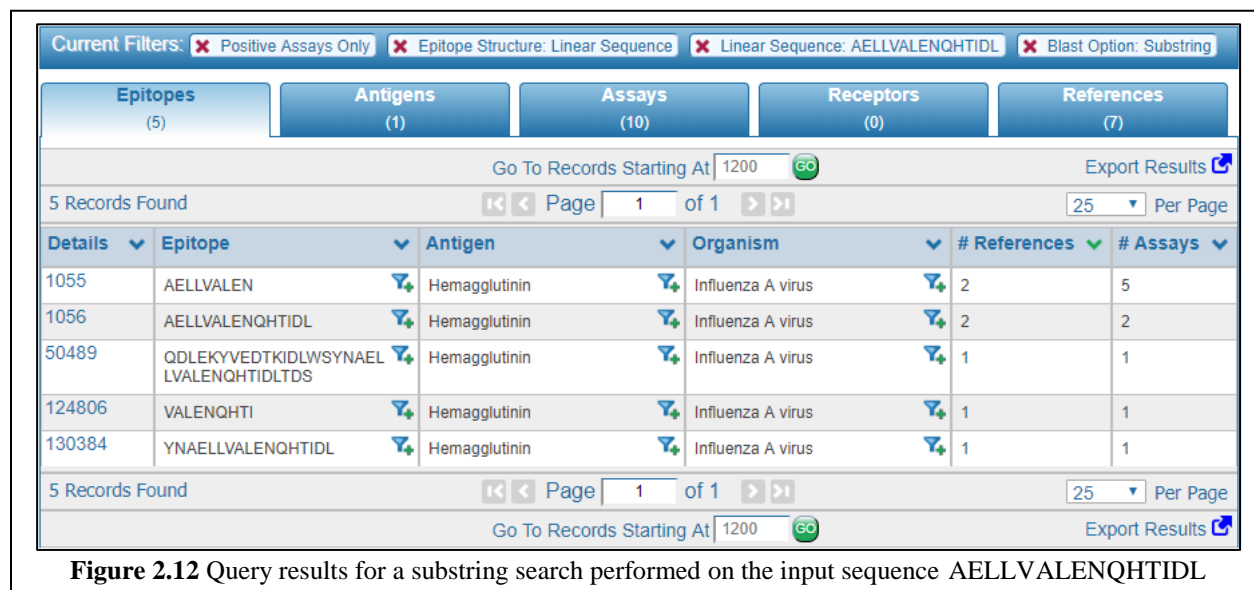
Figure 2.11 Home page simple search

Once the initial results are generated, the user can filter the results using additional filtering criteria. Auto-complete fields and associated hierarchical tree views reveal the available parameter choices. In this way, users can narrow their query to specific non-peptidic epitopes, assay types, MHC alleles, hosts, or diseases. Users can also specify article type (journal article and/or submission), the author, title, PubMed ID, and year. All filters that are applied are displayed at the top of the page. Results on most pages can be downloaded as comma separated value (CSV) files that can be read using a text editor or spreadsheet application. Users also have the ability to map positive and negative results to reference proteomes as a means of visualizing immunogenic regions using the Immunome Browser (Section 2.2.3.2).

An autocomplete feature is available for many of the fields on the left-hand side of the results page that contain additional filter criteria. This includes the non-peptidic epitope field in the Epitope box, the organism and antigen name fields in the Antigen box, the T cell, B cell, and MHC ligand assay fields in the Assay box, the specific MHC restriction field in the MHC Restriction box, the specific host field in the Host box, and the specific disease field in the Disease box. For example, as one types “hep” in the organism field in the Antigen box, several choices start to appear in a list below the text field, including “hepatitis C virus”. The user can click on a selection from the list or can continue typing to further narrow down choices. Likewise, a user may type “human” or “homo” to select “homo sapiens” in the Organism field. The list of matches includes scientific names and synonyms. These actions enable users to quickly specify their search parameters without using the finders. Use of the autocomplete circumvents the need to directly use the finders.

If the user decides not to use the autocomplete feature for a search field, they will need to use the finders. The epitope source can be prescribed for the source organism and source antigen by using the organism finder (Section 2.2.4.5) and molecule finder (Section 2.2.4.4), respectively. The user can decide whether to include B cell responses, T cell responses, and/or MHC ligand results in the search (at least one must be checked). The host organism, the MHC restriction, and the MHC class can also be specified with the help of the organism finder and the allele finder (Section 2.2.4.1). The fields using finders will allow multiple selections as search criteria. In these cases, the selections are treated as a set. Records will be considered a match if they include at least one of the selected values in the set. The search is executed by clicking the Search button and the query results can be viewed on the updated results page described in Section 2.2.3.

In addition to finding peptides in the database that contain the specified amino acid sequence, the substring search also finds epitopes within the input sequence itself. For example, when the sequence AELLVALENQHTIDL is submitted for a substring search, the query results, shown in Figure 2.12, yields five peptide sequences. Three of them (the second, third, and fifth) contain the input sequence. The two others (the first and fourth) are substrings of the input sequence that are also epitopes contained in the IEDB.



2.2.2 Specialized Searches

The Specialized Searches feature was largely retired in 2021 due to underutilization by the scientific community and high cost of maintenance. As a result, this section has been downscaled in the Annual Compendium. The two remaining features of the Specialized Searches includes the Identifier Search shown in Figure 2.13. If users know the epitope, reference, submission, or assay identifier they are seeking, they can enter that information into one of the four IEDB Identifiers fields.

IDENTIFIER SEARCH

IEDB Identifiers

Epitope ID

Reference ID

Submission ID

Assay ID

External Identifiers

PubMed ID

PDB ID

ChEBI ID

Search

Figure 2.13 Identifier Search input screen

The Browse by 3D Structure selection screen is shown in Figure 2.14. Users can select 3D structure of interest to them based on available structures within the database.

BROWSE BY STRUCTURE

- [-] B Cell Structure (3029 Assay(s)*)
- [-] T Cell Structure (321 Assay(s)*)
- [-] MHC molecule (312 Assay(s)*)
 - [+] class I (141 Assay(s)*)
 - [-] class II (84 Assay(s)*)
 - [-] human (34 Assay(s)*)
 - [+] HLA-DP (1 Assay(s)*)
 - [+] HLA-DQ (18 Assay(s)*)
 - [+] HLA-DR (15 Assay(s)*)
 - [+] mouse (30 Assay(s)*)
 - [+] non-classical (107 Assay(s)*)
- [+] mutant MHC molecule (8 Assay(s)*)
- [+] MHC Ligand Structure (1223 Assay(s)*)

* Indicates the number of distinct assays.

Figure 2.14 Browse by 3D Structure search

2.2.3 Search Results Page

The results of a search or query appear on the Results page. An example is shown in Figure 2.15. The search criteria are listed in the Current Filters section above the results table. There are five tabs for the results. Figure 2.15 displays the Epitopes tab. Results are initially sorted by the number of assays, but the user can click on the column header to change the sort order. A second click will reverse the sort order of the selected column. The table includes the epitope structure description, the corresponding antigen, source organism, the number of references in which the epitope appears, and the number of assays (positive and negative) that reside in the database. Clicking on the Epitope ID number in the far-left column will take the user to a Distinct Epitope Detail page, as seen in Figure 2.16. The bottom section of the figure contains links to

relevant tools in the Analysis Resource. The user can also refine the search results by clicking on the filter icon in the epitope, antigen, and organism fields.

The screenshot shows the IEDB search results page with the following components:

- Navigation:** Home, Specialized Searches, Analysis Resource, Help, View (1/20)
- Current Filters:** Include Positive Assays
- Filter Options (Left Sidebar):**
 - Epitope:** Any, Linear peptide, Length, Sequence, Discontinuous, Non-peptidic, 3D structure assays, Amino acid modification.
 - Epitope Source:** Organism (Ex: influenza, peanut), Antigen (Ex: core, capsid, myosin), Include related structure, Select multiple options.
 - Receptor:** Has sequence, Type (Any Type), Paired chains only.
 - Chain:** Any Type, Sequence (Exact Match).
 - Assay:** Outcome (Positive), T Cell, B Cell, MHC Ligand.
- Table Headers:** EIDB ID, Epitope, Antigen, Organism, # References, # Assays.
- Table Content (Sample Rows):**

EIDB ID	Epitope	Antigen	Organism	# References	# Assays
44920	SLPLMIAIV	65 kDa phosphoprotein	Human herpesvirus 5 (human cytomegalovirus)	148	85
123885	seaditpim			324	1048
20354	GLQPVFTL	Mario protein 1	Influenza A virus	254	655
58560	SINFEKL	Oral # 2	Salvus galvus (pisikan)	219	547
113645	MEVGWYRSPFSRVVHLYRNGK	Oligodendrocyte-myelin glycoprotein	Homo sapiens (human)	191	979
4602	ASINVEIEM	Nucleoprotein	Influenza A virus	190	448
20788	GLCTLVAML	mRNA export factor (CEP27 homolog)	Human herpesvirus 4 (Epstein Barr virus)	154	317
112741	2,4-dinitrophenyl group			140	476
130694	1-O-(alpha-D-galactosyl)-N-hexacosanoylphosphatidylserine			134	644
48237	PKYKQNTLGLAT			128	424
24766	HSLGKWLGHPOKPF	Myelin proteolipid protein	Homo sapiens (human)	118	699
53172	RAHWVIVTF	Other Human papillomavirus (Human Papilloma Virus) protein	Human papillomavirus (Human Papilloma Virus)	118	310
135815	alpha-D-Galp-(1->3)-beta-D-Galp-(1->6)-D-GlcNAcyl group			118	456
6435	GLDQVQITVY	Genome antigen	Hepatitis C virus	113	312
55748	TRHYVDDGAM	65 kDa phosphoprotein	Human herpesvirus 5 (human cytomegalovirus)	107	209
112742	2,4,6-trinitrophenyl group			107	308
32206	KVALGNIV	Genome polypeptin	Hepatitis C virus	103	289
16633	FLPDPFFRV	External core antigen	Hepatitis B virus (Human hepatitis B virus)	100	294
61086	SEEFARL	Envelope glycoprotein B	Human herpesvirus 1	99	323
6568	CLGGLLTAV	Lateral membrane protein 2	Human herpesvirus 4 (Epstein Barr virus)	98	298
30001	HAVYVFATC	Pre-glycoprotein polyprotein GP complex	Lymphocytic choriomeningitis mammarenavirus (Lymphocytic choriomeningitis virus)	94	273
61151	SLELRFRAIV	Polymerase acidic protein	Influenza A virus	94	273
16676	FLRDRIVQL	Epstein-Barr nuclear antigen 3	Human herpesvirus 4 (Epstein Barr virus)	89	280
17516	FQDPNGQFI	Nucleoprotein	Lymphocytic choriomeningitis mammarenavirus (Lymphocytic choriomeningitis virus)	85	207
37257	LLFDVPIVIV	Protein Tau-1	Primate T-lymphocyte virus 1	85	199
- Page Information:** Page 1 of 6222, Go To Records Starting At 1200, Export Results.
- Footer:** iedbAnnualCompendium2021 V8 - Word

Figure 2.15 The Epitope tab of the search results page



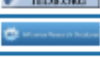


EPITOPE SUMMARY	
<p>GILGPFVTL is a linear peptidic epitope (epitope ID 20351) studied as part of Matrix protein 1 from Influenza A virus. This epitope has been studied for immune reactivity in 236 publication(s), tested in 524 T cell assays, 5 B cell assays, 115 MHC ligand assays and has 3D structure(s) 10GA, 5JHO, STEZ, ZVLJ, SISZ, SEGI, SEUD, ZVLK, ZVLL, ZVLR, 1HHI and 4NT6.</p>	
COMPILED DATA	
MHC Ligand Assay(s) 115	
MHC molecule	Positive / All
HLA-A*02:01	75 / 75
HLA-A2	10 / 10
HLA-A*02:06	4 / 5
HLA-A*02:02	4 / 4
HLA-A*02:03	4 / 4
HLA-C*08:01	4 / 4
HLA-A*68:02	1 / 3
HLA-E	0 / 2
HLA-E*01:03	1 / 1
SLA-3*02:02	1 / 1
HLA-A*01:01	0 / 1
HLA-A*03:01	0 / 1
HLA-A*11:01	0 / 1
HLA-A*24:02	0 / 1
HLA-A3	0 / 1
HLA-DRA*01:01/DRB1*01:01	0 / 1
B Cell Assay(s) 5	
Assay Type	Positive / All
qualitative binding	4 / 4
complement-dependent cytotoxicity	1 / 1
T Cell Assay(s) 524	
Assay Type	Positive / All
IFN γ release	155 / 160
qualitative binding	124 / 126
cytotoxicity	79 / 83
dissociation constant KD	28 / 36
off rate	14 / 14
on rate	14 / 14
proliferation	13 / 14
TNF α release	12 / 13
activation	10 / 10
3D structure	9 / 9
CCL4/MIP-1b release	6 / 6
pathogen burden after challenge	6 / 6
survival from challenge	5 / 6
degranulation	4 / 5
IL-2 release	4 / 5
perforin release	4 / 4
TNF release	3 / 4
granzyme B release	3 / 3
CXCL9/MIG release	1 / 1
decreased disease	1 / 1
IL-12 release	1 / 1
T cell- APC binding	1 / 1
IL-10 release	0 / 1
IL-4 release	0 / 1
EXTERNAL RESOURCES	
Resource	Link
 IEDB-AR: MHC-I Processing ↗	Predict MHC class I processing ↗
 IEDB-AR: MHC-I ↗	Predict MHC class I binding affinity ↗
 IEDB-AR: B cell scales ↗	Predict B cell epitopes ↗
 IRF ↗	View sequence feature: Influenza A_M1_experimentally-determined-epitope_58(9) details at IRF ↗
 IRF ↗	View sequence feature: Influenza A_M1_monoclonal-antibody-binding-site_58(9) details at IRF ↗

Figure 2.16 An example of the epitope detail page

The second tab on the results page is the Antigens tab (Figure 2.17). Results are initially sorted by the number of assays, but the user can click on the column header to change the sort order. The table includes the antigen name, the corresponding source organism, the number of associated antigen epitopes in the IEDB, and the number of assays (positive and negative) that reside in the database. The user can explore the data related to a particular antigen by clicking on the funnel icon in the righthand side of the Antigen column. The data in the other three tabs will change accordingly as seen in Figure 2.18.

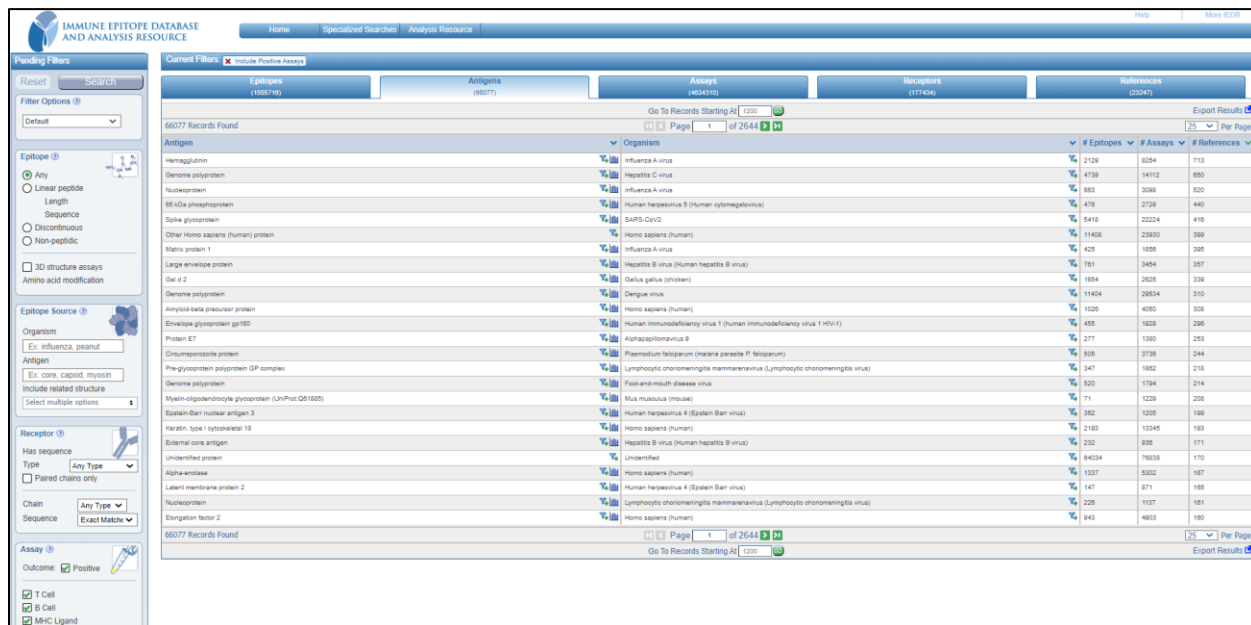


Figure 2.17 The Antigens tab of the search results page

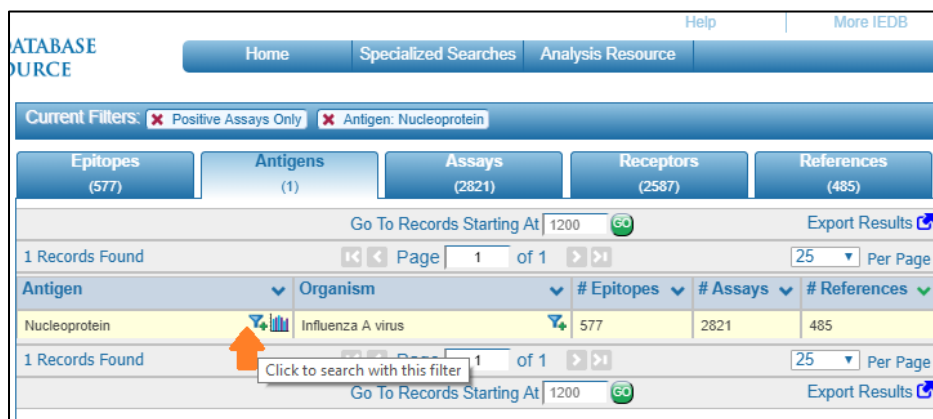


Figure 2.18 The Results filtered by the Nucleoprotein of the Influenza A virus. This was achieved by clicking on the filter icon in the antigen column. The content of the other tabs contain data specific to this antigen.

The third tab on the results page is the Assays tab (Figure 2.19). Results are initially sorted by the Assay Description, but the user can click on the column header to change the sort order. The table contains three additional tabs, one each for the three assay categories – T cell, B cell, and MHC Ligand. The table for the T cell assays includes the assay ID, the reference, epitope, host organism, immunization, assay antigen, antigen epitope relation, MHC restriction, and assay description. The content of the B cell assay tab is the same except it omits the MHC restriction column. The MHC ligand assay tab contains fields for assay ID, reference, epitope, antigen processing, MHC restriction, assay description, and quantitative measure. Clicking on the Assay ID in the far-left lane will take the user to an Assay Detail page. An example is given

in Figure 2.20. The actual page contains additional fields below what is shown in the figure, including sections for Epitope Reference Details, Immunization, Host Details, 1st In Vivo Process, Administration Details, 1st Immunogen, Immunogen Details, Immunization Comments, T Cell Assay, Effector Cells, Antigen Presenting Cells, MHC Allele, Antigen, Antigen Details, and Assay Reference Details. The user can also refine the results further by clicking on the filter icon that is available in several of the fields, such as reference, epitope, and host.

EIDB ID	Reference	Epitope	Host	Immunization	Assay Antigen	Antigen Epitope Relation	MHC Restriction	Assay Description
1505273	P. Chong, Infect Immun 1992	ADIVSEIFGCTTIV KTGGPADHYYSKVT A Pertussis toxin subunit 3 precursor (106-136) Borrelia pertussis	Mus musculus BALB/c	Administration in vivo with Pertussis toxin subunit 3 precursor (Source: Antigen)	ADIVSEIFGCTTIV KTGGPADHYYSKVT A Pertussis toxin subunit 3 precursor (106-136) Borrelia pertussis	Epitope	H2-d class II	3d-thymidine proliferation Positive-High
1810409	Xuefeng Wang, Parasit Vectors 2010	AKQVNICQPKKELD 22.6 kDa tegumental antigen (111-125) Sclerosoma japonicum	Mus musculus C57BL/6J	Administration in vivo with AKQVNICQPKKELD (Epitope)	AKQVNICQPKKELD 22.6 kDa tegumental antigen (111-125) Sclerosoma japonicum	Epitope	H2-b class II	3d-thymidine proliferation Positive-High
1737052	S. Chosh, Infect Immun 1999	ALNRFQIQVLEIKS hemagglutinin (511-525) influenza A virus	Mus musculus BALB/c	Administration in vivo with ALNRFQIQVLEIKS (Epitope)	ALNRFQIQVLEIKS hemagglutinin (511-525) influenza A virus	Epitope	H2-d class II	3d-thymidine proliferation Positive-High
1648562	A.G. Pisci, J Biol Chem 2009	ALSTLVNKKI 90 kDa chaperonin 2 (256-266) Mycobacterium tuberculosis	Rattus norvegicus (Norway rat)	Administration in vivo with ALSTLVNKKI (Fragment of Source: Antigen) Released by administration in vivo	ALSTLVNKKI 90 kDa chaperonin 2 (256-266) Mycobacterium tuberculosis	Epitope	RT1-B1	3d-thymidine proliferation Positive-High
1500620	Maria M Garcia-Brones, Virology 2004	ANHCSDAMNIFEEV polyprotein (2103-2117) Foot-and-mouth disease virus C-38c1	Bos taurus Landrace X Large White	Administration in vivo with Foot-and-mouth disease virus C-38c1 (Source: Organism)	ANHCSDAMNIFEEV polyprotein (2103-2117) Foot-and-mouth disease virus C-38c1	Epitope	SLA class II	3d-thymidine proliferation Positive-High
1853781	J. Erume, J Biol Health Sci 2011	ARLVSEAMHTEDK Nucleoprotein (391-405) Measles virus strain Edmonston (Measles morbillivirus strain Edmonston)	Mus musculus CBA	Administration in vivo with Nucleoprotein (Source: Antigen)	ARLVSEAMHTEDK Nucleoprotein (391-405) Measles virus strain Edmonston (Measles morbillivirus strain Edmonston)	Epitope	H2-K class II	3d-thymidine proliferation Positive-High
1481611	Gerard E. Hancock, J Biol Virol 2003	CSNPTCWAICKRIP Major surface glycoprotein G (178-190) Human respiratory syncytial virus A2 (human respiratory syncytial virus strain A2)	Mus musculus S.A.	Administration in vivo with Major surface glycoprotein G (Source: Antigen)	CSNPTCWAICKRIP Major surface glycoprotein G (178-190) Human respiratory syncytial virus A2 (human respiratory syncytial virus strain A2)	Epitope	H2-5 class II	3d-thymidine proliferation Positive-High
1685144	Piriam Das, Neurosci Aging 2003	DAEFRHDSQVEVHHQK Amyloid-beta precursor protein (672-687) Homo sapiens (human)	Mus musculus HLA-DQ8.02 Ig	Administration in vivo with DAEFRHDSQVEVHHQK (Fragment of Source: Antigen)	DAEFRHDSQVEVHHQK Amyloid-beta precursor protein (672-687) Homo sapiens (human)	Epitope	H2 class II	3d-thymidine proliferation Positive-High
1685131	Piriam Das, Neurosci Aging 2003	DAEFRHDSQVEVHHQ KLVFFAEDEVKNGA KSLRIVGQVIVA Amyloid-beta precursor protein (672-713) Homo sapiens (human)	Mus musculus HLA-DQ8.02 Ig	Administration in vivo with DAEFRHDSQVEVHHQ (Epitope)	DAEFRHDSQVEVHHQ KLVFFAEDEVKNGA KSLRIVGQVIVA Amyloid-beta precursor protein (672-713) Homo sapiens (human)	Epitope	H2 class II	3d-thymidine proliferation Positive-High
1643838	Michelle A Kutzler, Virology 2008	ANKEEELSSQVSTVLSKLS Hemagglutinin (1-120) Influenza A virus	Mus musculus BALB/c	Admin...	ANKEEELSSQVSTVLSKLS Hemagglutinin (1-120) Influenza A virus	Epitope	H2-Dd	3d-thymidine proliferation

Figure 2.19 The Assays tab of the search results page. The page contains three additional tabs for T Cell, B Cell, and MHC Ligand Assays

T Cell Assay	
Qualitative Measurement	Positive-High
Method/Technique	³ H-thymidine
Measurement of	proliferation
Effector Cells	
Effector Cell Tissue Type	Spleen
Effector Cell Type	T cell
Effector Cell Culture Conditions	Direct Ex Vivo
Antigen Presenting Cells	
Cell Tissue Type	Spleen
Cell Type	Splenocyte
Cell Culture Conditions	Direct Ex Vivo
Autologous or Syngeneic	Y
MHC Allele	
MHC Allele Name	H2-b class II
MHC Evidence Code	T cell assay -Biological process measured
Antigen	
Epitope Relation	Epitope
Chemical Type	Linear peptide
Linear Sequence	LFAAFPSFAGLRPTFDTRLM
Starting Position	21
Ending Position	40
Source Molecule Name	14 kDa antigen
Source Accession	P0A5B7.2 🔗
Source Organism ID	1773
Source Organism	Mycobacterium tuberculosis
Antigen Details	
Antigen Evidence Code	Exact match to reference information
Assay Reference Details	
Assay Comments by IEDB Curator	Spleen T cells from heat-killed H37Ra immunized mice or LNC cells from soluble H37Ra extracts immunized mice proliferated in response to the epitope.
Location of Assay Data in Reference	Figures 3 and 5

Figure 2.20 The top three sections of a sample T cell assay detail page.

The fourth tab on the results page is the Receptors tab (Figure 2.21). The Receptors tab indicates the distinct receptor group, the species, and the receptor types alongside the sequences of each chain component. The display is subdivided into two separate tabs for TCRs and BCRs. Clicking on the Group ID links brings the user to a receptor specific details page.

The screenshot shows the IEDB website interface with the 'Receptors' tab selected. The left sidebar contains various filters such as 'Epitope', 'Antigen', 'Assay', and 'Receptor'. The main content area displays a table of receptor results. The table has columns for 'Group ID', 'Type', 'Chain 1 CDR3', and 'Chain 2 CDR3'. The first few rows show entries for Homo sapiens (human) and Mus musculus (mouse) with various CDR3 sequences.

Figure 2.21 The Receptors tab of the search results page

The fifth tab on the results page is the References tab (Figure 2.22). Results are initially sorted by the year of publication or submission, the most recent references first. Again, the user can click on the column header to change the sort order. The table includes the IEDB-assigned reference ID, the PubMed ID, authors, article title, abstract, and the date. For journal articles, the information is downloaded directly from PubMed. Clicking on the Ref ID in the far-left lane will take the user to a Reference Detail page (Figure 2.23).

The screenshot shows the IEDB website interface with the 'References' tab selected. The left sidebar contains the same filters as in Figure 2.21. The main content area displays a table of reference results. The table has columns for 'IEDB ID', 'PMID', 'Author', 'Title', 'Journal', and 'Date'. The first few rows show recent publications, such as 'Insight into the molecular-level details of cost cell interactions with IgG' and 'Tumor-associated antigenic peptides are present in the HLA class I ligandome of cancer cell line derived extracellular vesicles'.

Figure 2.22 The References tab of the search results page

Reference	
Article Authors	Shira Yair-Sabag; Valentina Tedeschi; Carolina Vitulano; Elon Barnea; Fabian Glaser; Dgani t Melamed Kadosh; Joel D Taurog; Maria Teresa Fiorillo; Rosa Sorrentino; Arie Admon
Article Title	The Peptide Repertoire of HLA-B27 may include Ligands with Lysine at P2 Anchor Position.
Reference Detail	
Reference ID	1033511
Abstract	The HLA-B*27 peptidome has drawn significant attention due to the genetic association between some of the HLA-B*27 alleles and the inflammatory rheumatic disease ankylosing spondylitis (AS), for which a comprehensive biological explanation is still lacking. This study aims to expand the known limits of the HLA-B*27 peptidome to facilitate selection and testing of new peptides, possibly involved in the disease. The HLA peptidomes of HeLa and C1R cell lines stably transfected with the AS-associated HLA-B*27:05 allele, the nonassociated HLA-B*27:09 allele, or their cysteine 67 to serine mutants (C67S), are analyzed on a very large scale. In addition, the peptidomes of HLA-B*27:05 and HLA-B*27:05-C67S are analyzed from the spleens of rats transgenic for these alleles. The results indicate that C67S mutation increases the percentage of peptides with glutamine or lysine at their P2 position (P2-Lys), in both HLA-B*27:05 and HLA-B*27:09. Furthermore, a small fraction of HLA-B*27 peptides contains lysine at their second position (P2), in addition to the more commonly found peptides with arginine (P2-Arg) or the less common glutamine (P2-Gln) located at this anchor position. Overall these data indicate that peptides with P2-Lys should be considered as real ligands of HLA-B*27 molecules and taken into account while looking for putative peptides implicated in the AS.
Affiliations	Department of Biology, Technion-Israel Institute of Technology, Haifa, Israel; Department of Biology and Biotechnology "C. Darwin", Sapienza University of Rome, Rome, Italy; Bioinformatics Knowledge Unit, The Lorry I. Lokey Interdisciplinary Center for Life Sciences and Engineering, Technion-Israel Institute of Technology, Haifa, Israel; Department of Internal Medicine, University of Texas Southwestern Medical Center, Dallas, USA.
Date	2018
Reference Type	Literature
PubMed ID	29393594
Journal	Proteomics
Journal Volume	18
Article Pages	e1700249
Journal ISSN	1615-9861
Curation Last Updated	2019-01-03 23:39:42
Related Information	
Epitopes	34798 (Click for related results)
Bcell Assays	0
Tcell Assays	0
MHC Ligand Assays	42431 (Click for related results)

Figure 2.23 An example of the Reference detail page

2.2.3.1 Immunome Browser

The Immunome Browser is a feature integrated into the IEDB to more efficiently display large sets of immune epitope data. The purpose of the tool is to allow users to explore how often each protein region has been studied in immune assays and in how many assays the immune response was positive or negative. The Immunome Browser “maps” query results from the IEDB onto a reference proteome. A reference proteome is used because (1) epitopes reported in IEDB were identified for different strains and protein isoforms – mapping to the reference protein allows to visualize and study such epitopes as they would have the same antigen; (2) different mutant variants of the same epitope were tested and reported; and (3) immune response varies among studies and assays due to heterogeneity of samples and complexity of immune response. The Immunome Browser presents the mapped data along with calculated ‘epitope prominence scores’ that rank known epitopes based on how frequently subjects responded to each epitope, allowing identification of immunodominance. Both linear and discontinuous linear peptide epitopes with T cell or B cell responses

are supported for viewing using the Immunome Browser. It is accessible via the Immunome Browser icon on the Antigen tab on the results page, as seen in Figure 2.24.

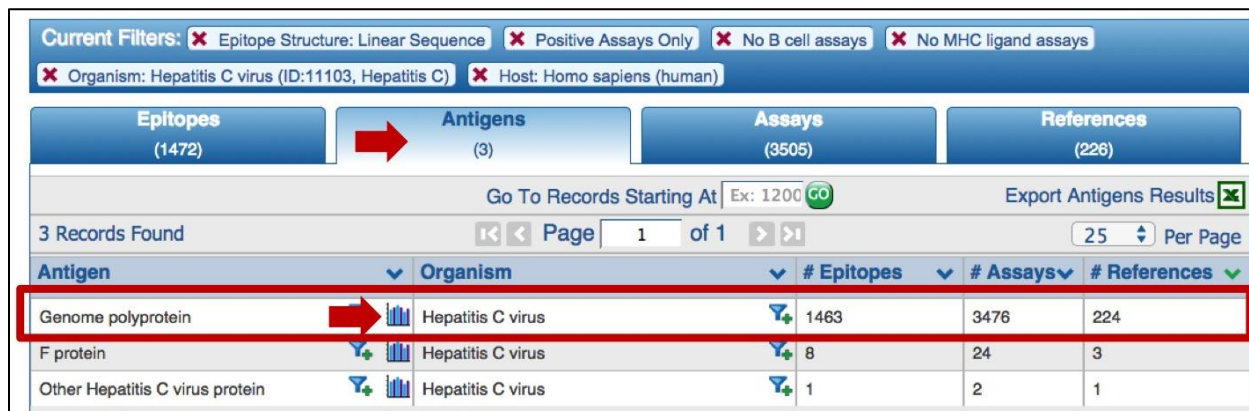


Figure 2.2 The Immunome Browser is accessed by clicking on the icon shown on the Antigen tab on the results page of a query

In order to make the Immunome Browser capability more prominent, a pop-up window was added that appears the first time a user goes to the Antigen tab after making a search, as seen in Figure 2.3. It can be easily dismissed by clicking on the large X in the upper right-hand corner or clicking anywhere else on the window.

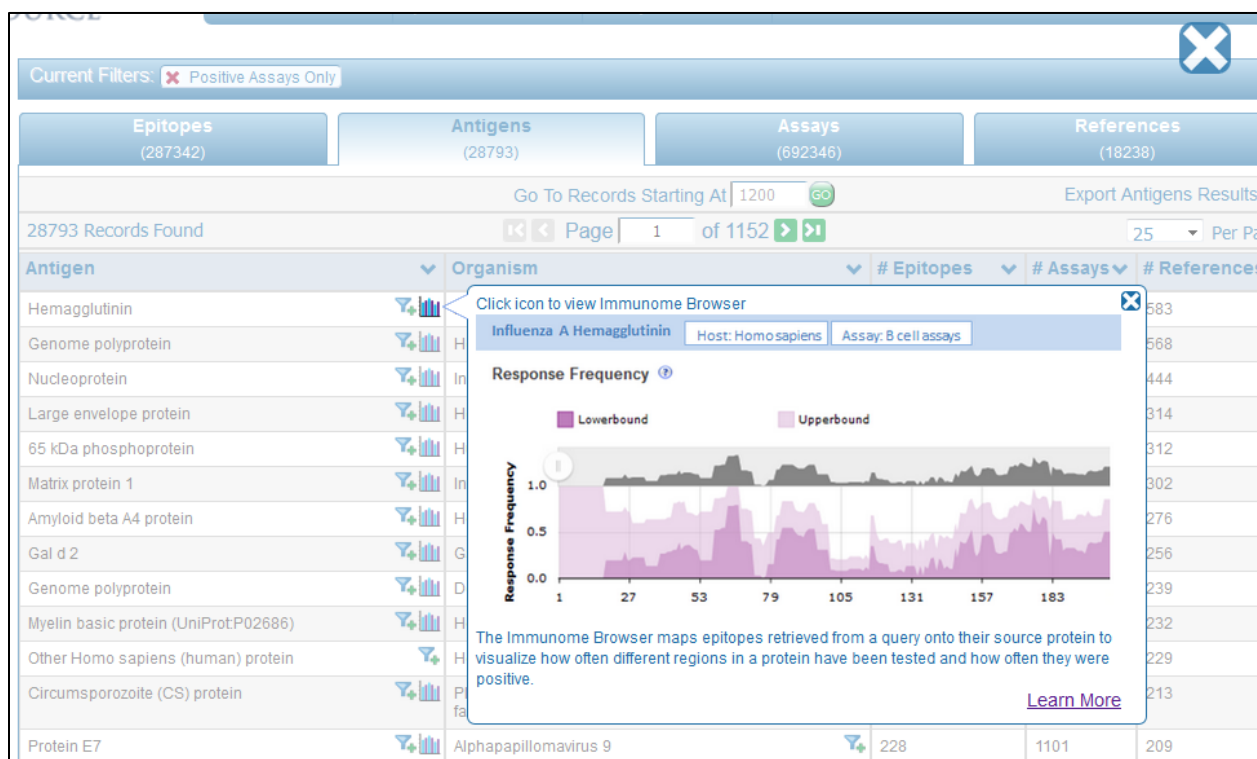


Figure 2.3 A pop-up window appears each time the user clicks on the Antigen tab on the Results page. It highlights the Immunome Browser icon and briefly explains the Immunome Browser's functionality.

In the Immunome Browser, each selected antigen is considered to be a target protein, and the tool by default maps and visualizes all epitopes in the query result that are from the source antigen, which is the target

protein or any of its taxonomic child. The mapping is done using the alignment between the source antigen and the target protein, and matching the epitope positions in the source antigen with the residue positions in the target protein. By default, epitopes that were tested as negative for the correspondent query will be mapped as well, even if the user queried only for positive assays. Also, the epitopes that failed to satisfy the default mapping criteria won't be mapped and visualized. The Immunome Browser output for query criteria of T cell assays, linear epitopes, Hepatitis C virus is shown in Figure 2.4.



Figure 2.4 Immunome Browser graphical output displaying assay results mapped onto a reference proteome by residue position

Two graphs are generated in the Immunome Browser. The upper plot titled “Response Frequency” renders the lower (dark color) and upper (light color) bounds of the 95% confidence interval (CI) of the response frequency (RF) for each position target protein position, averaged over all epitopes mapped to that position. RF is calculated as the number of positively responded subjects (or individuals in this case) relative to the total number of those tested summed up by mapped epitopes. The graph, however, displays only the upper and lower bounds of the 95% confidence interval (CI) for the RF score and not the RF score itself. The lower plot, titled “Epitope Assay Counts,” indicates the number of positive and negative assays for each residue position.

Epitope ID	Epitope Sequence	Mapped Position	Identity	Subjects Tested	Subjects Responded	Assays Positive	Assays Negative	Response Freq (95% CI)
42601	MSYVYDQCTKQYDQYDQ*	1-18	94%	1	0	0	1	0.00 (0.00-0.94)
42601	MSYVYDQCTKQYDQYDQ*	1-18	94%	1	0	0	1	0.00 (0.00-0.94)
42603	MSYVYDQCTKQYDQYDQ*	1-18	88%	2	0	0	2	0.00 (0.00-0.78)
42602	MSYVYDQCTKQYDQYDQ*	1-8	88%	4	0	0	2	0.00 (0.00-0.81)
1320773	MSYVYDQCTKQYDQYDQ*	1-18	88%	1	0	0	1	0.00 (0.00-0.94)
42606	MSYVYDQCTKQYDQYDQ*	1-18	100%	1	0	0	1	0.00 (0.00-0.94)
42673	MSYVYDQCTKQYDQYDQ*	1-14	92%	40	3	1	0	0.08 (0.02-0.14)
42605	MSYVYDQCTKQYDQYDQ*	1-20	90%	23	1	1	1	0.04 (0.00-0.18)
42609	MSYVYDQCTKQYDQYDQ*	1-20	100%	7	2	2	2	0.29 (0.05-0.84)
42607	MSYVYDQCTKQYDQYDQ*	1-18	100%	1	0	0	1	0.00 (0.00-0.94)
42605	MSYVYDQCTKQYDQYDQ*	1-14	100%	1	0	0	1	0.00 (0.00-0.94)
42691	MSYVYDQCTKQYDQYDQ*	1-8	100%	3	0	0	1	0.00 (0.00-0.81)
42679	MSYVYDQCTKQYDQYDQ*	1-14	85%	00	5	1	0	0.20 (0.04-0.18)
42679	MSYVYDQCTKQYDQYDQ*	1-18	85%	00	5	1	0	0.20 (0.04-0.18)
42698	MSYVYDQCTKQYDQYDQ*	1-18	100%	9	1	1	2	0.11 (0.00-0.41)
462390	MSYVYDQCTKQYDQYDQ*	1-18	94%	4	2	1	0	0.50 (0.10-0.88)
61789	STVYVYDQCTKQYDQYDQ*	2-26	91%	1	0	0	1	0.00 (0.00-0.94)
61790	STVYVYDQCTKQYDQYDQ*	2-10	100%	24	2	2	1	0.08 (0.01-0.23)
61787	STVYVYDQCTKQYDQYDQ*	2-8	87%	70	2	2	2	0.03 (0.01-0.10)
61788	STVYVYDQCTKQYDQYDQ*	2-10	88%	17	3	1	1	0.18 (0.05-0.38)
65387	TVYVYDQCTKQYDQYDQ*	3-11	77%	2	0	0	1	0.00 (0.00-0.78)

Figure 2.5 Immunome Browser tabular output.

In addition to the plots, a table is generated that displays the epitope ID, sequence, position, and identity, as well as the number of subjects tested, the number who responded, and number of positive and negative assays, and the response frequency (Figure 2.5). The user can interactively zoom in and out on the plots to a specific protein region and, if the check box for the table to correspond the plots is selected, the table is automatically updated to show only epitope from the selected region. Alternatively, the user can specify the residue start and end positions at the bottom of plots and click Update Display.

2.2.4 Finders Overview

Several finders (Allele, Assay, Disease, Molecule, Geolocation, and Organism) are available to help facilitate selections and control vocabulary usage, thus improving query results. At times the potential list of selections can be quite extensive, and the finders help users make selections from large lists. The finders can be utilized when performing all queries. Multiple selections can be made when utilizing finders during a query.

2.2.4.1 Allele Finder

The MHC Allele Finder facilitates the selection of one or more MHC alleles. The finder consists of four sections as shown in Figure 2.6. The top section lists the alleles that the user selects. The selections can be removed by clicking on the red X of an allele name or by clicking on the Reset button to delete all selected alleles. When the user has completed the selection process, the user needs to click the green Apply button. The middle-left section of the finder contains the “Search By” panel. Users can search by allele name, source organism, and/or class (I, II, or non-classical). After the user supplies their search criteria and clicks the Search button, the system will filter the list of MHC alleles using the organism, class, and allele provided. The allele finder uses wild card characters by default on both ends of criteria entered into the allele field. The system then returns in the bottom “Search Results” panel a list of any alleles that contain the value in the name field and match the class selected. Users can click on the green “+” icon to select an allele. Users can also click on the yellow highlighter icon to highlight the allele in the allele tree in the middle right section of the finder. Alternatively, a user can browse the hierarchical allele tree. Clicking on a node in this tree will add the allele to the Current Selections section of the finder.

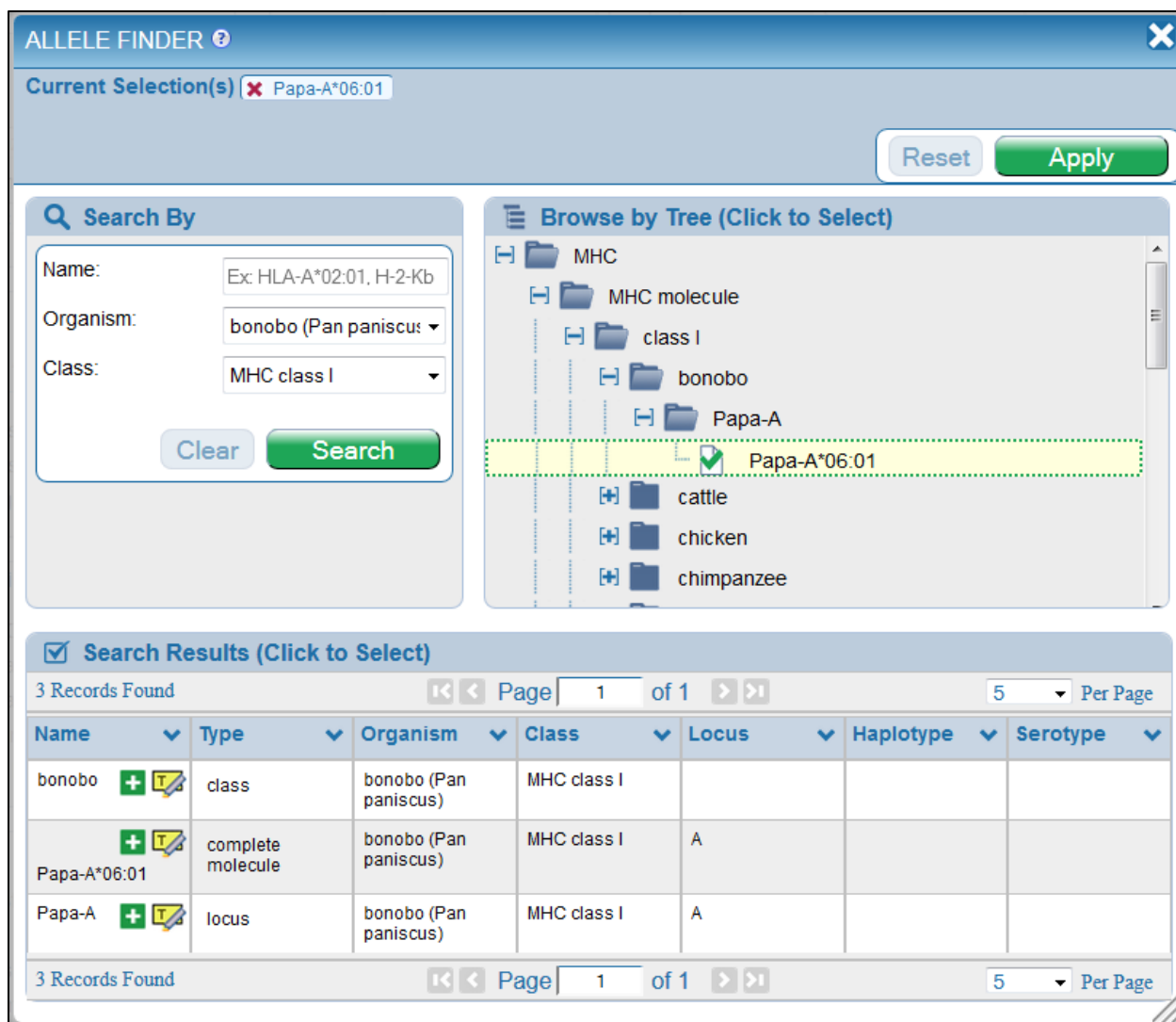


Figure 2.6 Allele Finder. Alleles are displayed in a tree structure and users can search by allele name, source organism, and MHC class (I, II, or non-classical)

2.2.4.2 Assay Finder

The IEDB contains experimental data characterizing antibody and T cell epitopes studied in humans, non-human primates, and other animal species. Search results may easily be narrowed to include only the specific assay categories of T Cell Assays, B Cell Assays, and MHC Ligand Assays. Additionally, one may further narrow search results to very specific experimental methods using the Assay Finder.

The Assay Finder is used to facilitate the selection of one or more assay types and lists all assay types in the selected assay category. The Assay Finder for T Cell, B cell, and MHC Ligand assays can be found in the Assay section on the home page search interface, the left-hand side of the Results page, and on their respective Specialized Search pages. These versions of the Assay Finder organize and display assays in a tree structure, as seen in Figure 2.7 for the T Cell assays. Users can also search for assays by their method/technique and by the characters within the assay's name. The figure shows the results for ELISA type assays for "CCL1/TCA-3 release". Users can click on the green "+" icon to select an assay. Users can also click on the yellow highlighter icon to highlight the assay in the assay tree in the middle right section

of the finder, as is shown in the figure. Alternatively, a user can browse the hierarchical assay tree. Clicking on a node in this tree will add the assay to the Current Selections section of the finder.

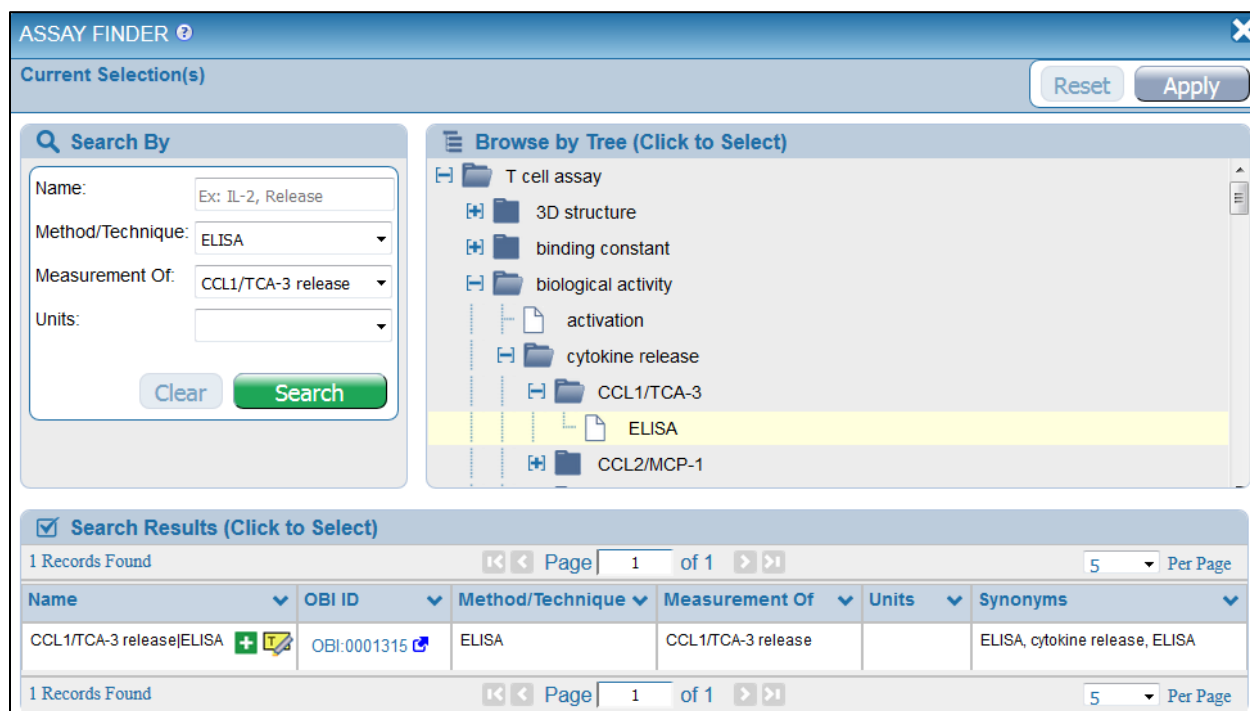


Figure 2.7 Assay Finder on the T Cell Advanced Search page. Assays are displayed in a tree structure and users can find displays by their method/technique and by the characters with which the assay name starts

2.2.4.3 Disease Finder

This query feature allows users to select a disease association based on the host disease state or status as captured in curation. Diseases are organized into a browsable tree structure. Upon the selection of host disease state/status, the system provides the user with a hierarchical *Disease Tree* that organizes the data at the highest level according to the five disease categories (Allergy, Autoimmunity, Infectious Disease, Additional Diseases by category, and Transplant-related Disease and Allo-reactivity) and then by the anatomical location of the disease (e.g. respiratory tract, gastrointestinal tract). In this way the user can either type in the disease name or synonym of interest, or navigate the Disease Tree by disease manifestation and physical location to see what is available. A node for healthy subjects is also included. The Disease Finder is fully functional for only Autoimmunity. The finder will be revised with an updated disease ontology and hierarchical tree during 2017 based on the Disease Ontology that will make it functional for the other disease categories. Infectious disease searches are best accommodated using the Organism Finder to specify the antigen.

Figure 2.8 shows the Disease Tree with diabetes highlighted. DOID refers to the Disease Ontology identifier. The DTREE designates an IEDB-specific identifier that will be updated later with a DOID number when the Disease Ontology incorporates the terms.



2.2.4.4 Molecule Finder

The Molecule Finder is used to facilitate the selection of source antigens, immunogens, and epitopes. There are two versions of the Molecule Finder – one specifically for non-peptidic molecules found in the Epitope box in the results page, and another that includes peptidic and non-peptidic branches, found in the Antigen box in the results page. The former contains the structures curated by the Chemical Entities of Biological Interest (ChEBI) database. An example of the non-peptidic finder is shown in Figure 2.9.

NON-PEPTIDIC MOLECULE FINDER

Current Selection(s) ✖ penicillin Reset Apply

Search By

Name:

Molecule ID:

Search

Browse by Tree (Click to Select)

- gatifloxacin
- indometacin
- lomefloxacin
- methotrexate
- monocarboxylic acid
- norfloxacin
- ochratoxin A
- oxolinic acid
- penicillin

Search Results (Click to Select)

18 Records Found Page 1 of 4 5 Per Page

Molecule Name	Synonyms	Database ID	Organism Name
penicillin	Penicillin, penicillins, penicillins	ChEBI:17334	
benzylpenicillin	2,2-dimethyl-6beta-(phenylacetamido)penam-3alpha-carboxylic acid, Benzylpenicillin, PENICILLIN G, (2S,5R,6R)-3,3-dimethyl-7-oxo-6-(phenylacetamido)-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid, ...more...	ChEBI:18208	
penicillin O	6beta-[(prop-2-en-1-ylsulfanyl)acetamido]-2,2-dimethylpenam-3alpha-carboxylic acid, almeceiline, (2S,5R,6R)-6-[[[(allylsulfanyl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carb...more...	ChEBI:51207	

Figure 2.9 The non-peptidic version of the Molecule Finder. This finder is accessed in the Epitope filter on the search results page.

With regard to the protein or peptidic branch of the Molecule Finder, individual GenPept proteins utilized by IEDB data are assigned to parent proteins from reference proteomes by sequence homology.

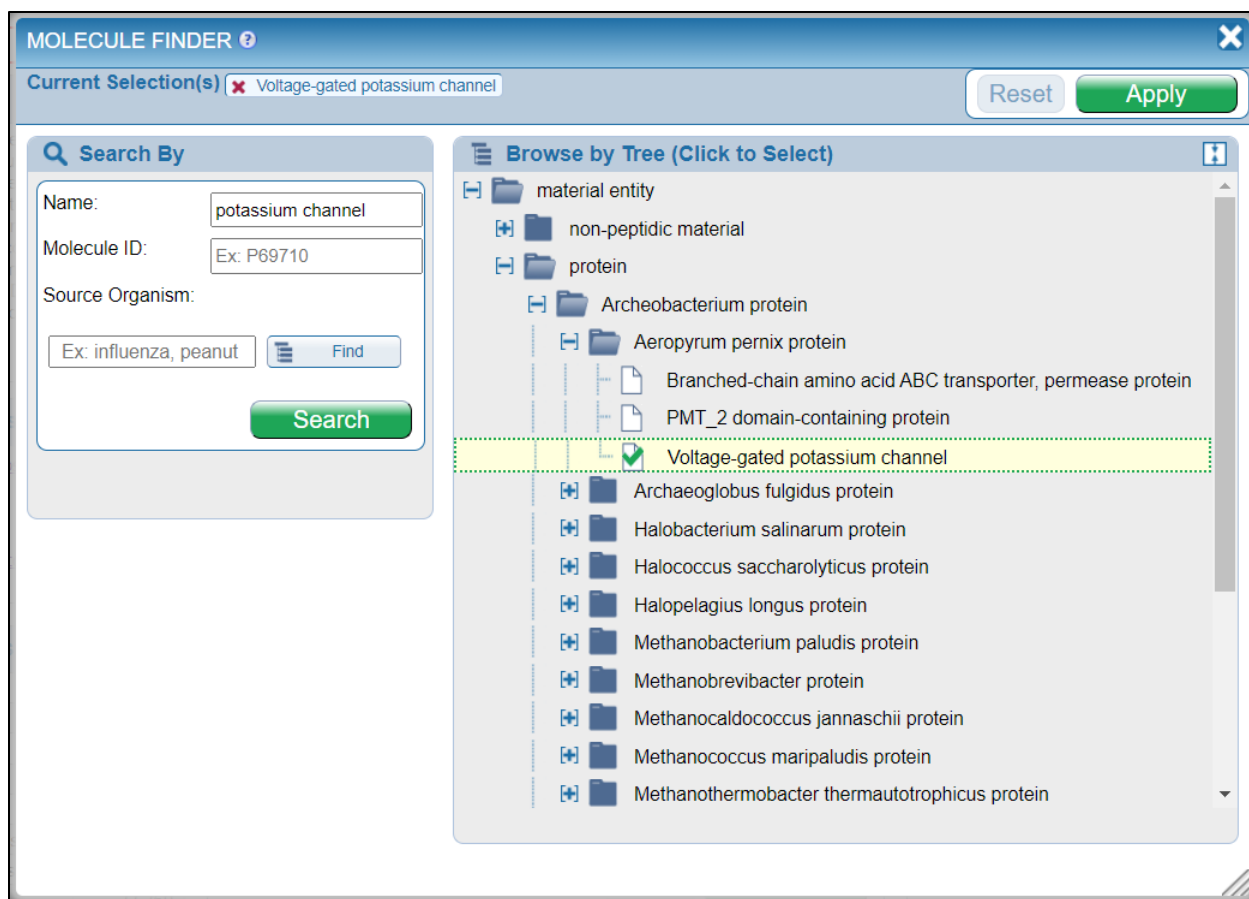


Figure 2.10 An example of the protein branch of the Molecule Finder

As one can see in Figure 2.32, a user can search the molecule tree by entering text, including synonyms, in the Name field in the upper left-hand corner of the finder. The user can also specify the source organism of the molecule of interest using the autocomplete field or the organism finder. For example, this can facilitate the specification of a hemagglutinin is a particular strain of Influenza.

2.2.4.5 Organism Finder

The organism finder is used to facilitate the selection of a species or strain from the NCBI Taxonomy Database. The Organism finder allows the user to find species using their name, synonyms, or taxonomy identifier (assigned by NCBI). When the user performs a search, the system will display a tree structure. The user can search for a name or Organism ID in the "Search By" panel in the upper left-hand portion of the Finder. In the example shown in

Figure 2.11, a search for "dengue" is performed, which results in 47 items being found. The user can select one or more of the items by clicking on the green "+" icon or clicking a branch in the tree. The user can also choose to highlight the item in the tree, as shown in the figure, by clicking the corresponding yellow highlighter icon. Synonyms for a selection are listed next to the organism name column in the search results table. When the Organism Finder is adjacent to a Host Organism field, only the source organisms for which data exist in the IEDB are displayed in the tree structure.

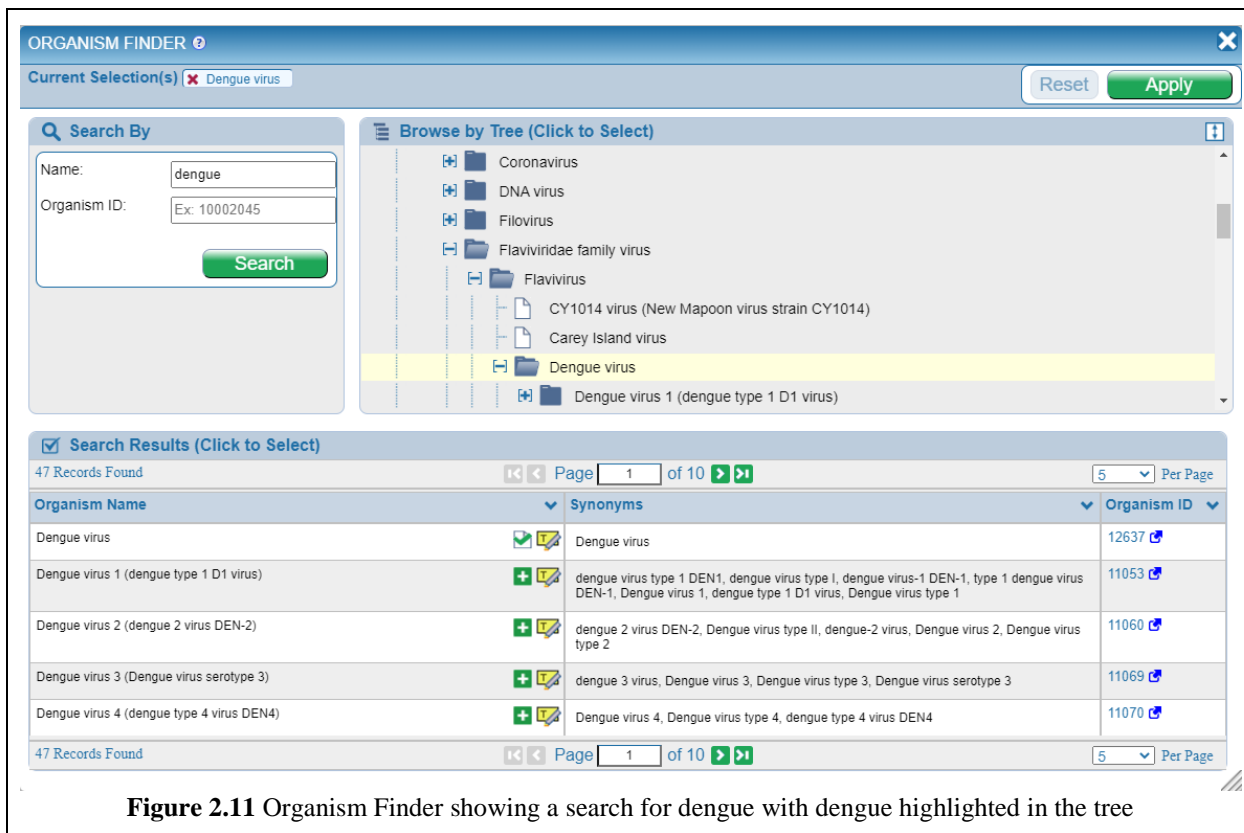


Figure 2.11 Organism Finder showing a search for dengue with dengue highlighted in the tree

2.3 Tools

The Analysis Resource pull-down menu at www.iedb.org accesses the IEDB Analysis Resource and has five items – Analysis Resource Overview, T Cell Epitope Prediction, B Cell Epitope Prediction, Epitope Analysis Tools, and Tool Licensing Information. The purpose of the Analysis Resource of the IEDB is to provide computational tools that enhance the value of the IEDB database to the user. Providing access to tools in one centralized location helps make users aware of available solutions to their problems. All of the information contained within the Analysis Resource, including analysis tools and algorithms developed by the IEDB staff, are freely available to the scientific community. For commercial entities that wish to access the tools in-house, licensing information for the tools can be accessed from the pull-down menu. The Analysis Resource can also be accessed via the rightmost panel on the IEDB home page, although only the most frequently used tools are listed.

The tools provided in the analysis resource fall into two categories – prediction tools and analysis tools. Prediction tools extrapolate beyond data held in the database. They can be used to predict epitopes in protein sequences or predict properties of known epitopes, such as their MHC binding affinity. Analysis tools help extract and interpret data contained in the database.

For prediction tools, it is important to differentiate between the **tool** making predictions, and the **method** used to generate that tool, given a set of training data. For example, the artificial neural network (ANN) method or approach, when trained on a particular data set, will yield a prediction tool. As the ANN method is trained on different data sets, different corresponding prediction tools will result. In this way, the ANN method can be used to develop an MHC class I prediction tool and a separate MHC class II prediction tool. These tools can be refined as more data are available for training. One benefit of the IEDB is that it allows implementing methods to automatically generate new prediction tools as the database grows.

The AR Labs concept was introduced in early 2016. Tools under AR Labs are experimental and are considered to be beta versions, not quite ready for production yet. They are intended for further research, updates and testing. This means they are functional, but the team is still actively developing on improving them and more substantial changes are expected in upcoming releases.

The next subsections will describe each prediction and analytical tool in more detail.

2.3.1 T Cell and B Cell Prediction Tools

Prediction tools can be subdivided into categories by what they predict. The current tools fall into the subcategories listed below:

- T cell epitopes – MHC class I and II binding prediction
- T cell epitopes – Processing prediction
- T cell epitopes – Immunogenicity prediction
- TCR analysis
- B cell epitope prediction
- Structure tools

2.3.1.1 T Cell Epitopes - MHC binding prediction

The Analysis Resource provides tools for predicting peptide binding to MHC class I and II molecules. For class I binding predictions, users can select predictions performed with tools derived from ten different methods – IEDB Recommended, artificial neural network (ANN), stabilized matrix method (SMM), SMM

with a peptide:MHC binding energy covariance matrix (SMMPMBEC), Scoring matrices derived from combinatorial peptide libraries (Comblib_Sidney2008), Consensus, NetMHCpan, PickPocket, NetMHCcons, and NetMHCstabpan. The Consensus method is derived from the ANN, SMM, and CombLib methods. For class II binding predictions, users can select IEDB Recommended, the Combinatorial Library method, the NN_align, the SMM_align method, a method devised by Sturniolo et al. and used in TEPITOPE, NetMHCIIpan, and a consensus method derived from the NN-align, SMM_align, Sturniolo methods, and the Combinatorial Library. Tutorials and example data are available for both the class I and II tools.

A RESTful interface is available for the MHC class I and class II prediction tools. This allows users to perform predictions on the IEDB server in batch mode without having to install any software on their own systems. Additionally, users will always be assured that they are using the latest version of the tools. More information is available in Section 2.3.5. These tools are also available in standalone form on the Download tab and can be downloaded at <http://tools.iedb.org/main/download/>.

2.3.1.1.1 Peptide Binding to MHC Class I Molecules

Users can select from ten different methods for predicting class I epitopes – ANN, SMM, SMMPMBEC, Comblib_Sidney2008, Consensus, NetMHCpan, NetMHCcons, PickPocket, NetMHCstabpan, and IEDB recommended, which are described further below. A check box can be used to show only frequently occurring alleles. This allows the selection of only those alleles that occur in at least 1% of the human population or allele frequency of 1% or higher. However, un-checking the check-box will allow selection of all the alleles and corresponding peptide lengths for a particular species. Users can also upload an allele file instead of entering allele on the page one at a time.

Artificial Neural Network

Artificial neural networks (ANN) are computer algorithms modeled after the brain. They consist of many simple processing units which are wired together in a communication network. Each unit is a simplified model of a neuron which sends off a new signal if it receives a sufficiently strong input signal from the other units to which it is connected. The strength of these connections can be varied in order for the network to perform a desired pattern of node signal activity, which is learned from a set of input training data.

ANNs have been trained for 81 different Human MHC alleles including HLA-A, -B, -C and -E. Furthermore, predictions for 41 animal (Monkey, Cattle, Pig, and Mouse) alleles are available. The current version of ANN is based on NetMHC 4.0. The method is described in Nielsen et al. (Protein Science, 2003) and Andreatta and Nielsen (Bioinformatics, 2016).

Stabilized Matrix Method (SMM)

The Stabilized Matrix Method (SMM) described by Peters and Sette (BMC Bioinformatics, 2005) can be applied to calculate matrices from quantitative affinity data of peptides binding to MHC molecules. The advantage of this method is that it suppresses the noise present in the training data, caused by the inevitable experimental error as well as the limited number of data points.

Stabilized matrix Method with a Peptide:MHC Binding Energy Covariance matrix (SMMPMBEC)

SMMPMBEC is an improved version of SMM. It is different from SMM in that it addresses sparseness of peptide sequence coverage that is often found in binding data sets by using Peptide:MHC Binding Energy Covariance (PMBEC). The PMBEC matrix was derived from experimentally determined binding affinity measurements using combinatorial peptide libraries. SMMPMBEC is described in Kim et al. BMC Bioinformatics 2009.

Scoring matrices derived from combinatorial peptide libraries (Comblib_Sidney2008)

Comblib_Sidney2008 refers to a set of predictors (i.e. scoring matrices) that were derived from binding affinity measurements of combinatorial peptide libraries against a panel of MHC alleles. This work is described in Sidney et al. (Immunome Res. 2008). This class of predictors is unique in that average binding energy contribution of a given residue at a position is directly measured, without worrying about limited peptide sequence coverage.

Consensus

The Consensus predictor was motivated by an idea that predictions made by consulting “consensus” of individual predictions from multiple predictors may result in improved performance over that of any individual ones. For MHC-I, a work describing an early implementation can be found in Moutaftsi et al. (Nat Biotech 2006). The methods used for Consensus are ANN, SMM, and ComLib_Sidney2008. The Consensus method uses as many of these three component methods as possible, depending on their applicability for the chosen allele and length.

NetMHCpan

NetMHCpan version 4.0 predicts binding of peptides to any MHC class I molecule of known sequence using artificial neural networks (ANNs). The method is trained on binding affinity and eluted ligand data leveraging the information from both data types as described in Jurtz et al. (J Immunol, 2017). To train this method, data on all class I MHC ligand elution assays available in IEDB database were collected including the ligand sequence, details of the source protein, position of the ligand in the source protein and the restricting allele of the ligand. There were 160,527 distinct assays in total and the length of the ligands ranged from 4–37. All lengths with a count of ligands at least 0.5% of total ligands were selected for further analysis which included lengths 8–15 and comprised of 99% of the assay entries. After refinement described in Jurtz et al. (J Immunol, 2017), the final training data set had 85,217 entries in total with ligand length ranging from 8 to 15. The ligands originated from 14,797 source antigens and were restricted by 55 unique HLA molecules. Random artificial negatives were generated for each MHC molecule covered by eluted ligand data by sampling randomly $10 \times N$ peptides of each length 8–15 amino acids from the antigen source protein sequences, where N is the number of 9mer ligands for the given MHC molecule.

In 2020, NetMHCpan was updated to version 4.1, as described in Reynisson et al. (Nucleic Acids Res, 2020). The updated version of NetMHCpan differs from its predecessor in two critical aspects: the training data and the machine-learning modeling framework. The training data have been vastly extended by accumulating MHC BA and EL data from the public domain. In particular, EL data were extended to include MA data. The combined dataset used for training of NetMHCpan-4.1 consists of 13,245,212 data points covering 250 distinct MHC class I molecules. The machine learning framework was updated from NNAlign to NNAlign_MA to allow for effective handling of these MA data. In short, the NNAlign framework is a single-allele framework permitting the integration of mixed data types (BA and EL) in the model training, which allows information to be leveraged across the different data types, resulting in a boosted predictive power. NNAlign_MA extends this training framework to allow for the incorporation of EL MA data. This is achieved by iteratively annotating the best single-allele to the MA data during the model training, effectively deconvoluting the MA binding motifs. In 2020, NetMHCpan EL 4.1 became the ‘IEDB recommended’ method based on benchmarking results.

PickPocket

PickPocket predicts binding of peptides to any known MHC molecule using position specific weight matrices. The method is trained on more than 150,000 quantitative binding data covering more than 150 different MHC molecules. Predictions can be made for HLA-A, B, C, E and G alleles, as well as for non-human primates, mouse, cattle, and pig. Version 1.1 has been retrained on extensive data sets, including 10 prevalent HLA-C and 7 prevalent BoLA MHC-I molecules. The matrices of pocket-library are generated using the SMM_pmbec method described in Kim et al. (BMC Bioinformatics, 2009). Predictions can be made for 8 – 12mer peptides. All non-9mer predictions are made using approximations. The prediction values are given in nM IC50 values. The algorithm is described in Zhang et al. (Bioinformatics, 2009).

NetMHCcons

NetMHCcons predicts binding of peptides to any known MHC class I molecule. This is a consensus method for MHC class I predictions integrating three state-of-the-art methods, NetMHC, NetMHCpan, and PickPocket, to give the most accurate predictions. NetMHC is an artificial neural network-based (ANN) allele-specific method which has been trained using 94 MHC class I alleles. Version 3.4 is used as part of NetMHCcons-1.1. NetMHCpan is a pan-specific ANN method trained on more than 115,000 quantitative binding data covering more than 120 different MHC molecules. Version 2.8 is used as part of NetMHCcons-1.1. PickPocket Version 1.1 is described above. NetMHCcons can generate predictions for peptides of 8 – 15 residues in length. The prediction values are given in nM IC50 values. The method is described in detail in Karosiene et al. (Immunogenetics, 2012).

NetMHCstabpan

NetMHCstabpan was introduced in 2016. It predicts binding stability of peptides to any known MHC molecule using artificial neural networks (ANNs). The method is trained on more than 25,000 quantitative stability data covering 75 different HLA molecules. The user can upload full length MHC protein sequences, and have the server predict MHC restricted peptides from any given protein of interest. Predictions can be made for 8-14 mer peptides, although all non 9mer predictions are made using approximations. Most HLA molecules have a strong preference for binding 9mers. The prediction values are given in half life time in hours values and as %-Rank to a set of 200,000 random natural peptides. The method is described in Rasmussen et al. (Journal of Immunology, 2016).

IEDB Recommended

IEDB recommended is the default prediction method selection. Prior to 2020, IEDB recommended considered all alleles and their corresponding peptide lengths for a particular species. For each allele-length combination, consensus method was used, which includes ANN, SMM, and ComLib. However, as of 2020, NetMHCpan EL 4.1 became the ‘IEDB recommended’ method based on benchmarking results.

2.3.1.1.2 Peptide Binding to MHC Class II Molecules

Users can select from seven different methods for predicting class II epitopes – IEDB recommended, SMM-align, Sturniolo, Combinatorial Library, Consensus, NN-align, and NetMHCIIpan. By default, the overall best method (IEDB recommended) is selected. However, not all methods can currently make predictions for all alleles, so only the alleles available will be displayed. The seven methods are described further below.

SMM-align

The MHC class II binding groove is open at both ends making the correct alignment of a peptide in the binding groove a crucial part of identifying the core of an MHC class II binding motif. The stabilization matrix alignment method, SMM-align, allows for direct prediction of peptide:MHC binding affinities. The method uses amino terminal peptide flanking residues (PFR) to get a consistent gain in predictive performance by favoring binding registers with a minimum PFR length of two amino acids. The method predicts quantitative peptide:MHC binding affinity values. The method has been trained and evaluated on

a data set that covers the nine HLA-DR supertypes suggested and three mouse H2-IA allele. The method is described by Nielsen et al. (BMC Bioinformatics, 2007).

Sturniolo

This matrix-based approach is used in the TEPITOPE class II epitope prediction program. It is described in Sturniolo et al. (Nat. Biotechnol., 1999).

Combinatorial Library

The positional scanning combinatorial libraries approach utilized a pool of random peptide libraries to systematically measure the contribution to MHC binding from each amino acid at each of the nine positions at the binding peptide. Each pool in the library contains 9-mer peptides with one fixed residue at a single position. With each of the 20 naturally occurring residues represented at each position along the 9-mer backbone, the entire library consisted of 180 peptide mixtures. Competitive binding assays were then carried out to determine the IC₅₀ values for each pool. IC₅₀ values for each mixture were standardized as a ratio to the geometric mean IC₅₀ value of the entire set of 180 mixtures, and then normalized at each position so that the value associated with the optimal value at each position corresponds to 1. For each position, an average (geometric) relative binding affinity (ARB) was calculated, and then the ratio of the ARB for the entire library to the ARB for each position was derived. The final results are a set of 9 by 20 scoring matrices which could predict the binding of novel peptides to MHC molecules.

The methodology is the same as that used for the MHC class I combinatorial library tool as described in Sidney et al. Immunome Res. 2008. A description of the ARB method is contained in Bui et al., Immunogenetics, 2005.

Consensus

The consensus method was developed by the IEDB team by exploiting features of three aforementioned methods. The Consensus method uses NN-align, SMM-align, and the combinatorial peptide scanning library. When the scanning library is not available for an allele, the Sturniolo method is used instead. A paper describing the original method was published by Wang et al. (PLoS Comput Biol, 2008), and a subsequent paper was published by Wang et al. (BMC Bioinformatics, 2010). The datasets used in assessing the performance of the NN-align, SMM-align, and Sturniolo methods and in developing the Consensus method, as described in Wang et al. (2010), can be found at <http://tools.iedb.org/main/datasets/>.

NN-align

NN-align is an artificial neural network-based alignment algorithm for MHC class II peptide binding prediction. It simultaneously identifies the MHC class II binding core and binding affinity. The method is trained using an algorithm that corrects bias in the training data caused by redundancy in binding core representation. Prediction accuracy has been shown to improve significantly when information about the residues flanking the peptide-binding core is considered. A 2009 paper in BMC Bioinformatics by Nielsen and Lund describes the method in detail (PMID: 19765293).

NetMHCIIpan

NetMHCIIpan predicts binding of peptides for all three human MHC class II isotypes- HLA-DR, HLA-DP, and HLA-DQ, in addition to mouse H-2-I molecules. A paper describing the method was published by Nielsen et al. in PLoS Computational Biology, 2008, and a paper describing the latest version 3.1 was published by Andreatta et al. in Immunogenetics, 2015.

In 2022, NetMHCIIpan was updated to version 4.1, as described in Kaabinejadian et al. (Front Immunol., 2022). The combined dataset used for training of NetMHCIIpan-4.0 consists of 4 086 230 data points covering a total of 116 distinct MHC class II molecules. The machine learning framework was updated from NNAlign to NNAlign_MA to allow for effective handling of these MA data. In short, the NNAlign

framework is a single-allele framework permitting the integration of mixed data types (BA and EL) in the model training, which allows information to be leveraged across the different data types, resulting in a boosted predictive power. NNAlign_MA extends this training framework to allow for the incorporation of EL MA data. This is achieved by iteratively annotating the best single-allele to the MA data during the model training, effectively deconvoluting the MA binding motifs.

IEDB Recommended

IEDB recommended 2.22 is the default prediction method selection. It considers all alleles and their corresponding peptide lengths for a particular species. For each allele-length combination, consensus method is used, as described above. If none of these methods are available for the allele, NetMHCIIpan is used.

2.3.1.1.3 Tepitool

The tool is designed as a wizard for the prediction of T cell epitope candidates from a given set of amino acid sequences, based on predicted peptide binding to MHC class I and class II molecules, where the user is led through a series of well-defined steps to complete the task. In contrast to the independent methods described above, Tepitool guides the user step-by-step as a client-side web form that takes user input data that is in turn processed at the server-side when the user submits the entire form. All fields except sequences and alleles are filled with default recommended settings for prediction and selection of optimum peptides. The input parameters can be adjusted as per the user's specific needs, and the user can go back to previous steps to change the selection before final submission of the job. The TepiTool has six steps:

- (i) Provide sequence data.
- (ii) Select the host species and MHC allele class.
- (iii) Select the alleles for binding prediction.
- (iv) Select peptides to be included in prediction.
- (v) Select preferred methods for binding prediction and peptide selection and cutoff values.
- (vi) Review selections, enter job details and submit data.

The tool is described in Paul et al. (Curr. Protoc. Immunol., 2016).

2.3.1.2 T Cell Epitopes – MHC Processing Prediction

2.3.1.2.1 Proteasomal cleavage/TAP transport/MHC class I combined predictor

For the prediction of antigen processing through the MHC class I antigen presentation pathway, we incorporated predictions of proteasomal cleavage and TAP transport similar to the MHCPathway website described in (Tenzer et al, CMLS, 2005). The predictions are based on in vitro experiments characterizing the sequence specificity of proteasomal cleavage and TAP transport. The goal of the prediction is to identify MHC-I ligands (peptides that are naturally processed from their source proteins and presented by MHC class I molecules).

The proteasomal cleavage predictions evaluate how efficiently a peptide or its N-terminally prolonged precursors can be liberated from its source protein. The TAP transport predictions evaluate how efficiently a peptide or its N-terminal prolonged precursors are transported into the ER by TAP (Peters et al., Immunol, 2003). When this information is taken together and combined with MHC class I binding predictions, the tool yields a prediction of the efficiency with which a peptide is presented on the cell surface. The methods available are ANN, SMM, SMMPMBEC, Comblib_Sidney2008, NetMHCpan, PickPocket, NetMHCcons, and IEDB Recommended, which are described in Section 2.3.1.1.1.

A check box can be used to show only frequently occurring alleles. This allows the selection of only those alleles that occur in at least 1% of the human population or allele frequency of 1% or higher. However, un-

checking the check-box will allow selection of all the alleles and corresponding peptide lengths for a particular species. Users can also upload an allele file instead of entering allele on the page one at a time.

2.3.1.2.2 *Neural network-based prediction of proteasomal cleavage sites (NetChop) and T cell epitopes (NetCTL/NetCTLpan)*

NetChop produces neural network predictions for cleavage sites of the human proteasome (Kesmir et al., 2002). NetChop considers the characteristics of the structurally modified proteasomes found in cells stimulated by gamma-interferon under physiological conditions. The NetChop algorithm was trained on in vitro data and MHC Class I ligand data. The use of this training set, combined with the artificial neural network methodology, makes the prediction of cleavage sites more accurate. NetChop has been trained only on human data, but since the proteasome structure is quite conserved, the algorithm developers believe that the tool is capable of making reliable predictions for at least the other mammalian proteasomes.

NetCTL predicts CTL epitopes in protein sequences integrating prediction of peptide MHC binding, proteasomal C terminal cleavage and TAP transport efficiency. The method is described in detail in Larsen et al. (Eur J Immunol., 2005). NetCTLpan is an update to the original NetCTL server that allows for prediction of CTL epitope with restriction to any MHC molecules of known protein sequence (Stranzl et al., Immunogenetics, 2010). This tool is also available in a standalone form on the Download tab and can be downloaded at <http://tools.iedb.org/main/download/>.

2.3.1.2.3 *MHC-NP: Prediction of peptides naturally processed by the MHC*

MHC-NP is a tool for predicting peptides naturally processed by the MHC pathway and was introduced to the Analysis Resource in 2013. MHC-NP employs data obtained from MHC elution experiments in order to assess the probability that a given peptide is naturally processed and binds to a given MHC molecule. This tool was the winner of the 2nd Machine Learning Competition in Immunology, where it yielded state-of-the-art accuracy for the prediction of peptides eluted from human HLA-A*02:01, HLA-B*07:02, HLA-B*35:01, HLA-B*44:03, HLA-B*53:01, HLA-B*57:01 and mouse H2-D(b) and H2-K(b) MHC molecules. The method is described in Giguère et al. (J Immunol Methods, 2013). Because the tool was developed by researchers external to the IEDB team, it is not included in the virtual machine image.

2.3.1.2.4 *MHC-II-NP: Prediction of peptides naturally processed by the MHC*

Although algorithms for the prediction of peptide binding affinity to MHC II molecules exist, there is a lack of methods that predict the ligands resulting from natural antigen processing. To address this gap MHC-II-NP was developed.

The training ligand dataset, obtained through IEDB database query, contained more than 14,000 naturally processed ligands identified by MS of peptides eluted from MHC class II-expressing cells (from approximately 2,600 unique protein sequences). Analysis into sequence signatures which liberate peptides from their source antigens revealed preferred amino acids surrounding both N- and C- ligand terminuses. These cleavage motifs were used to develop this method for predicting naturally processed MHC II ligands as described in Paul et al. (Front. Immunol., 2018).

2.3.1.3 *T Cell Class I pMHC Immunogenicity Predictor*

An MHC class I immunogenicity tool was introduced in 2013. T-cells have to recognize peptides presented on MHC molecules to be activated and elicit their effector functions. Several studies have demonstrated that some peptides are more immunogenic than others and therefore more likely to be T-cell epitopes. This tool is based on an analysis of a large set of data describing the immunogenicity of peptides presented on various MHC-I molecules.

Two main conclusions could be drawn from this analysis. First, in line with previous observations, the developers showed that positions P4-6 of a presented peptide are more important for immunogenicity. Second, some amino acids, especially those with large and aromatic side chains, are associated with immunogenicity. This information was combined into a simple model that was used to demonstrate that immunogenicity is, to a certain extent, predictable. This model was validated with data from two independent epitope discovery studies. This model indicated that T-cells are equipped to better recognize viral than human (self) peptides. The method is described in Calis et al. (PLoS Comput Biol., 2013). This tool is also available in a standalone form on the Download tab and can be downloaded at <http://tools.iedb.org/main/download/>.

2.3.1.4 Deimmunization

The deimmunization tool is attempt to identify immunodominant regions in a given therapeutically important protein, and suggest amino-acid substitutions that create non-immunogenic versions of the proteins. Therefore, we have opted for a two steps process; 1) In the first step, the deimmunization tool will list all the immunogenic regions or peptides based on selected threshold. These peptides will be generated from the protein with 15mer window size and 10mer overlap. 2) In the second step, the user can select one or more peptides listed in the results and final result window will display the non-immunogenic substitution of each selected peptides. The default threshold is 8.5 (which is difference in the median of percentile rank from 26 reference alleles set for MHC class II). In the final result window, the tools will also take care of the fact that non-immunogenic substitution in the immunogenic peptides, should not create new immunogenic site in the neighboring peptides. Therefore, the result window will also display the effect of substitution on the neighboring peptides. This tool is described in Dhanda et al. (Immunology, 2018).

2.3.1.5 CD4 T cell immunogenicity

The server is developed to predict the allele independent CD4 T cell immunogenicity at population level, overcoming complications related to the high level of variability of HLA molecules, potential other factors beyond HLA as well as a frequent lack of HLA typing data. With this tool, users can predict the T cell immunogenicity using 7-allele method (Paul et. al. 2015), immunogenicity method and combined method (IEDB recommended). The combined method predicts the final score that combines the predictions from 7-allele method and immunogenicity method. This tool is described in Dhanda et al. (Front Immunol, 2018).

2.3.1.6 Antigen Expression Based Epitope Likelihood-Function (Axel-F)

AXEL-F incorporates antigen abundance estimates with MHC binding predictions to enhance epitope predictions. This tool is described in Koşaloğlu-Yalçın et al. (iScience, 2020).

2.3.2 TCR Analysis

2.3.2.1 TCRMatch

TCRMatch compares input CDR3b sequences against curated CDR3b sequences in the IEDB to find matches that are predicted to share epitope specificity. Matches are determined by sequence similarity, which is scored using a comprehensive k-mer comparison. This tool is described in Chronister et al. (Front Immunol, 2021).

2.3.3 B Cell Epitope Prediction

2.3.3.1 Prediction of linear epitopes from protein sequence

Six different tools are provided that predict antibody epitope candidates from amino acid sequences. Five are based on amino acid property scales and a sixth method uses a Hidden Markov Model. Parameters such as hydrophilicity, flexibility, accessibility, and antigenic propensity of polypeptides chains have been correlated with the location of continuous epitopes in a few well-characterized proteins. Based on these observations, amino acid property scales have been developed to predict antigenic determinants. Each scale consists of 20 values assigned to each of the amino acid residues on the basis of their relative propensity to possess the property described by the scale. The following amino acid property scales have been selected and implemented based on their popularity and coverage of different categories.

- Secondary structure - Chou and Fasman beta turn prediction
- Surface exposure - Emini surface accessibility prediction
- Flexibility - Karplus and Schulz flexibility prediction
- Antigenicity - Kolaskar and Tongaonkar antigenicity prediction
- Hydrophobicity/hydrophilicity - Parker hydrophilicity prediction

BepiPred combines the predictions of a hidden Markov model and the propensity scale of Parker et al. (Biochemistry, 1986). It is described in Larsen et al. (Immunome Research, 2006). The linear B cell epitope predictor is also available as a standalone tool. In 2017, BepiPred 2.0 was developed and published (Nucleic Acids Res, 2017), PMID: 28472356. BepiPred-2.0 uses structural predictions, such as surface accessibility and secondary structure as part of the feature space. Using a Random Forest machine-learning algorithm trained on structural epitopes obtained from 160 3D-structures, they improved the performance significantly compared to the prior version and other state-of-the-art prediction tools.

2.3.3.2 DiscoTope - Prediction of epitopes from protein structure

DiscoTope was augmented in 2013 to include version 2.0 as well as version 1.1. DiscoTope is designed specifically to predict discontinuous epitopes. It uses protein three-dimensional structural data in addition sequence data. The method is based on amino acid statistics, spatial information, and surface accessibility in a compiled data set of discontinuous epitopes determined by X-ray crystallography of antibody/antigen protein complexes. DiscoTope 2.0 works with a novel definition of the spatial neighborhood used to sum propensity scores and half-sphere exposure as a surface measure. The results are more accurate than version 1.1 but involve more complex calculations that increase run times by a factor of approximately 30.

DiscoTope 1.1 is described in Haste Andersen et al. (Protein Sci., 2006), and DiscoTope 2.0 is described in Kringelum et al. (PLoS Comp. Bio, 2012).

2.3.3.3 ElliPro - Epitope prediction based upon structural protrusion

ElliPro predicts linear and discontinuous antibody epitopes based on a protein antigen's 3D structure. ElliPro accepts either a protein structure (preferred) or a protein sequence as an input. If a protein sequence is used, ElliPro will predict its 3D structure by homology modeling. Its use is described in the Tutorial tab of the ElliPro section of the Analysis Resource. The method is described in Julia Ponomarenko et al. (BMC Bioinformatics, 2008). It is also available as a standalone tool.

2.3.3.4 Methods for modeling and docking of antibody and protein 3D structures

This web page provides information on methods for modeling and docking of antibody and protein 3D structures. The methods are available as web servers and are hosted outside the IEDB except one method, LYRA, described above. One of the approaches at B-cell epitope prediction involves antibody-antigen docking, or 3D modeling of an antibody-antigen complex, for known 3D structures of both antibody and

antigen. When the structures of either antibody or protein or both are unknown (that is, not available in PDB) but sequences are known, the user can attempt to obtain the 3D structural models of the antibody and the antigen, following the chart displayed on the web page and using the described web servers. Links are provided to these servers for the user's convenience.

2.3.4 Structure Tools

2.3.4.1 LYmphocyte Receptor Automated modeling (LYRA)

The LYRA web server implements a complete and automated method for building of B- and T-cell receptor structural models starting from their amino acid sequence alone. The webservice is freely available and easy to use for non-specialists. Upon submission, LYRA automatically generates alignments using ad hoc profiles, predicts the structural class of each hypervariable loop, selects the best templates in an automatic fashion, and provides within minutes a complete 3D model that can be downloaded or inspected online. Experienced users can manually select or exclude template structures according to case specific information. LYRA is based on the canonical structure method, that in the last 30 years has been successfully used to generate antibody models of high accuracy, and in our benchmarks this approach proves to achieve similarly good results on TCR modeling, with a benchmarked average RMSD accuracy of 1.29 and 1.48 Å for B- and T-cell receptors, respectively. The tool is described in detail by Klausen et al. (Nucleic Acids Research, 2015).

2.3.4.2 Structural Complexes of Epitope Receptor (SCEptRe)

SCEptRe provides weekly updated, non-redundant, user customized benchmark datasets with information on the immune receptor features for receptor-specific epitope predictions. This tool extracts weekly updated 3D complexes of antibody-antigen, TCR-pMHC and MHC-ligand from the Immune Epitope Database (IEDB) and clusters them based on antigens, receptors and epitopes to generate benchmark datasets. Users can customize structural quality and clustering parameters (e.g., resolution, R free factors, antigen or epitope sequence identity) to generate these datasets based on their need. The tool is described in detail by Mahajan et al. (BMC Bioinformatics, 2019).

2.3.4.3 Docktope

Docktope is a web-based tool, based on the D1-EM-D2 approach, intended to allow the pMHC-I modeling. Some applications allowed from the pMHC-I construction include the study of the pMHC-I tridimensional structure, epitope-MHC-I interaction patterns, cross-reactivity assessment and molecular dynamics studies. The tool allows the user to construct pMHC-I structures for the following alleles: HLA-A*02:01 (9-mer epitopes), HLA-B*27:05 (9-mer epitopes), H2-Db (9-mer/10-mer epitopes) and H2-Kb (8-mer epitopes). The tool is described in detail by Rigo et al. (Scientific Reports, 2015).

2.3.5 Epitope Analysis Tools

2.3.5.1 Population Coverage

T cells recognize a complex between a specific MHC type and a particular pathogen-derived epitope and thus a given epitope will elicit a response only in individuals that express an MHC molecule capable of binding that particular epitope. MHC molecules are extremely polymorphic (over a thousand different variants are known in humans). Therefore, selecting multiple peptides with different MHC binding specificities will afford increased coverage of the patient population targeted as vaccine recipients. The issue of population coverage in relation to MHC polymorphism is further complicated by the fact that different MHC types are expressed at dramatically different frequencies in different ethnicities. Thus, without careful consideration, a vaccine with ethnically biased population coverage could result. To address this issue, the actual/predicted binding capacity of potential epitopes to as many different MHC molecules possible (and when available, also restriction data of T cell responses recognizing the epitope) can be used

to project the population coverage in different ethnicities of different vaccine candidates or epitope sets. Accordingly, epitope-based vaccines or diagnostics can be designed to maximize population coverage, while minimizing complexity (that is, the number of different epitopes included in the diagnostic or vaccine), and also minimizing the variability of coverage obtained or projected in different ethnic groups.

An important consideration in the process of epitope selection is that the patient population coverage afforded by a given set is not simply corresponding to the sum of the coverage of its individual components. Thus, to calculate the coverage afforded by a given mixture of epitopes, a more comprehensive approach and a suitable algorithm has been developed for this specific purpose (Bui et al. BMC Bioinformatics 2006). This method calculates the fraction of individuals predicted to respond to a given epitope set on the basis of HLA genotypic frequencies, assuming non-linkage disequilibrium between HLA loci, and on the basis of MHC binding and/or T cell restriction data. The algorithm is briefly explained here. First, genotypic frequencies of various MHC are tabulated. Each time a peptide binds to a given MHC, a “hit” is recorded for that MHC. The process is repeated for all peptides. Then the hits for MHC are tallied. Next, the frequency of each possible diploid MHC combination (phenotype) is calculated. For n MHC types, this corresponds to an $n \times n$ tabulation of the frequency at which each specific pair of MHCs will be found in the population from which the MHC frequencies are derived. A similar table is generated to contain the number of hits per each of the MHC combinations by adding the number of hits associated with each of the two alleles of MHC in the combination (a simple exception is the case of homozygous combinations, where the number of hits is simply the number of hits of the given MHC). From these two tables, a frequency distribution is assembled, tabulating the genotypic frequency of all MHC combinations associated with a certain number of hits. The result of the analysis is displayed as a frequency distribution histogram and a cumulative frequency plot.

HLA allele genotypic frequencies were obtained from Allele Frequency Net Database (<http://www.allelefreqencies.net/>). At present, Allele Frequency Net Database (Gonzalez-Galarza FF et al., Nucleic Acids Res. 2015) provides allele frequencies for 115 countries and 20 different ethnicities grouped into 16 different geographical areas. In addition, the program also accepts custom populations with allele frequencies defined by users. Multiple population coverages can be simultaneously calculated and an average population coverage is generated. Since MHC class I and MHC class II restricted T cell epitopes elicit immune responses from two different T cell populations (CTL and HTL, respectively), the program provides three calculation options to accommodate different coverage modes: (1) class I separate, (2) class II separate, and (3) class I and class II combined. For each population coverage, the tool computes the following: (1) projected population coverage, (2) average number of epitope hits / HLA combinations recognized by the population, and (3) minimum number of epitope hits / HLA combinations recognized by 90% of the population (PC90).

2.3.5.2 Epitope Conservancy

In a diagnostic or epitope-based vaccine setting, focusing on conserved epitopes allows for targeting responses around pathogen variability, whether it exists prior to infection, or develops in the natural course of disease. The use of conserved epitopes would be expected to focus the immune response on sequences crucial for retaining biological function of the pathogen proteins, and thus with intrinsically lower variability, even under immune pressure. The epitope conservancy analysis tools implemented here aims to address the issue of variability (or conservation) of epitopes, and to assist in the selection of epitopes with the desired pattern of conservation. The algorithm has been implemented to calculate the degree of conservancy of an epitope within a given protein sequence set at different degree of identities. The degree of conservation is defined as the number of protein sequences that contain the epitope at a given identity level, divided by the total number of protein sequences found in the dataset analyzed (Bui et al. BMC Bioinformatics 2007).

Users can now also indicate whether they want duplicated protein sequences removed from the results set by checking a box in the “Specify calculation options” section of the input page. Previously redundant results were removed. For users wanting to study different populations, keeping redundant sequences can be very informative.

2.3.5.3 Epitope Cluster Analysis

Version 2.0 of the Epitope Cluster Analysis tool (Dhanda et al., Immunology. 2018) was released in 2018. This new version expanded tool functionality to include graphical representation, to provide connectivity within a cluster, to generate epitope clusters based on representative or consensus sequences, and to calculate overhang sequence identity. The base functionality of the tool remained the same.

The tool groups epitopes into clusters based on sequence identity. A cluster is defined as a group of sequences that has a sequence similarity greater than the minimum sequence identity threshold specified. Epitope sequences can be either directly entered in the text area or uploaded from a file. Two acceptable sequence formats are PLAIN and FASTA. The user can select the sequence identity threshold at which they want to calculate epitope clusters. Clusters are displayed in a table format where clusters are indicated by table rows which have the same color.

2.3.5.4 Computational Methods for Mapping Mimotopes to Protein Antigens

A web page devised by Dr. Julia Ponomarenko was added to the Analysis Tools section in 2014 that provides guidelines for mapping mimotopes to protein antigens. Mimotopes are molecules, often peptides, which are recognized by antibodies or T cells that were primed to recognize a different epitope. For antibody epitope mapping, peptide mimotopes can be selected from a combinatorial library (usually peptides from a specific protein antigen) on binding to a specific monoclonal antibody with high affinity. They are commonly obtained using phage display. Experimental data on mimotopes can be found in the IEDB, MimoDB (<http://immunet.cn/mimodb/index.html>), and journal publications. Mimotopes can mimic essential features of genuine epitopes and therefore their mapping to the antigen is a logical and essential step of the mimotope analysis. This web page provides information on available methods for mimotope mapping, how to search the IEDB for mimotopes, and an example of a mimotope dataset and the results of its mapping, using the available web servers hosted outside the IEDB.

Methods available as web servers and operational as of July 2014 (they all require the user submitting an antigen protein 3D structure, providing either PDB ID of the structure or the pdb file) are as follows:

- **MIMOX** (<http://immunet.cn/mimox/>)
- **Pepitope server** (<http://pepitope.tau.ac.il/>) provides three algorithms: Mapitope, PepSurf, or combined
- **EpiSearch** (<http://curie.utmb.edu/episearch.html>)

The web page also lists other published methods that are unavailable as web servers. They include the following:

- **PepMapper** (<http://www.ncbi.nlm.nih.gov/pubmed/22701536>) (2012) - provided URL didn't respond
- **MimoPro** (<http://www.ncbi.nlm.nih.gov/pubmed/21609501>) (2011) - provided URL didn't respond
- **Pep-3D-Search** (<http://www.ncbi.nlm.nih.gov/pubmed/19087303>) (2008) - available for download at <http://kyc.nenu.edu.cn/Pep3DSearch/>
- **MEPS** (<http://www.ncbi.nlm.nih.gov/pubmed/17430573>) (2007)
- **MIMOP** (<http://www.ncbi.nlm.nih.gov/pubmed/16434442>) (2006)

- **3D-Epitope-Explorer (3DEX)** (<http://www.ncbi.nlm.nih.gov/pubmed/15834923>) (2005) - available for download at <http://www.schreiber-abc.com/3dex/>
- **Enshell-Seijffers et al.** (<http://www.ncbi.nlm.nih.gov/pubmed/14596802>) (2003)
- **FINDMAP** (<http://www.ncbi.nlm.nih.gov/pubmed/12935344>) (2003)
- **SiteLight** (<http://www.ncbi.nlm.nih.gov/pubmed/12824481>) (2003)

The webpage also contains instructions on how to search for mimotopes in the IEDB.

2.3.5.5 Restrictor Analysis Tool for Epitopes (RATE)

RATE is an automated method that can infer HLA restriction for a set of given epitopes from large datasets of T cell responses in HLA typed subjects. The tool takes two data files, one containing the alleles expressed by the subjects and the other containing the response of the peptides in the subjects. The tool calculates the odds ratio and estimates its significance using Fisher's exact test. It also calculates a parameter called relative frequency similar to odds ratio. The tool was developed with a focus on class II alleles but can also be applied to class I alleles. A description of the tool can be found in Paul et al. (Journal of Immunology, 2015).

In 2017, RATE was validated using experimentally defined restriction data from a set of 191 tuberculosis-derived epitopes and 63 healthy individuals with MTB infection from the Western Cape Region of South Africa. Additionally, the parameters used by the RATE tool to infer restriction were optimized. New HLA restrictions were identified using the optimized RATE tool. A description of this study can be found in Paul et al. (BMC Immunol, 2017).

2.3.5.6 ImmunomeBrowser

This tool aggregates all data relevant to the user query and allows one to visualize the known immune response to a specific antigen, as well as illustrating knowledge gaps in a reference protein. It provides the immune reactivity in terms of response frequency (RF) and the number of subjects tested/responded and/or number of independent assays performed along the length of reference protein. The tool was originally implemented in the results page of the database section of the IEDB. To further extend the usability to predicted epitopes and propriety epitopes or non-IEDB data, a stand-alone version of the tool was developed in 2018 (Dhanda et. al., Bioinformatics 2018). The standalone version maps user provided peptide sets and associated response data to a user-provided protein reference sequence. This now allows the user to analyze and visualize immunodominant regions within their own dataset. The results are presented both as a user interactive JavaScript based web interface and a tabular format in a selected reference sequence.

2.3.5.7 PepSySco

Given a set of peptide sequences, Peptide Synthesis Score (PepSySco) predicts the likelihood that they can be synthesized successfully. The tool is described in detail by Gutman et al. (ACS Omega, 2022).

2.3.5.8 PepX (Peptide Expression Annotation)

This tool identifies from which proteins a list of peptides can be derived, and returns an estimate of the expression level of those peptides from selected public databases.

2.3.6 IEDB Analysis Resource Labs

The AR Labs concept was introduced in early 2016. Tools designated as within AR Labs are experimental and are considered to be beta versions, not quite ready for production yet. They are intended for further research, updates and testing. This means they are functional, but the team is still actively developing on improving them and more substantial changes are expected in upcoming releases. Therefore, users are asked not to expect the same level of stability for these tools as for other tools in the IEDB analysis resource. Users are welcome to send bug reports and suggestions for improvements for these tools via the help desk.

Of the tools described above, 11 tools are currently available under the Labs designation:

Prediction LABS Tools

MHCII-NP
Deimmunization
CD4 T Cell Immunogenicity
LYRA
SCEptRe
Docktope
Axel-F
TCRMatch

Analysis LABS Tools

RATE
ImmunomeBrowser
PepSySco
PepX

2.3.7 Benchmark References and Data Sets

The “Datasets” tab contains data sets and references related to benchmarking results of the IEDB’s epitope prediction tools. There are three accordion sliders that can be expanded to reveal details. The three are for MHC class I binding prediction, MHC class II binding prediction, and B cell epitope prediction. In each case, the journal article title, authors, journal name, and year are listed along with a description of the data set, the date of its generation, details on the data set generation, the data format, and where the data set is available.

For the MHC class I binding predictions, the references are:

- Reynisson B, Alvarez B, Paul S, Peters B, Nielsen M. NetMHCpan-4.1 and NetMHCIIpan-4.0: improved predictions of MHC antigen presentation by concurrent motif deconvolution and integration of MS MHC eluted ligand data. *Nucleic Acids Res.* 2020 Jul 2;48(W1):W449-W454. doi: 10.1093/nar/gkaa379. PMID: 32406916; PMCID: PMC7319546.
- Jurtz V, Paul S, Andreatta M, Marcatili P, Peters B, Nielsen M. NetMHCpan-4.0: Improved Peptide-MHC Class I Interaction Predictions Integrating Eluted Ligand and Peptide Binding Affinity Data. *J Immunol.* 2017 Nov 1;199(9):3360-3368. doi: 10.4049/jimmunol.1700893. Epub 2017 Oct 4. PMID: 28978689; PMCID: PMC5679736.
- Trolle T, Metushi IG, Greenbaum JA, Kim Y, Sidney J, Lund O, Sette A, Peters B, Nielsen M. Automated benchmarking of peptide-MHC class I binding predictions. *Bioinformatics.* 2015 Jul 1;31(13):2174-81. doi: 10.1093/bioinformatics/btv123. Epub 2015 Feb 25. PMID: 25717196; PMCID: PMC4481849
- Kim Y, Sidney J, Buus S, Sette A, Nielsen M, Peters B. Dataset size and composition impact the reliability of performance benchmarks for peptide-MHC binding predictions. *BMC Bioinformatics.* 2014 Jul 14;15:241. doi: 10.1186/1471-2105-15-241. PMID: 25017736; PMCID: PMC4111843.
- Kim Y, Sidney J, Pinilla C, Sette A, Peters B. Derivation of an amino acid similarity matrix for peptide: MHC binding and its application as a Bayesian prior. *BMC Bioinformatics.* 2009 Nov 30;10:394. PMID: 19948066.

- Peters B, Bui HH, Frankild S, Nielson M, Lundegaard C, Kostem E, Basch D, Lamberth K, Harndahl M, Fleri W, Wilson SS, Sidney J, Lund O, Buus S, Sette A., A community resource benchmarking predictions of peptide binding to MHC-I molecules, *PLoS Comput Biol.* 2006 Jun 9;2(6):e65. Epub 2006 Jun 9. PMID: 16789818

For the MHC class II binding predictions, the references are:

- Kaabinejadian S, Barra C, Alvarez B, Yari H, Hildebrand WH, Nielsen M. 2022. Accurate MHC Motif Deconvolution of Immunopeptidomics Data Reveals a Significant Contribution of DRB3, 4 and 5 to the Total DR Immunopeptidome. *Front Immunol.* 13:835454.
- Reynisson B, Alvarez B, Paul S, Peters B, Nielsen M. NetMHCpan-4.1 and NetMHCIIpan-4.0: improved predictions of MHC antigen presentation by concurrent motif deconvolution and integration of MS MHC eluted ligand data. *Nucleic Acids Res.* 2020 Jul 2;48(W1):W449-W454. doi: 10.1093/nar/gkaa379. PMID: 32406916; PMCID: PMC7319546.
- Andreatta M, Trolle T, Yan Z, Greenbaum JA, Peters B, Nielsen M. An automated benchmarking platform for MHC class II binding prediction methods. *Bioinformatics.* 2018 May 1;34(9):1522-1528. doi: 10.1093/bioinformatics/btx820. PMID: 29281002; PMCID: PMC5925780.
- Jensen KK, Andreatta M, Marcatili P, Buus S, Greenbaum JA, Yan Z, Sette A, Peters B, Nielsen M. Improved methods for predicting peptide binding affinity to MHC class II molecules. *Immunology.* 2018 Jul;154(3):394-406. doi: 10.1111/imm.12889. Epub 2018 Feb 6. PMID: 29315598; PMCID: PMC6002223.
- Wang P, Sidney J, Kim Y, Sette A, Lund O, Nielsen M, Peters B. Peptide binding predictions for HLA DR, DP and DQ molecules. *BMC Bioinformatics.* 2010 Nov 22;11:568. PMID: 21092157; PMCID: PMC2998531.
- Wang, P., J. Sidney, C. Dow, B. Mothe, A. Sette, B. Peters. (2008). A Systematic Assessment of MHC Class II Peptide Binding Predictions and Evaluation of a Consensus Approach. *PLoS Computational Biology* 4(4). PMID: 18389056

For the B cell predictions, the references are:

- Kringelum JV, Nielsen M, Padkjær SB, Lund O. Structural analysis of B-cell epitopes in antibody:protein complexes. *Mol Immunol.* 2013 Jan;53(1-2):24-34. doi: 10.1016/j.molimm.2012.06.001. Epub 2012 Jul 10. PMID: 22784991; PMCID: PMC3461403
- Kringelum JV, Lundegaard C, Lund O, Nielsen M. Reliable B cell epitope predictions: impacts of method development and improved benchmarking. *PLoS Comput Biol.* 2012;8(12):e1002829. doi: 10.1371/journal.pcbi.1002829. Epub 2012 Dec 27. PMID: 23300419; PMCID: PMC3531324.
- Rubinstein ND, Mayrose I, Martz E, Pupko T. Epitopia: a web-server for predicting B-cell epitopes. *BMC Bioinformatics.* 2009 Sep 14;10:287. doi: 10.1186/1471-2105-10-287. PMID: 19751513; PMCID: PMC2751785.
- Liang S, Zheng D, Standley DM, Yao B, Zacharias M, Zhang C. EPSVR and EPMeta: prediction of antigenic epitopes using support vector regression and multiple server results. *BMC Bioinformatics.* 2010 Jul 16;11:381. doi: 10.1186/1471-2105-11-381. PMID: 20637083; PMCID: PMC2910724.
- Ponomarenko JV, Bourne PE., Antibody-protein interactions: benchmark datasets and prediction tools evaluation. *BMC Struct Biol.* 2007 Oct 2;7(1):64. PMID: 17910770

2.3.8 Other Ways to Access Tools

There are several ways that users can access many of the tools in the Analysis Resource in addition to using the tools directly on the Analysis Resource website. On the Analysis Resource home page (<http://tools.iedb.org>), there are tabs for Tools-API and Download. The former takes the user to information about the RESTful interface. The RESTful (Representational State Transfer) web service is available for the MHC class I and II binding predictions, MHC class I processing predictions, and MHC-NP for T cell epitopes, and linear epitope prediction methods for B cells. The service sends POST request to the tool's server, and relies on user supplied parameters. The Tools-API web page contains examples for accessing the service using the 'curl' program, although any program that sends POST requests to the server will work just as well (including a web browser). This feature allows users to perform predictions on the IEDB server in batch mode without having to install any software on their own systems. Additionally, users will always be assured that they are using the latest version of the tools.

The Download tab takes the user to the IEDB Tools Download web page. For users from non-commercial organizations with batch processing needs, the MHC class I and II binding prediction tools, linear B cell epitope prediction tool, ElliPro, the epitope cluster analysis tool, the MHC class I immunogenicity tool, and proteasomal cleavage prediction tool (added in 2016) are available as standalone scripts for download. These command line tools are kept in sync with the web tools and should therefore produce the same results as clicking through the web interface. They are freely available to non-profit/academic users through an open source license.

For commercial users that would like to run the analysis resource locally, licenses for a virtual machine image file or standalone downloads are available upon request for a modest annual fee. For the virtual machine, all tools are included except MHC-NP which was not developed by the IEDB team. The virtual machine image is kept in sync with the current version of the IEDB Analysis Resource and is updated approximately every six months. Commercial licenses are also available. Requests for either license can be sent to license@iedb.org.

2.4 Help Overview

The Help pull-down menu contains items that aid the user to understand and utilize the features of the website. Users can access relevant documentation, request help, and provide feedback.

2.4.1 Support

The IEDB Solutions Center is the primary resource for information on using the website's features. The user can submit help requests, check on the status of requests, browse and search the knowledge base and forums, and link to help documentation, such as the Curation Manual. In order to submit a help request via the Solutions Center, users must provide an email address, a subject line, and a description of the issue password. The Solutions Center can also be accessed by a link at the bottom of each web page.

The Solutions Center has extensive documentation organized in forums that address a variety of topics, including the epitope analysis and prediction tools, submitting data to the IEDB, querying the database, and general announcements and updates. The Solutions Center is also the repository for the Annual Compendia, the list of papers published by the IEDB team, release notes for the main website and the Analysis Resource, and the News and Noteworthy forum. In addition, written and video tutorials, including the presentations from the annual IEDB User Workshops can be found there.

2.4.2 Help Request

Users can submit help requests in two ways. At the bottom of most pages is a "Help request" link. This will initiate a submission to the IEDB Solutions Center. The second method also utilizes the IEDB Solutions Center. The top menu bar on the IEDB Solutions Center web page has a "Help Request" link. Selecting this link brings the user to a "Submit a request" web page. The user must fill in their email address, the subject line, and a description of the help request. All help requests are forwarded to the same help desk.

Help requests should consist of problems that users have with the application, such as a certain function of the system not working. Questions on how to use features of the application should be covered in the online help available in the Solutions Center. After submitting a help request, a confirmation e-mail will be sent to the user's e-mail address, which will include the help request number. When the help request issue has been resolved, an e-mail will be sent to notify the user that the help request has been satisfied.

Help requests are generally responded to within one business day and solved within five business days. Requests that are actually bug reports by users are answered initially to inform the requestor that their issue is being forwarded to the appropriate developers and they will be notified when a fix is released. Requests involving the curation or re-curation of articles will typically take longer to resolve.

2.4.3 Provide Feedback

All users are able to submit feedback, which will transmit an email message to the IEDB team. Feedback is intended for questions, input, and suggestions, such as new features they would like to see added in the future. Feedback helps the IEDB team update the system to provide users with the best possible experience. If users need help using the system or handling an unexpected result, a Help Request is more appropriate. The feedback feature can be accessed with the Support pull-down menu and the "Provide Feedback" at the bottom of each web page. Both initiate a submission to the IEDB Solutions Center.

2.4.4 Video Tutorials

There is a link to the IEDB video tutorials and information articles under the Help menu at the top of each web page. This link takes the user to the "How-to Videos" article in the Solutions Center in the Tutorials and Reference Materials folder. Most of the videos are edited versions of live presentations made at the annual IEDB User Workshops. The videos from the 2020-2022 events are currently posted for user viewing.

An overview of the resource and information regarding the data structure and curation are presented first. The next category addresses the query and reporting features. Videos for all the tools in the Analysis Resource are listed next. Videos for the legacy website have been removed because the legacy site has been retired and can no longer be accessed by users.

2.5 More IEDB

The "More IEDB" pull-down menu contains links to other relevant information that may be of interest to users. Its component links are described below.

2.5.1 Database Export

The contents of the Immune Epitope Database are exported weekly to files in XML format. The database export page (Figure 2.35) contains a complete database export in XML format along with exports of the various records that are referenced in the complete database export. The complete dataset export is provided as a ZIP archive (iedb_export.zip) containing one XML file for each reference contained in the database. The export also contains ZIP archives of XML files that provide users with the full record for the various identifiers that are referenced in the full database export, such as the IEDB source organism accession identifier list, the MHC allele name list, and the organism list. The supporting XML files only contain full records when the record is not otherwise available from the original source.

The database export page also provides the relevant XML Schema Definition (XSD) files for each of the XML files provided. The XSD file(s) for a particular XML file are located in the IEDB Schema box in the export table. In the case of the complete database export, multiple schemas are provided, with the primary schema being listed first, followed by any supporting schemas.

In addition to the XML export, the contents of the IEDB are exported weekly as a MySQL Structure Query Language (SQL) database. The database export page contains the complete database export in two formats, a single SQL script (iedb_public.sql.gz), and a TAR archive of MyISAM binary tables (iedb_public.tar.gz). Both export formats are generated from a 5.5.52-MariaDB database server. The database export page also provides an entity-relationship diagram (ERD) for the MySQL database.

Archives of the data as they appeared in IEDB 1.0, 2.0, 2.1, and 2.3 are available by clicking on the *Archived Versions*. For each version, there is a compressed file that contains an XML for each reference. The corresponding XSD files are also available for download.

Select compilations of data are available for download from the Database Export web page in CSV format in the section titled CSV Metric Exports. Users can download these files to conveniently obtain all the information in the IEDB for specific categories. Files are available for all epitopes, antigens, T cell assays, B cell assays, MHC ligand assays, and references. In addition, an export for antibody and T cell receptors was added in 2016. In addition to the files available on the Database Export web page, comma separated values (CSV) file exports are available on-demand from all list pages and list tabs throughout the site.

2.5.2 Meta-Analyses

The IEDB team has conducted a variety of analyses that examine data in the IEDB for several disease conditions and have been published in scientific journals. Over 20 papers have been published covering topics ranging from allergies, diabetes, and multiple sclerosis, to influenza, tuberculosis, malaria. When a user selects this menu item, they are directed to the Meta-analyses folder in the Knowledgebase and Forums section of the IEDB Solutions Center. The forum is updated as new meta-analyses are published. Users who subscribe to the folder will be automatically informed when content is added.

2.5.3 Citing the IEDB

Data and tools within the IEDB are presented as a public resource. Users are requested to consider citing the IEDB when they present information obtained from the IEDB or use tools contained in the Analysis Resource. It is expected that the authors of an entry as well as the IEDB are properly cited whenever their work is referred to:

1. The IEDB website should be cited using the URL: <http://www.iedb.org>
2. The journal reference for the IEDB was updated in 2018; and should be cited as:

Vita R, Mahajan S, Overton JA, Dhanda SK, Martini S, Cantrell JR, Wheeler DK, Sette A, Peters B. The Immune Epitope Database (IEDB): 2018 update. *Nucleic Acids Res.* 2019 Jan 8;47(D1):D339-D343. doi: 10.1093/nar/gky1006. PMID: 30357391; PMCID: PMC6324067.

2.5.4 Release Notes

Release notes are provided whenever a new release of the main website or the Analysis Resource is deployed to the public. When a user selects this menu item, they are directed to the Release Notes folder in the Knowledgebase and Forums section of the IEDB Solutions Center. The forum is updated as new releases are deployed. Users who subscribe to the folder will be automatically informed when content is added.

2.5.5 Links to External Sources

The IEDB system provides a list of links to external resources solely for the convenience of Immune Epitope Database visitors. The Immune Epitope Database has no interest in, responsibility for, or control over the linked-site. The Immune Epitope Database makes no promises or warranties of any kind, express or implied, including those of fitness for any particular purpose, as to the content of the linked-site. To view the links available, select the Links menu item. The hyperlinks on the links page are grouped by category:

- Public Databases, Prediction Algorithms, and Other Tools
- Antibody Related Links
- Bioinformatics Resource Centers
- MHC and TCR Related Links
- Protein Related Links
- Laboratory Resources
- Biodefense Resources

2.6 Learn More

The Learn More web page can be accessed from the link at the bottom of the Welcome box in the upper left-hand corner of the IEDB home page. The Learn More page has five major sections, as seen in Figure 2.36. The upper left section has a carousel of content from the New and Noteworthy forum in the Solutions Center. A graphic is usually included in the forum article that is displayed in the left-hand portion of the section. Below this section are three other sections titled Support, About the Data, and About Us. The Support section has links to submit a help request or feedback. The right-hand panel of the page lists the release notes for the most recent versions of the main website and the Analysis Resource.

The screenshot shows the IEDB website's 'Learn More' page. At the top, there is a navigation bar with 'Home', 'Specialized Searches', and 'Analysis Resource'. Below this, the page is organized into several sections. On the left, there is a 'News and Updates' section with a carousel of images and text. The first image shows a workshop, and the text states: 'The 2020 IEDB User Workshop will be held 5-6 November 2020 virtually via Zoom. Information is available at workshop.iedb.org.' Below this are three columns: 'Support' with links like 'Ask a question or submit an idea', 'About the Data' with charts and 'Export all or part of the IEDB' options, and 'About Us' with a group photo and 'Citation guidelines'. On the right side, there is a 'Latest Release Notes' section listing updates for IEDB Analysis Resource v2.23 (2 Jul 2020), v2.22 (4 Sep 2019), and v2.21 (26 Mar 2019), as well as IEDB v3.10.0 release notes. A 'View Previous Release Notes >>' link is at the bottom right.

Figure 2.12 The Learn More page, accessed from the Welcome message in the upper left corner of the IEDB home page

2.6.1 Support

Each link is briefly described below.

Ask a question or submit an idea

This link allows the user to submit a help request or suggestion via the IEDB Solutions Center. The user must specify their email address, the subject of their request, and description of the problem or suggestion. Users can also attach relevant files.

Browse support topics

The IEDB Solutions Center home page has a variety of user help, including several resources for getting started with the IEDB, recent Release Notes, and timely articles.

Get help with common searches

There is a category in the Solutions Center that contains a list of common queries and questions. These queries have been designed to parallel the IEDB meta-analysis efforts. Meta-analyses are performed at intervals to query the database for all data relating to subjects of interest (ex. Influenza, Malaria, Tuberculosis). These queries represent a simplified version of such analysis.

Browse links to epitope resources

The IEDB system provides a list of links to external resources solely for the convenience of Immune Epitope Database visitors as described in Section 2.5.5.

2.6.2 About the Data

Each link is briefly described below.

Export all or part of the IEDB

Clicking on this link will bring the user to the Database Export page that is described in Section 2.5.1.

Meta-analyses

The meta-analyses conducted by the IEDB team are described in Section 2.5.2.

Read about the data fields

This link gives the user access to a subset of the Curation Manual hosted in a Wiki that describes and defines the various data fields. Using the Find function of their web browser, users can search for the data field name of interest in order to gain a fuller understanding of the field definition and the possible values.

See how the data is entered

Users can access the full IEDB Curation Manual. The rules for curating all data fields are described in this Wiki.

Learn about the IEDB ontology

Selecting IEDB Ontology takes the user to <http://ontology.iedb.org>, the home page for the Ontology of Immune Epitopes (ONTIE). The IEDB ontology is available to download in a format that can be viewed with the Protégé OWL editor. The file ONTIE.owl contains the classes and relations used in the IEDB export. IEDB-export.owl is the ONTIE-annotated export of the IEDB T cell epitope data. Smaller versions of the file are available for download. The ONTIE home page also lists related ontologies that are used in constructing ONTIE.

2.6.3 About Us

Citation Guidelines

This link brings the user to the “Citing the IEDB” web page described in Section 2.5.3.

IEDB Publications

A list of publications relevant to the IEDB can be found on the Publications page. They have been grouped into four categories – General IEDB, Epitope Meta-analyses, Analysis Resource, and Curation. This list is updated with each new release of the main website. A list of IEDB related publications can also be found in the Publications folder in the IEDB Solutions Center.

Acknowledgements

A host of talented individuals have worked hard to make the Immune Epitope Database a reality. A roster of the current team members can be viewed on this page.

Terms of Use

The Terms of Use page is a collection of statements that outline the conditions related to using the IEDB system. The Terms of use includes our privacy notice, copyright information, and various disclaimers. The IEDB privacy policy describes what user information is collected, the circumstances for collecting it, and how it is used. Personal information about users is not collected unless the users choose to provide it.

Annual Compendia

An archive of the IEDB Annual Compendia can be found at this [link](#) to the IEDB Solutions Center. The list includes all Annual Compendia going back to 2005.

3 Scientific Publications

This section lists the scientific publications for which the IEDB played a contributory role. The first section lists publications authored by the IEDB contractor team over the past 20 years. The second section lists references that cited the IEDB. This list was compiled by using Google Scholar and the ISI Web of Knowledge to find citations for each one of the IEDB team authored papers.

3.1 IEDB Sponsored Publications

Scientific articles written by the IEDB team members that are relevant to the IEDB are listed below (organized by year of publication) through 2022. This publication list was re-evaluated in December 2022 to ensure that the list was accurate and up to date. As a result of this review, the publication list now contains 187 IEDB sponsored publications (including book chapters) between program inception in 2003 and 31 December 2022. We have added the number of citations for that publication in 2022, as well as the cumulative citations from 2003 to 2022.

Publication Year	Publication Type	IEDB Publication	2022 Citations	Cumulative Citations (2003-2022)
2022	Tools	Kaabinejadian S, Barra C, Alvarez B, Yari H, Hildebrand WH, Nielsen M. Accurate MHC Motif Deconvolution of Immunopeptidomics Data Reveals a Significant Contribution of DRB3, 4 and 5 to the Total DR Immunopeptidome. <i>Front Immunol.</i> 2022 Jan 26;13:835454. doi: 10.3389/fimmu.2022.835454. PMID: 35154160; PMCID: PMC8826445.	8	8
2022	Tools	Vita R, Mody A, Overton JA, Buus S, Haley ST, Sette A, Mallajosyula V, Davis MM, Long DL, Willis RA, Peters B, Altman JD. Minimal Information about MHC Multimers (MIAMM). <i>J Immunol.</i> 2022 Feb 1;208(3):531-537. doi: 10.4049/jimmunol.2100961. PMID: 35042788; PMCID: PMC8830768.	0	0
2022	General	Valencia A, Vergara C, Thio CL, Vince N, Douillard V, Grifoni A, Cox AL, Johnson EO, Kral AH, Goedert JJ, Mangia A, Piazzolla V, Mehta SH, Kirk GD, Kim AY, Lauer GM, Chung RT, Price JC, Khakoo SI, Alric L, Cramp ME, Donfield SM, Edlin BR, Busch MP, Alexander G, Rosen HR, Murphy EL, Wojcik GL, Carrington M, Gourraud PA, Sette A, Thomas DL, Duggal P. Trans-ancestral fine-mapping of MHC reveals key amino acids associated with spontaneous clearance of hepatitis C in HLA-DQB1. <i>Am J Hum Genet.</i> 2022 Feb 3;109(2):299-310. doi: 10.1016/j.ajhg.2022.01.001. Epub 2022 Jan 31. PMID: 35090584; PMCID: PMC8874224.	0	0
2022	General	Marrama D, Mahita J, Sette A, Peters B. Lack of evidence of significant homology of SARS-CoV-2 spike sequences to myocarditis-associated antigens. <i>EBioMedicine.</i> 2022 Jan;75:103807. doi: 10.1016/j.ebiom.2021.103807. Epub 2022 Jan 6. PMID: 34998242; PMCID: PMC8733122.	8	8
2022	Tools	Koşaloğlu-Yalçın Z, Lee J, Greenbaum J, Schoenberger SP, Miller A, Kim YJ, Sette A, Nielsen M, Peters B. Combined assessment of MHC binding and antigen abundance improves T cell epitope predictions. <i>iScience.</i> 2022 Feb 18;25(2):103850. doi: 10.1016/j.isci.2022.103850. Epub 2022 Feb 1. PMID: 35128348; PMCID: PMC8806398.	1	1

2022	Tools	Zierrep PF, Vita R, Blazeska N, Moumbock AFA, Greenbaum JA, Peters B, Günther S. Towards the prediction of non-peptidic epitopes. <i>PLoS Comput Biol.</i> 2022 Feb 18;18(2):e1009151. doi: 10.1371/journal.pcbi.1009151. PMID: 35180214; PMCID: PMC8893639.	0	0
2022	Tools	Trevizani R, Yan Z, Greenbaum JA, Sette A, Nielsen M, Peters B. A comprehensive analysis of the IEDB MHC class-I automated benchmark. <i>Brief Bioinform.</i> 2022 Jul 18;23(4):bbac259. doi: 10.1093/bib/bbac259. PMID: 35794711; PMCID: PMC9618166.	0	0
2022	Tools	Gutman I, Gutman R, Sidney J, Chihab L, Mishto M, Liepe J, Chiem A, Greenbaum J, Yan Z, Sette A, Koşaloğlu-Yalçın Z, Peters B. Predicting the Success of Fmoc-Based Peptide Synthesis. <i>ACS Omega.</i> 2022 Jun 27;7(27):23771-23781. doi: 10.1021/acsomega.2c02425. PMID: 35847273; PMCID: PMC9280948.	1	1
2022	Tools	Garcia Alvarez HM, Koşaloğlu-Yalçın Z, Peters B, Nielsen M. The role of antigen expression in shaping the repertoire of HLA presented ligands. <i>iScience.</i> 2022 Aug 17;25(9):104975. doi: 10.1016/j.isci.2022.104975. PMID: 36060059; PMCID: PMC9437844.	0	0
2022	General	Koşaloğlu-Yalçın Z, Blazeska N, Vita R, Carter H, Nielsen M, Schoenberger S, Sette A, Peters B. The Cancer Epitope Database and Analysis Resource (CEDAR). <i>Nucleic Acids Res.</i> 2023 Jan 6;51(D1):D845-D852. doi: 10.1093/nar/gkac902. PMID: 36250634; PMCID: PMC9825495.	0	0
2021	General	Edwards L, Jackson R, Overton J, Vita R, Blazeska N, Peters B, Sette A. An immunologically friendly classification of non-peptidic ligands. <i>Database (Oxford).</i> 2021 Mar 27;baab014. doi: 10.1093/database/baab014. PubMed PMID: 33772585	0	0
2021	General	Vita R, Zheng J, Jackson R, Dooley D, Overton J, Miller M, Berrios D, Scheuermann R, He Y, McGinty HK, Brochhausen M, Lin A, Jain S, Chibucos M, Judkins J, Giglio M, Feng I, Burns G, Brush M, Peters B, Stoeckert C. Standardization of assay representation in the Ontology for Biomedical Investigations. <i>Database (Oxford).</i> 2021 Jul 9;baab040. doi: 10.1093/database/baab040. PubMed PMID: 34244718	3	3
2021	Tools	Chronister W, Crinklaw A, Mahajan S, Vita R, Kosaloglu-Yalcin Z, Yan Z, Greenbaum J, Jessen L, Nielsen M, Christley S, Cowell L, Sette A, Peters B. TCRMatch: Predicting T-Cell Receptor Specificity Based on Sequence Similarity to Previously Characterized Receptors. <i>Front Immunol.</i> 2021 Mar 11;12:640725. doi: 10.3389/fimmu.2021.640725. PubMed PMID: 33777034	16	25
2021	General	Tarke A, Sidney J, Kidd C, Dan J, Ramirez S, Yu E, Mateus J, da Silva Antunes R, Moore E, Rubiro P, Methot N, Philips E, Mallal S, Frazier A, Rawlings S, Greenbaum J, Peters B, Smith D, Crotty S, Weiskopf D, Grifoni A, Sette A. Comprehensive analysis of T cell immunodominance and immunoprevalence of SARS-CoV-2 epitopes in COVID-19 cases. <i>Cell Rep Med.</i> 2021 Feb 16;2(2):100204. doi: 10.1016/j.xcrm.2021.100204. PubMed PMID: 33521695	136	266

2021	General	Grifoni A, Sidney J, Vita R, Peters B, Crotty S, Weiskopf D, Sette A. SARS-CoV-2 human T cell epitopes: Adaptive immune response against COVID-19. <i>Cell Host Microbe</i> . 2021 Jul 14;29(7):1076-1092. doi: 10.1016/j.chom.2021.05.010. PubMed PMID: 34237248	106	135
2021	Tools	Reardon B, Kosaloglu-Yalcin Z, Paul S, Peters B, Sette A. Allele-Specific Thresholds of Eluted Ligands for T-Cell Epitope Prediction. <i>Mol Cell Proteomics</i> . 2021;20:100122. doi: 10.1016/j.mcpro.2021.100122. PubMed PMID: 34303001	2	2
2021	Tools	Nilsson J, Grifoni A, Tarke A, Sette A, Nielsen M. PopCover-2.0. Improved Selection of Peptide Sets With Optimal HLA and Pathogen Diversity Coverage <i>Front Immunol</i> . 2021 Aug 17;12:728936. doi: 10.3389/fimmu.2021.728936. PubMed PMID: 34484239	2	4
2021	General	Eberhardt C, Kissick H, Patel M, Cardenas M, Prokhnevskaya N, Obeng R, Nasti T, Griffith C, Im S, Wang X, Shin D, Carrington M, Chen Z, Sidney J, Sette A, Saba N, Wieland A, Ahmed R. Functional HPV-specific PD-1 + stem-like CD8 T cells in head and neck cancer. <i>Nature</i> . 2021 Sep;597(7875):279-284. doi: 10.1038/s41586-021-03862-z. PubMed PMID: 34471285	51	63
2021	Tools	Refsgaard C, Barra C, Peng Z, Ternette N, Nielsen M. NetMHCphosPan - Pan-specific prediction of MHC class I antigen presentation of phosphorylated ligands. <i>Immunoinformatics</i> . 20 April 2021. doi: 10.1016/j.immuno.2021.100005	3	3
2021	Tools	Montemurro A, Schuster V, Povlsen H, Bentzen A, Jurtz V, Chronister W, Crinkloaw A, Hadrup S, Winther O, Peters B, Jessen L, Nielsen M. NetTCR-2.0 enables accurate prediction of TCR-peptide binding by using paired TCR α and β sequence data. <i>Commun Biol</i> . 2021 Sep 10;4(1):1060. doi: 10.1038/s42003-021-02610-3. PubMed PMID: 34508155	29	29
2020	Tools	Paul S, Croft N, Purcell A, Tschärke D, Sette A, Nielsen M, Peters B. Benchmarking predictions of MHC class I restricted T cell epitopes in a comprehensively studied model system. <i>PLoS Comput Biol</i> . 2020 May 26;16(5):e1007757. doi: 10.1371/journal.pcbi.1007757. PubMed PMID: 32453790	18	37
2020	General	Dimou A, Grewe P, Sidney J, Sette A, Norman P, Doebele R. HLA Class I Binding of Mutant EGFR Peptides in NSCLC Is Associated With Improved Survival. <i>J Thorac Oncol</i> . 2021 Jan;16(1):104-112. doi: 10.1016/j.jtho.2020.08.023. PubMed PMID: 32927123	0	2
2020	Tools	Peters B, Nielsen M, Sette A. T Cell Epitope Predictions. <i>Annu Rev Immunol</i> . 2020 Apr 26;38:123-145. doi: 10.1146/annurev-immunol-082119-124838. PubMed PMID: 32045313	35	79

2020	Tools	Reynisson B, Alvarez B, Paul S, Peters B, Nielsen M. NetMHCpan-4.1 and NetMHCIIpan-4.0: improved predictions of MHC antigen presentation by concurrent motif deconvolution and integration of MS MHC eluted ligand data. <i>Nucleic Acids Res.</i> 2020 Jul 2;48(W1):W449-W454. doi: 10.1093/nar/gkaa379. PubMed PMID: 32406916	240	486
2020	Tools	Reynisson B, Barra C, Kaabinejadian S, Hildebrand WH, Peters B, Nielsen M. Improved Prediction of MHC II Antigen Presentation through Integration and Motif Deconvolution of Mass Spectrometry MHC Eluted Ligand Data. <i>J Proteome Res.</i> 2020 Jun 5;19(6):2304-2315. doi: 10.1021/acs.jproteome.9b00874. PubMed PMID: 32308001	76	161
2020	General	Grifoni A, Sidney J, Zhang Y, Scheuerman RH, Peters B, Sette A. A Sequence Homology and Bioinformatic Approach Can Predict Candidate Targets for Immune Responses to SARS-CoV-2. <i>Cell Host Microbe.</i> 2020 Apr 8;27(4):671-680.e2. doi: 10.1016/j.chom.2020.03.002. PubMed PMID: 32183941	162	759
2020	General	Salimi N, Edwards L, Foos G, Greenbaum JA, Martini S, Reardon B, Shackelford D, Vita R, Zalman L, Peters B, Sette A. A behind-the-scenes tour of the IEDB curation process: an optimized process empirically integrating automation and human curation efforts. <i>Immunology.</i> 2020 Oct;161(2):139-147. doi: 10.1111/imm.13234. PubMed PMID: 32615639.	2	2
2020	Tools	Kosaloglu-Yacin Z, Sidney J, Chronister W, Peters B, Sette A. Comparison of HLA ligand elution data and binding predictions reveals varying prediction performance for the multiple motifs recognized by HLA-DQ2.5. <i>Immunology.</i> 2021 Feb;162(2):235-247. doi: 10.1111/imm.13279. PubMed PMID: 33064841	2	2
2020	General	Sidne J, Peters B, Sette A. Epitope prediction and identification- adaptive T cell responses in humans. <i>Semin Immunol.</i> 2020 Aug;50:101418. doi: 10.1016/j.smim.2020.101418. PubMed PMID: 33131981	8	16
2020	Tools	Vita R, Overton JA, Dunn P, Cheung K, Kleinstein SH, Sette A, Peters B. A structured model for immune exposures. <i>Database (Oxford).</i> 2020 Jan 1;2020:baaa016. doi: 10.1093/database/baaa016. PubMed PMID: 32283555	0	2
2020	Tools	Sachs A, Moore E, Kosaloglu-Yalcin Z, Peters B, Sidney J, Rosenberg SA, Robbins PF, Sette A. Impact of Cysteine Residues on MHC Binding Predictions and Recognition by Tumor-Reactive T Cells. <i>J Immunol.</i> 2020 Jul 15;205(2):539-549. doi: 10.4049/jimmunol.1901173. PubMed PMID: 32571843	2	6
2020	Tools	Nielsen M, Andreatta M, Peters B, Buus S. Immunoinformatics: Predicting Peptide–MHC Binding. <i>Annual Review of Biomedical Science.</i> 2020 July 3. doi: 10.1146/annurev-biodatasci-021920-100259.	11	21
2020	Tools	Barra C, Ackaert C, Reynisson B, Schockaert J, Jessen LE, Watson M, Jang A, Comtois-Marotte S, Goulet J, Pattijn S, Paramithiotis E, Nielsen M. Immunopeptidomic Data Integration to Artificial Neural Networks Enhances Protein-Drug Immunogenicity Prediction. <i>Front Immunol.</i> 2020 Jun 23;11:1304. doi: 10.3389/fimmu.2020.01304. PubMed PMID: 32655572	2	9

2019	General	Viscaino JA, Kubinok P, Kovalchik K, Ma Q, Duquette J, Mongrain I, Deutsch E, Peters B, Sette A, Sirois I, Caron E. The Human Immunopeptidome Project: A Roadmap to Predict and Treat Immune Diseases. <i>Mol Cell Proteomics</i> . 2020; 19(1): 31-49. doi: 10.1074/mcp.R119.001743. PubMed PMID: 31744855	12	61
2019	Tools	Paul S, Grifoni A, Peters B, Sette A. Major Histocompatibility Complex Binding, Eluted Ligands, and Immunogenicity: Benchmark Testing and Predictions. <i>Front Immunol</i> . 2020 Feb 5;10:3151. doi: 10.3389/fimmu.2019.03151. eCollection 2019. PubMed PMID: 32117208.	8	17
2019	Tools	Dhanda SK, Mahajan S, Paul S, Yan Z, Kim H, Jespersen MC, Jurtz V, Andreatta M, Greenbaum JA, Marcatili P, Sette A, Nielsen M, Peters B. Immune Epitope Database - Analysis Resource (IEDB-AR) in 2019. <i>Nucleic Acids Res</i> . 2019 Jul 2;47(W1):W502-W506. doi: 10.1093/nar/gkz452. PubMed PMID: 31114900	63	169
2019	General	Heide J, Vaughan KC, Sette A, Jacobs T, Schulze Zur Wiesch J. Comprehensive Review of Human Plasmodium falciparum-Specific CD8+ T Cell Epitopes. <i>Front Immunol</i> . 2019 Mar 21;10:397. doi: 10.3389/fimmu.2019.00397. eCollection 2019. PubMed PMID: 30949162	7	20
2019	General	Grifoni A, Mahajan S, Sidney J, Martini S, Scheuerman RH, Peters B, Sette A. A survey of known immune epitopes in the enteroviruses associated with acute flaccid myelitis. <i>Hum Immunol</i> . 2019 Nov;80(11):923-929. doi: 10.1016/j.humimm.2019.08.004. Epub 2019 Aug 23. PubMed PMID: 31451291	2	3
2019	Tools	Mahajan S, Yan Z, Jespersen MC, Jensen KK, Marcatili P, Nielsen M, Sette A, Peters B. Benchmark datasets of immune receptor-epitope structural complexes. <i>BMC Bioinformatics</i> . 2019 Oct 10;20(1):490. doi: 10.1186/s12859-019-3109-6. PubMed PMID: 31601176	2	7
2018	Tools/ Analysis Resource	Andreatta M, Nielsen M. Bioinformatics Tools for the Prediction of T-Cell Epitopes. <i>Methods Mol Biol</i> . 2018; 1785:269-281. doi: 10.1007/978-1-4939-7841-0_18. PubMed PMID: 29714025	3	19
2018	General	Vaughan K, Xu X, Peters B, Sette A. Investigation of outbreak-specific nonsynonymous mutations on Ebolavirus GP in the context of known immune reactivity. <i>J Immunol Res</i> . 2018 Nov 15;2018:1846207. doi: 10.1155/2018/1846207. eCollection 2018. PubMed PMID: 30581874	0	1
2018	General	Vita R, Overton JA, Mungall CJ, Sette A, Peters B. FAIR principles and the IEDB: Short-term improvements and a Long-term vision of OBO-Foundry mediated machine-actionable interoperability. <i>Database (Oxford)</i> . 2018 Jan 1;2018. doi: 10.1093/database/bax105. PubMed PMID: 29688354	1	11
2018	General	Vita R, Overton JA, Peters B. Identification of Errors in the IEDB Using Ontologies. <i>Database (Oxford)</i> . 2018 Jan 1;2018. doi: 10.1093/database/bay005. PubMed PMID: 29688357	0	0

2018	General	Mahajan S, Vita R, Shackelford D, Lane J, Schulten V, Zarebski L, Jespersen MC, Marcatili P, Nielsen M, Sette A, Peters B. Epitope Specific Antibodies and T Cell Receptors in the Immune Epitope Database. <i>Front Immunol.</i> 2018 Nov 20;9:2688. doi: 10.3389/fimmu.2018.02688. eCollection 2018. PubMed PMID: 30515166	7	24
2018	General	Vita R, Mahajan S, Overton JA, Dhanda SK, Martini S, Cantrell JR, Wheeler DK, Sette A, Peters B. The Immune Epitope Database (IEDB): 2018 update. <i>Nucleic Acids Res.</i> 2019 Jan 8;47(D1):D339-D343. doi: 10.1093/nar/gky1006. PubMed PMID: 30357391	285	1054
2018	General	Tian Y, da Silva Antunes R, Sidney J, Lindestam Arlehamn CS, Grifoni A, Dhanda SK, Paul S, Peters B, Weiskopf D, Sette A. A Review on T Cell Epitopes Identified Using Prediction and Cell-Mediated Immune Models for Mycobacterium tuberculosis and Bordetella pertussis. <i>Front Immunol.</i> 2018 Nov 29;9:2778. doi: 10.3389/fimmu.2018.02778. eCollection 2018. PubMed PMID: 30555469	5	32
2018	Tools/ Analysis Resource	Dhanda SK, Vaughan K, Schulten V, Grifoni A, Weiskopf D, Sidney J, Peters B, Sette A. Development of a novel clustering tool for linear peptide sequences. <i>Immunology.</i> 2018 Nov;155(3):331-345. doi: 10.1111/imm.12984. Epub 2018 Aug 6. PubMed PMID: 30014462	14	55
2018	Tools/ Analysis Resource	Dhanda SK, Karosiene E, Edwards L, Grifoni A, Paul S, Andreatta M, Weiskopf D, Sidney J, Nielsen M, Peters B, Sette A. Predicting HLA CD4 immunogenicity in human populations. <i>Front Immunol.</i> 2018 Jun 14;9:1369. doi: 10.3389/fimmu.2018.01369. eCollection 2018. PubMed PMID: 29963059	16	69
2018	Tools/ Analysis Resource	Barra C, Alvarez B, Paul S, Sette A, Peters B, Andreatta M, Buus S, Nielsen M. Footprints of antigen processing boost MHC class II natural ligand binding predictions. <i>Genome Med.</i> 2018 Nov 16;10(1):84. doi: 10.1186/s13073-018-0594-6. PubMed PMID: 30446001.	11	55
2018	Tools/ Analysis Resource	Alvarez B, Barra C, Nielsen M, Andreatta M. Computational Tools for the Identification and Interpretation of Sequence Motifs in Immuno-peptidomes. <i>Proteomics.</i> 2018 Jun;18(12):e1700252. doi: 10.1002/pmic.201700252. Epub 2018 Feb 26. PubMed PMID: 29327813.	3	28
2018	Tools/ Analysis Resource	Dhanda SK, Vita R, Ha B, Grifoni A, Peters B, Sette A. ImmunomeBrowser: A tool to aggregate and visualize complex and heterogeneous epitopes in reference protein. <i>Bioinformatics.</i> 2018 Nov 15;34(22):3931-3933. doi: 10.1093/bioinformatics/bty463. PubMed PMID: 29878047	7	21
2018	Tools/ Analysis Resource	Jensen KK, Andreatta M, Marcatili P, Buus S, Greenbaum JA, Yan Z, Sette A, Peters B, Nielsen M. Improved methods for predicting peptide binding affinity to MHC class II molecules. <i>Immunology.</i> 2018 Jul;154(3):394-406. doi: 10.1111/imm.12889. Epub 2018 Feb 6. PubMed PMID: 29315598	110	533
2018	Tools/ Analysis Resource	Paul S, Karosiene E, Dhanda SK, Jurtz V, Edwards L, Nielsen M, Sette A, Peters B. Determination of a Predictive Cleavage Motif for Eluted Major Histocompatibility Complex Class II Ligands. <i>Front Immunol.</i> 2018 Aug	9	41

		6;9:1795. doi: 10.3389/fimmu.2018.01795. eCollection 2018. PubMed PMID: 30127785		
2018	Tools/ Analysis Resource	Carrasco Pro S, Lindestam Arlehamn CS, Dhanda SK, Carpenter C, Lindvall M, Faruqi AA, Santee CA, Renz H, Sidney J, Peters B, Sette A. Microbiota epitope similarity either dampens or enhances the immunogenicity of disease-associated antigenic epitopes. PLoS One. 2018 May 7;13(5):e0196551. doi: 10.1371/journal.pone.0196551. eCollection 2018. PubMed PMID: 29734356	6	26
2017	General	Vita R, Overton JA, Sette A, Peters B. Better living through ontologies at the Immune Epitope Database. Database (Oxford). 2017 Jan 1;2017(1). doi: 10.1093/database/bax014. PubMed PMID: 28365732	0	1
2017	General	Remesh SG, Andreatta M, Ying G, Kaefer T, Nielsen M, McMurtrey C, Hildebrand W, Peters B, Zajonc DM. Unconventional peptide Presentation by Major Histocompatibility Complex (MHC) Class I Allele HLA-A*02:01: Breaking Confinement. J Biol Chem. 2017 Mar 31; 292(13):5262-5270. doi:10.1074/jbc.M117.776542. Epub 2017 Feb 8. PubMed PMID: 28179428	6	46
2017	Tools/ Analysis Resource	Fleri W, Paul S, Dhanda SK, Mahajan S, Xu X, Peters B, Sette A. The Immune Epitope Database and Analysis Resource in Epitope Discovery and Synthetic Vaccine Design. Front Immunol. 2017 Mar 14;8:278. doi:10.3389/fimmu.2017.00278. eCollection 2017. Review. PubMed PMID: 28352270; PubMed Central PMCID: PMC5348633.	75	301
2017	Tools/ Analysis Resource	Nielsen M, Andreatta M. NNAlign: a platform to construct and evaluate artificial neural network models of receptor-ligand interactions. Nucleic Acids Res. 2017 April 12. doi:10.1093/nar/gkx276. [Epub ahead of print] PubMed PMID: 28407117.	12	39
2017	Tools/ Analysis Resource	Andreatta M, Alvarez B, Nielsen M. GibbsCluster: unsupervised clustering and alignment of peptide sequences. Nucleic Acids Res. 2017 Apr 12. doi:10.1093/nar/gkx248. [Epub ahead of print] PubMed PMID: 28407089.	27	125
2017	Tools/ Analysis Resource	Jespersen MC, Peters B, Nielsen M, Marcatili P. BepiPred-2.0: improving sequence-based B-cell epitope predictions using conformational epitopes. Nucleic Acids Res. 2017 May 2. doi:10.1093/nar/gkx346. [Epub ahead of print] PubMed PMID: 28472356.	267	976
2017	General	Sidney J, Becart S, Zhou M, Duffy K, Lindvall M, Moore EC, Moore EL, Rao T, Rao N, Nielsen M, Peters B, Sette A. Citrullination only infrequently impacts peptide binding to HLA class II MHC. PLoS One. 2017 May 8;12(5):e0177140. doi:10.1371/journal.pone.0177140. eCollection 2017. PubMed PMID: 28481943; PubMed Central PMCID: PMC5421785	5	38
2017	Tools/ Analysis Resource	Andreatta M, Jurtz VI, Kaefer T, Sette A, Peters B, Nielsen M. Machine learning reveals a non-canonical mode of peptide binding to MHC class II molecules. Immunology. 2017 May 24. doi:10.1111/imm.12763. [Epub ahead of print] PubMed PMID: 28542831.	2	17

2017	General	Fleri W, Vaughan K, Salimi N, Vita R, Peters B, Sette A. The Immune Epitope Database: How Data Are Entered and Retrieved. <i>J Immunol Res.</i> 2017; 2017:5974574. doi:10.1155/2017/5974574. Epub 2017 May 29. Review. PubMed PMID: 28634590; PubMed Central PMCID: PMC5467323.	1	17
2017	Tools/ Analysis Resource	Paul S, Arlehamm CSL, Schulten V, Westernberg L, Sidney J, Peters B, Sette A. Experimental validation of the RATE tool for inferring HLA restrictions of T cell epitopes. <i>BMC Immunol.</i> 2017 Jun 21; 18(Suppl 1):20. doi: 10.1186/s12865-017-0204-1. PubMed PMID: 28681704.	3	11
2017	General	Shao W, Pedrioli PGA, Wolski W, Scurtescu C, Schmid E, Vizcano JA, Courcelles M, Schuster H, Kowalewski D, Marino F, Arlehamm CSL, Vaughan K, Peters B, Sette A, Ottenhoff THM, Meijgaarden KE, Nieuwenhuizen N, Kaufmann SHE, Schlapbach R, Castle JC, Nesvizhskii AI, Nielsen M, Deutsch EW, Campbell DS, Moritz RL, Zubarev RA, Ytterberg AJ, Purcell AW, Marcilla M, Paradela A, Wang Q, Costello CE, Ternette N, van Rammensee HG, Thibault P, Perreault C, Bassani-Sterberg M, Aebersold R, Caron E. The SystemMHC Atlas project. <i>Nucleic Acids Res.</i> 2017 Jul 29. doi:10.1093/nar/gkx664. [Epub ahead of print] PubMed PMID: 28985418.	14	123
2017	General	Vaughan K, Xu X, Cron E, Peters B, Sette A. Deciphering the MHC-associated peptidome: a review of naturally processed ligand data. <i>Expert Rev Proteomics.</i> 2017 Sep; 14(9):729-736. doi:10.1080/14789450.2017.1361825. Epub 2017 Aug 11. PubMed PMID: 28756714.	0	22
2017	Tools/ Analysis Resource	Dhanda SK, Grifoni A, Pham J, Vaughan K, Sidney J, Peters B, Sette A. Development of a strategy and computational application to select candidate protein analogues with reduced HLA binding and immunogenicity. <i>Immunology.</i> 2017 Aug 22. doi:10.1111/imm.12816. [Epub ahead of print] PubMed PMID: 28833085.	3	23
2017	Tools/ Analysis Resource	Jurtz VI, Rosenberg Johansen A, Nielsen M, Almagro Armenteros JJ, Nielsen H, Kaae Sonderby C, Winther O, Kaae Sonderby S. An introduction to Deep learning on biological sequence data - Examples and solutions. <i>Bioinformatics.</i> 2017 Aug 23 doi:10.1093/bioinformatics/btx531. [Epub ahead of print] PubMed PMID: 28961695.	26	140
2017	Tools/ Analysis Resource	Jurtz V, Paul S, Andreaata M, Marcatili P, Peters B, Nielsen M. NetMHCpan-4.0: Improved Peptide-MHC Class I Interaction Predictions Integrating Eluted Ligand and Peptide Binding Affinity Data. <i>J Immunol.</i> 2017 Nov 1;199(9):3360-3368. doi:10.4049/jimmunol/1700893. Epub 2017 Oct 4. PubMed PMID: 28978689; PubMed Central PMCID: PMC5679736.	184	1011
2017	Tools/ Analysis Resource	Andreata M, Trolle T, Yan Z, Greenbaum JA, Peters B, Nielsen M. An automated benchmarking platform for MHC class II binding prediction methods. <i>Bioinformatics.</i> 2017 Dec 21. doi:10.1093/bioinformatics/btx820. [Epub ahead of print] PubMed PMID: 29281002.	10	65

2016	Tools/ Analysis Resource	Rasmussen M, Fenoy E, Harndahl M, Kristensen AB, Nielsen IK, Nielsen M, Buus S. Pan-Specific Prediction of Peptide-MHC Class I Complex Stability, a Correlate of T Cell Immunogenicity. <i>J Immunol.</i> 2016 Aug 15;197(4):1517-24. doi: 10.4049/jimmunol.1600582. Epub 2016 Jul 8. PubMed PMID: 27402703.	22	127
2016	General	Bandrowski A, Brinkman R, Brochhausen M, Brush MH, Bug B, Chibucos MC, Clancy K, Courtot M, Derom D, Dumontier M, Fan L, Fostel J, Fragoso G, Gibson F, Gonzalez-Beltran A, Haendel MA, He Y, Heiskanen M, Hernandez-Boussard T, Jensen M, Lin Y, Lister AL, Lord P, Malone J, Manduchi E, McGee M, Morrison N, Overton JA, Parkinson H, Peters B, Rocca-Serra P, Ruttenberg A, Sansone SA, Scheuermann RH, Schober D, Smith B, Soldatova LN, Stoeckert CJ Jr, Taylor CF, Torniai C, Turner JA, Vita R, Whetzel PL, Zheng J. The Ontology for Biomedical Investigations. <i>PLoS One.</i> 2016 Apr 29;11(4):e0154556. doi: 10.1371/journal.pone.0154556. eCollection 2016. PubMed PMID: 27128319	53	267
2016	Tools/ Analysis Resource	Bresciani A, Paul S, Schommer N, Dillon MB, Bancroft T, Greenbaum J, Sette A, Nielsen M, Peters B. T-cell recognition is shaped by epitope sequence conservation in the host proteome and microbiome. <i>Immunology.</i> 2016 May;148(1):34-9. doi: 10.1111/imm.12585. Epub 2016 Feb 8. PubMed PMID: 26789414	3	21
2016	General	Fleri W, Salimi N, Vita R, Peters B, Sette A. Immune Epitope Database and Analysis Resource, <i>Encyclopedia of Immunobiology</i> , Vol. 2, Elsevier Ltd., 20 May 2016, doi:10.1016/B978-0-12-374279-7.06004-5	0	0
2016	Tools/ Analysis Resource	McMurtrey C, Trolle T, Sansom T, Remesh SG, Kaever T, Bardet W, Jackson K, McLeod R, Sette A, Nielsen M, Zajonc DM, Blader IJ, Peters B, Hildebrand W. Toxoplasma gondii peptide ligands open the gate of the HLA class I binding groove. <i>Elife.</i> 2016 Jan 29;5. pii: e12556. doi: 10.7554/eLife.12556. PubMed PMID: 26824387	8	64
2016	Tools/ Analysis Resource	Nielsen M, Andreatta M, NetMHCpan-3.0; improved prediction of binding to MHC class I molecules integrating information from multiple receptor and peptide length data sets. <i>Genome Medicine</i> , March 2016, Mar 30;8(1):33. doi: 10.1186/s13073-016-0288-x. PubMed PMID: 27029192	50	443
2016	Tools/ Analysis Resource	Paul S, Sidney J, Sette A, Peters B. TepiTool: A Pipeline for Computational Prediction of T Cell Epitope Candidates. <i>Curr Protoc Immunol.</i> 2016 Aug 1;114:18.19.1-18.19.24. doi: 10.1002/cpim.12. PubMed PMID: 27479659	39	156
2016	General	Ponomarenko J, Vaughan K, Paul S, Peters B, Sette A, Haeussler M, Maurer-Stroh S. Ebola: an analysis of immunity at the molecular level, 2015 International Workshop on Artificial Immune Systems (AIS), Taormina, 2015, pp. 1-8. doi: 10.1109/AISW.2015.7469230, published online at IEEE Xplore Digital Library, URL: http://ieeexplore.ieee.org/stamp/stamp.jsp?tp=&arnumber=7469230&isnumber=7469223	0	5

2016	Tools/ Analysis Resource	Trolle T, McMurtrey CP, Sidney J, Bardet W, Osborn SC, Kaefer T, Sette A, Hildebrand WH, Nielsen M, Peters B. The Length Distribution of Class I-Restricted T Cell Epitopes Is Determined by Both Peptide Supply and MHC Allele-Specific Binding Preference. <i>J Immunol.</i> 2016 Feb 15;196(4):1480-7. doi: 10.4049/jimmunol.1501721. Epub 2016 Jan 18. PubMed PMID: 26783342	25	139
2016	General	Vita R, Overton JA, Seymour E, Sidney J, Kaufman J, Tallmadge RL, Ellis S, Hammond J, Butcher GW, Sette A, Peters B. An ontology for major histocompatibility restriction. <i>J Biomed Semantics.</i> 2016 Jan 11;7:1. doi:10.1186/s13326-016-0045-5. eCollection 2016. PubMed PMID: 26759709	0	4
2016	General	Xu X, Vaughan K, Weiskopf D, Grifoni A, Diamond MS, Sette A, Peters B. Identifying Candidate Targets of Immune Responses in Zika Virus Based on Homology to Epitopes in Other Flavivirus Species. <i>PLoS Curr.</i> 2016 Nov 15;8. pii: ecurrents.outbreaks.9aa2e1fb61b0f632f58a098773008c4b. doi: 10.1371/currents.outbreaks.9aa2e1fb61b0f632f58a098773008c4b. PubMed PMID: 28018746	9	79
2015	General	Cheloha RW, Sullivan JA, Wang T, Sand JM, Sidney J1, Sette A1, Cook ME, Suresh M, Gellman SH. Consequences of periodic α -to- β (3) residue replacement for immunological recognition of peptide epitopes. <i>ACS Chem Biol.</i> 2015 Mar 20;10(3):844-54. doi: 10.1021/cb500888q. Epub 2015 Jan 5. PubMed PMID: 25559929.	1	12
2015	General	Vita R, Vasilevsky N, Bandrowski A, Haendel M, Sette A, Peters B. Reproducibility and Conflicts in Immune Epitope Data. <i>Immunology.</i> 2015 Dec 17. doi: 10.1111/imm.12566. [Epub ahead of print] PubMed PMID: 26678806	0	4
2015	Tools/ Analysis Resource	Trolle T, Metushi IG, Greenbaum JA, Kim Y, Sidney J, Lund O, Sette A, Peters B, Nielsen M. "Automated benchmarking of peptide-MHC class I binding predictions". <i>Bioinformatics.</i> 2015 Feb 25. pii: btv123. [Epub ahead of print] PubMed PMID: 25717196.	7	118
2015	Tools/ Analysis Resource	Sela-Culang, I, I, Ofra, Y., and Peters, B. "Antibody specific epitope prediction - emergence of a new paradigm", <i>Current Opinion in Virology</i> , Volume 11, April 2015, Pages 98–102, PMID: 25837466.	12	57
2015	Tools/ Analysis Resource	Paul S, Lindestam Arlehamn CS, Scriba TJ, Dillon MB, Oseroff C, Hinz D, McKinney DM, Carrasco Pro S, Sidney J, Peters B, Sette A. Development and validation of a broad scheme for prediction of HLA class II restricted T cell epitopes. <i>J Immunol Methods.</i> 2015 Jul;422:28-34. doi: 10.1016/j.jim.2015.03.022. Epub 2015 Apr 7. PubMed PMID: 25862607.	24	132
2015	Tools/ Analysis Resource	Paul S, Dillon MB, Lindestam Arlehamn CS, Huang H, Davis MM, McKinney DM, Scriba TJ, Sidney J, Peters B, Sette A. A Population Response Analysis Approach To Assign Class II HLA-Epitope Restrictions. <i>J Immunol.</i> 2015 Jun 15;194(12):6164-76. doi: 10.4049/jimmunol.1403074. Epub 2015 May 6. PubMed PMID: 25948811; PubMed Central PMCID: PMC4458389.	3	28

2015	Tools/ Analysis Resource	Klausen MS, Anderson MV, Jespersen MC, Nielsen M, Marcatili P. LYRA, a webserver for lymphocyte receptor structural modeling. <i>Nucleic Acids Res.</i> 2015 Jul 1;43(W1):W349-W355. Epub 2015 May 24. PubMed PMID: 26007650.	13	54
2015	Tools/ Analysis Resource	Andreatta M, Karosiene E, Rasmussen M, Stryhn A, Buus S, Nielsen M. Accurate pan-specific prediction of peptide-MHC class II binding affinity with improved binding core identification. <i>Immunogenetics.</i> 2015 Sep 29. [Epub ahead of print] PubMed PMID: 26416257.	29	250
2015	Tools/ Analysis Resource	Carrasco Pro S, Sidney J, Paul S, Lindestam Arlehamn C, Weiskopf D, Peters B, Sette A. Automatic Generation of Validated Specific Epitope Sets. <i>J Immunol Res.</i> 2015;2015:763461. doi: 10.1155/2015/763461. Epub 2015 Oct 19. PubMed PMID: 26568965.	8	55
2015	Tools/ Analysis Resource	Andreatta M, Nielsen M. Gapped sequence alignment using artificial neural networks: application to the MHC class I system. <i>Bioinformatics.</i> 2015 Oct 29. pii: btv639. [Epub ahead of print] PubMed PMID: 26515819.	125	841
2015	General	Vaughan K, Ponomarenko J, Peters B, Sette A. Analysis of Human RSV Immunity at the Molecular Level: Learning from the Past and Present. <i>PLoS One.</i> 2015 May 22;10(5):e0127108. doi: 10.1371/journal.pone.0127108. eCollection 2015. PubMed PMID: 26001197 .	0	3
2015	General	Sette A, Paul S, Vaughan K, Peters B. The Use of the Immune Epitope Database to Study Autoimmune Epitope Data Related to Alopecia Areata. <i>J Investig Dermatol Symp Proc.</i> 2015 Nov;17(2):36-41. doi: 10.1038/jidsymp.2015.39. Review. PubMed PMID: 26551944.	1	7
2014	Tools/ Analysis Resource	Hansen AM, Rasmussen M, Svitek N, Harndahl M, Golde WT, Barlow J, Nene V, Buus S, Nielsen M. Characterization of binding specificities of bovine leucocyte class I molecules: impacts for rational epitope discovery. <i>Immunogenetics.</i> 2014 Dec;66(12):705-18. doi: 10.1007/s00251-014-0802-5. Epub 2014 Sep 4. PubMed PMID: 25186069.	2	11
2014	General	Ponomarenko J, Vaughan K, Sette A, Maurer-Stroh S. Conservancy of mAb Epitopes in Ebolavirus Glycoproteins of Previous and 2014 Outbreaks. <i>PLOS Currents Outbreaks.</i> 2014 Nov 3. Edition 1.	1	14
2014	General	Vaughan K, Peters B, Mallone R, von Herrath M, Roep BO, Sette A. Navigating diabetes-related immune epitope data: resources and tools provided by the Immune Epitope Database (IEDB). <i>Immunome Res.</i> 2013;9(1). doi: 10.4172/1745-7580.1000063. PubMed PMID: 25140192; PubMed Central PMCID: PMC4134942	0	6
2014	General	Vaughan K, Seymour E, Peters B, Sette A. Substantial gaps in knowledge of <i>Bordetella pertussis</i> antibody and T cell epitopes relevant for natural immunity and vaccine efficacy. <i>Hum Immunol.</i> 2014 May;75(5):440-451. doi: 10.1016/j.humimm.2014.02.013. Epub 2014 Feb 12. Review. PubMed PMID: 24530743	0	21
2014	Tools/ Analysis Resource	Trolle T, Nielsen M. NetTepi: an integrated method for the prediction of T cell epitopes. <i>Immunogenetics.</i> 2014 May 27. [Epub ahead of print] PubMed PMID: 24863339.	8	47

2014	Tools/ Analysis Resource	Kim Y, Sidney J, Buus S, Sette A, Nielsen M, Peters B. Dataset size and composition impact the reliability of performance benchmarks for peptide-MHC binding predictions. BMC Bioinformatics. 2014 Jul 14;15:241. doi: 10.1186/1471-2105-15-241. PubMed PMID: 25017736; PubMed Central PMCID: PMC4111843	1	100
2014	General	Vita R, Overton JA, Greenbaum JA, Ponomarenko J, Clark JD, Cantrell JR, Wheeler DK, Gabbard JL, Hix D, Sette A, Peters B. The immune epitope database (IEDB) 3.0. Nucleic Acids Res. 2014 Oct 9. pii: gku938. [Epub ahead of print] PubMed PMID: 25300482.	91	946
2013	General	Kim Y, Yewdell JW, Sette A, Peters B. Positional bias of MHC class I restricted T-cell epitopes in viral antigens is likely due to a bias in conservation. PLoS Comput Biol. 2013 Jan;9(1):e1002884. doi: 10.1371/journal.pcbi.1002884. Epub 2013 Jan 24. PubMed PMID: 23357871; PubMed Central PMCID: PMC3554532.	0	11
2013	General	Vita R, Overton JA, Greenbaum JA, Sette A; OBI consortium, Peters B. Query enhancement through the practical application of ontology: the IEDB and OBI. J Biomed Semantics. 2013 Apr 15;4 Suppl 1:S6. doi: 10.1186/2041-1480-4-S1-S6. Epub 2013 Apr 15. PubMed PMID: 23734660; PubMed Central PMCID: PMC3633001	0	6
2013	Tools/ Analysis Resource	Jessen LE, Hoof I, Lund O, Nielsen M. SigniSite: Identification of residue-level genotype-phenotype correlations in protein multiple sequence alignments. Nucleic Acids Res. 2013 Jul 1;41(Web Server issue):W286-91. doi: 10.1093/nar/gkt497. Epub 2013 Jun 12. PMID: 23761454; PMCID: PMC3692133	1	21
2013	Tools/ Analysis Resource	Jørgensen KW, Rasmussen M, Buus S, Nielsen M. NetMHCstab - predicting stability of peptide:MHC-I complexes; impacts for CTL epitope discovery. Immunology. 2013 Aug 8. doi: 10.1111/imm.12160. PubMed PMID: 23927693; PubMed Central PMCID: PMC3893846	7	119
2013	Tools/ Analysis Resource	Schubert B, Lund O, Nielsen M. Evaluation of peptide selection approaches for epitope-based vaccine design. Tissue Antigens. 2013 Oct;82(4):243-51. doi: 10.1111/tan.12199. PubMed PMID: 24461003.	1	25
2013	Tools/ Analysis Resource	Karosiene E, Rasmussen M, Blicher T, Lund O, Buus S, Nielsen M. NetMHCIIpan-3.0, a common pan-specific MHC class II prediction method including all three human MHC class II isotypes, HLA-DR, HLA-DP and HLA-DQ. Immunogenetics. 2013 Oct;65(10):711-24. doi: 10.1007/s00251-013-0720-y. Epub 2013 Jul 31. PubMed PMID: 23900783	23	241
2013	General	Vaughan K, Peters B, O'Connor KC, Martin R, Sette A. A molecular view of multiple sclerosis and experimental autoimmune encephalitis: What can we learn from the epitope data? J Neuroimmunol. 2013 Dec 12. pii: S0165-5728(13)00340-8. doi: 10.1016/j.jneuroim.2013.12.009. PubMed PMID: 24365494	0	12

2013	Tools/ Analysis Resource	Paul S, Kolla RV, Sidney J, Weiskopf D, Fleri W, Kim Y, Peters B, Sette A. Evaluating the immunogenicity of protein drugs by applying in vitro MHC binding data and the immune epitope database and analysis resource. Clin Dev Immunol. 2013;2013:467852. doi: 10.1155/2013/467852. Epub 2013 Oct 8. PubMed PMID: 24222776; PubMed Central PMCID: PMC3816028	3	47
2013	Tools/ Analysis Resource	Calis JJ, Maybeno M, Greenbaum JA, Weiskopf D, De Silva AD, Sette A, Kesmir C, Peters B. Properties of MHC class I presented peptides that enhance immunogenicity. PLoS Comput Biol. 2013 Oct;9(10):e1003266. doi: 10.1371/journal.pcbi.1003266. Epub 2013 Oct 24. PubMed PMID: 24204222; PubMed Central PMCID: PMC3808449	110	555
2013	Tools/ Analysis Resource	Paul S, Weiskopf D, Angelo MA, Sidney J, Peters B, Sette A. HLA Class I Alleles Are Associated with Peptide-Binding Repertoires of Different Size, Affinity, and Immunogenicity. J Immunol. 2013 Dec 15;191(12):5831-9. doi: 10.4049/jimmunol.1302101. Epub 2013 Nov 4. PubMed PMID: 24190657; PubMed Central PMCID: PMC3872965	31	220
2012	Tools/ Analysis Resource	Lundegaard C, Lund O, Nielsen M. Predictions versus high-throughput experiments in T-cell epitope discovery: competition or synergy? Expert Rev Vaccines. 2012 Jan;11(1):43-54. doi: 10.1586/erv.11.160. PubMed PMID: 22149708.	0	13
2012	Tools/ Analysis Resource	Karosiene E, Lundegaard C, Lund O, Nielsen M. NetMHCcons: a consensus method for the major histocompatibility complex class I predictions. Immunogenetics. 2012 Mar;64(3):177-86. doi: 10.1007/s00251-011-0579-8. Epub 2011 Oct 20. PubMed PMID: 22009319.	26	210
2012	Tools/ Analysis Resource	Kim Y, Ponomarenko J, Zhu Z, Tamang D, Wang P, Greenbaum J, Lundegaard C, Sette A, Lund O, Bourne PE, Nielsen M, Peters B. Immune epitope database analysis resource. Nucleic Acids Res. 2012 Jul;40 (Web Server issue):W525-30. Epub 2012 May 18. PubMed PMID: 22610854.	33	418
2012	General	Kim Y, Vaughan K, Greenbaum J, Peters B, Law M, Sette A. A Meta-Analysis of the Existing Knowledge of Immunoreactivity against Hepatitis C Virus (HCV). PLoS One. 2012;7(5):e38028. Epub 2012 May 31. PubMed PMID: 22675428; PubMed Central PMCID: PMC3364976.	0	17
2012	Tools/ Analysis Resource	Kringelum JV, Nielsen M, Padkjær SB, Lund O. Structural analysis of B-cell epitopes in antibody:protein complexes. Mol Immunol. 2013 Jan;53(1-2):24-34. doi: 10.1016/j.molimm.2012.06.001. Epub 2012 Jul 10. PubMed PMID: 22784991; PubMed Central PMCID: PMC3461403	29	240
2012	General	Salimi N, Fleri W, Peters B, Sette A. The Immune Epitope Database: A Historical Retrospective of the First Decade. Immunology. 2012 Jun 8. doi:10.1111/j.1365-2567.2012.03611.x. [Epub ahead of print] PubMed PMID: 22681406.	1	35
2012	General	Vaughan K, Kim Y, Sette A. A comparison of epitope repertoires associated with myasthenia gravis in humans and nonhuman hosts. Autoimmune Dis. 2012;2012:403915. doi:	0	6

		10.1155/2012/403915. Epub 2012 Dec 2. PubMed PMID: 23243503; PubMed Central PMCID: PMC3518085.		
2012	General	Vaughan K, Peters B, Larche M, Pomes A, Broide D, Sette A. Strategies to Query and Display Allergy-Derived Epitope Data from the Immune Epitope Database. <i>Int Arch Allergy Immunol.</i> 2012 Nov 21;160(4):334-345. [Epub ahead of print] PubMed PMID: 23172234.	1	11
2011	Tools/ Analysis Resource	Lundegaard C, Lund O, Nielsen M. Prediction of epitopes using neural network based methods. <i>J Immunol Methods.</i> 2011 Nov 30;374(1-2):26-34. doi: 10.1016/j.jim.2010.10.011. Epub 2010 Oct 31. PubMed PMID: 21047511.	5	42
2011	Tools/ Analysis Resource	Wang P, Sidney J, Sette A, Peters B. A computational pipeline to generate MHC binding motifs. <i>Immunome Res.</i> 2011 May;7(2). pii: 3. PubMed PMID: 28747991	0	0
2011	General	Seymour E., Damle R., Sette A., Peters B. (2011). "Cost sensitive hierarchical document classification to triage PubMed abstracts for manual curation." <i>BMC Bioinformatics</i> 12(1): 482. [Epub ahead of print]. PMID: 22182279.	0	10
2011	General	Vaughan K, Greenbaum J, Kim Y, Vita R, Chung J, Peters B, Broide D, Goodman R, Grey H, Sette A. Towards defining molecular determinants recognized by adaptive immunity in allergic disease: an inventory of the available data. <i>J Allergy (Cairo).</i> 2010;2010:628026. Epub 2011 Feb 13. PubMed PMID: 21403821.	0	12
2011	General	Vita R, Peters B, Josephs Z, de Matos P, Ennis M, Turner S, Steinbeck C, Seymour E, Zarebski L, Sette A. (2011). "A Model for Collaborative Curation, The IEDB and ChEBI Curation of Non-peptidic Epitopes." <i>Immunome Res.</i> 2011 Apr 1;7(1):1-8. PubMed PMID: 21897450.	1	3
2010	Tools/ Analysis Resource	Stranzl T, Larsen MV, Lundegaard C, Nielsen M. NetCTLpan: pan-specific MHC class I pathway epitope predictions. <i>Immunogenetics.</i> 2010 Jun;62(6):357-68. doi: 10.1007/s00251-010-0441-4. Epub 2010 Apr 9. PubMed PMID: 20379710.	39	177
2010	General	Oseroff C, Sidney J, Kotturi MF, Kolla R, Alam R, Broide DH, Wasserman SI, Weiskopf D, McKinney DM, Chung JL, Petersen A, Grey H, Peters B, Sette A. Molecular determinants of T cell epitope recognition to the common Timothy grass allergen. PubMed PMID: 20554959.	3	37
2010	General	Sidney J, Steen A, Moore C, Ngo S, Chung J, Peters B, Sette A. Divergent motifs but overlapping binding repertoires of six HLA-DQ molecules frequently expressed in the worldwide human population. <i>J Immunol.</i> 2010 Oct 1;185(7):4189-98. doi: 10.4049/jimmunol.1001006. Epub 2010 Sep 1. PubMed PMID: 20810981.	5	29
2010	General	Sidney J, Steen A, Moore C, Ngo S, Chung J, Peters B, Sette A. Five HLA-DP molecules frequently expressed in the worldwide human population share a common HLA supertypic binding specificity. <i>J Immunol.</i> 2010 Mar 1;184(5):2492-503. doi: 10.4049/jimmunol.0903655. Epub 2010 Feb 5. PubMed PMID: 20139279	6	42

2010	General	Moutaftsi M, Tschärke DC, Vaughan K, Koelle DM, Stern L, Calvo-Calle M, Ennis F, Terajima M, Sutter G, Crotty S, Drexler I, Franchini G, Yewdell JW, Head SR, Blum J, Peters B, Sette A. Uncovering the interplay between CD8, CD4 and antibody responses to complex pathogens. <i>Future Microbiol.</i> 2010 Feb;5(2):221-39. doi: 10.2217/fmb.09.110. PubMed: 20143946	3	31
2010	Tools/ Analysis Resource	Rapin N, Hoof I, Lund O, Nielsen M. The MHC motif viewer: a visualization tool for MHC binding motifs. <i>Curr Protoc Immunol.</i> 2010 Feb;Chapter 18:Unit 18.17. doi: 10.1002/0471142735.im1817s88. PubMed PMID: 20143317	1	15
2010	General	Brinkman RR, Courtot M, Derom D, Fostel JM, He Y, Lord P, Malone J, Parkinson H, Peters B, Rocca-Serra P, Ruttenberg A, Sansone SA, Soldatova LN, Stoeckert CJ Jr, Turner JA, Zheng J; OBI consortium. Modeling biomedical experimental processes with OBI. <i>J Biomed Semantics.</i> 2010 Jun 22;1 Suppl 1:S7. PubMed PMID: 20626927.	8	231
2010	Tools/ Analysis Resource	Kim Y, Sette A, Peters B. Applications for T-cell epitope queries and tools in the Immune Epitope Database and Analysis Resource. <i>J Immunol Methods.</i> 2010 Oct 31. [Epub ahead of print] PubMed PMID: 21047510.	1	48
2010	Tools/ Analysis Resource	Nielsen M, Lund O, Buus S, Lundegaard C. MHC class II epitope predictive algorithms. <i>Immunology.</i> 2010 Jul;130(3):319-28. Epub 2010 Apr 12. Review. PubMed PMID: 20408898;	20	195
2010	General	Ponomarenko J, Papangelopoulos N, Zajonc DM, Peters B, Sette A, Bourne PE. IEDB-3D: structural data within the immune epitope database. <i>Nucleic Acids Res.</i> 2010 Oct 28. [Epub ahead of print] PubMed PMID: 21030437.	5	58
2010	General	Salimi N, Fleri W, Peters B, Sette A. Design and utilization of epitope-based databases and predictive tools. <i>Immunogenetics.</i> 2010 Apr;62(4):185-96. Epub 2010 Mar 6. Review. PubMed PMID: 20213141; PubMed Central PMCID: PMC2843836.	3	39
2010	General	Vaughan K, Greenbaum J, Blythe M, Peters B, Sette A. Meta-analysis of all immune epitope data in the Flavivirus genus: inventory of current immune epitope data status in the context of virus immunity and immunopathology. <i>Viral Immunol.</i> 2010 Jun;23(3):259-84. PubMed PMID: 20565291; PubMed Central PMCID: PMC2942863.	4	54
2010	Tools/ Analysis Resource	Wang P, Sidney J, Kim Y, Sette A, Lund O, Nielsen M, Peters B. Peptide binding predictions for HLA DR, DP and DQ molecules. <i>BMC Bioinformatics.</i> 2010 Nov 22;11:568. PubMed PMID: 21092157; PubMed Central PMCID: PMC2998531.	67	546
2010	Tools/ Analysis Resource	Zhang H, Wang P, Papangelopoulos N, Xu Y, Sette A, Bourne PE, Lund O, Ponomarenko J, Nielsen M, Peters B. Limitations of Ab initio predictions of peptide binding to MHC class II molecules. <i>PLoS One.</i> 2010 Feb 17;5(2):e9272. PubMed PMID: 20174654; PubMed Central PMCID: PMC2822856.	3	43
2009	Tools/ Analysis Resource	Zhang H, Lundegaard C, Nielsen M. Pan-specific MHC class I predictors: a benchmark of HLA class I pan-specific prediction methods. <i>Bioinformatics.</i> 2009 Jan 1;25(1):83-9.	1	18

		doi: 10.1093/bioinformatics/btn579. Epub 2008 Nov 7. PubMed PMID: 18996943.		
2009	General	Loffredo JT, Sidney J, Bean AT, Beal DR, Bardet W, Wahl A, Hawkins OE, Piaskowski S, Wilson NA, Hildebrand WH, Watkins DI, Sette A. Two MHC class I molecules associated with elite control of immunodeficiency virus replication, Mamu-B*08 and HLA-B*2705, bind peptides with sequence similarity. PubMed PMID: 19494300.	4	36
2009	General	Walsh SR, Gillis J, Peters B, Mothé BR, Sidney J, Sette A, Johnson RP. Diverse recognition of conserved orthopoxvirus CD8+ T cell epitopes in vaccinated rhesus macaques. Vaccine. 2009 Aug 6;27(36):4990-5000. doi: 10.1016/j.vaccine.2009.05.077. Epub 2009 Jun 14. PubMed PMID: 19531389.	1	2
2009	General	Sette A, Grey H, Oseroff C, Peters B, Moutaftsi M, Crotty S, Assarsson E, Greenbaum J, Kim Y, Kolla R, Tschärke D, Koelle D, Johnson RP, Blum J, Head S, Sidney J. Definition of epitopes and antigens recognized by vaccinia specific immune responses: their conservation in variola virus sequences, and use as a model system to study complex pathogens. Vaccine. 2009 Dec 30;27 Suppl 6:G21-6. doi: 10.1016/j.vaccine.2009.10.011. PubMed PMID: 20006135.	3	19
2009	General	Davies V, Vaughan K, Damle R, Peters B, Sette A. Classification of the universe of immune epitope literature: representation and knowledge gaps. PLoS One. 2009 Sep 14;4(9):e6948. PubMed PMID: 19774228;	0	13
2009	General	Greenbaum JA, Kotturi MF, Kim Y, Oseroff C, Vaughan K, Salimi N, Vita R, Ponomarenko J, Scheuermann RH, Sette A, Peters B. Pre-existing immunity against swine-origin H1N1 influenza viruses in the general human population. Proc Natl Acad Sci U S A. 2009 Dec 1;106(48):20365-70. Epub 2009 Nov 16. PubMed PMID: 19918065; PubMed Central PMCID: PMC2777968.	10	301
2009	Tools/ Analysis Resource	Hoof I, Peters B, Sidney J, Pedersen LE, Sette A, Lund O, Buus S, Nielsen M, NetMHCpan, a method for MHC class I binding prediction beyond humans. Immunogenetics. 2009 Jan;61(1):1-13. PMID: 19002680.	49	653
2009	Tools/ Analysis Resource	Kim Y, Sidney J, Pinilla C, Sette A, Peters B. Derivation of an amino acid similarity matrix for peptide: MHC binding and its application as a Bayesian prior. BMC Bioinformatics. 2009 Nov 30;10:394. PubMed PMID: 19948066.	21	156
2009	General	Vaughan K, Blythe M, Greenbaum J, Zhang Q, Peters B, Doolan DL, Sette A, Meta-analysis of immune epitope data for all Plasmodia: overview and applications for malarial immunobiology and vaccine-related issues. Parasite Immunol. 2009 Feb;31(2):78-97. [PMID: 19149776]	1	2
2009	General	Vita R, Zarebski L, Greenbaum JA, Emami H, Hoof I, Salimi N, Damle R, Sette A, Peters B. The Immune Epitope Database 2.0. Nucleic Acids Res. 2009 Nov 11. [Epub ahead of print] PubMed PMID: 19906713.	14	550

2009	Tools/ Analysis Resource	Zhang H, Lund O, Nielsen M. The PickPocket method for predicting binding specificities for receptors based on receptor pocket similarities: Application to MHC-peptide binding. <i>Bioinformatics</i> . 2009 Mar 17. PMID: 19297351.	24	167
2008	General	Sidney J, Assarsson E, Moore C, Ngo S, Pinilla C, Sette A, Peters B. Quantitative peptide binding motifs for 19 human and mouse MHC class I molecules derived using positional scanning combinatorial peptide libraries. <i>Immunome Res</i> . 2008 Jan 25;4:2. doi: 10.1186/1745-7580-4-2. PubMed PMID: 18221540.	27	183
2008	General	Sidney J, Peters B, Frahm N, Brander C, Sette A. HLA class I supertypes: a revised and updated classification. <i>BMC Immunol</i> . 2008 Jan 22;9:1. doi: 10.1186/1471-2172-9-1. PubMed: 18211710.	48	322
2008	General	Kim M, Taylor J, Sidney J, Mikloska Z, Bodsworth N, Lagios K, Dunckley H, Byth-Wilson K, Denis M, Finlayson R, Khanna R, Sette A, Cunningham AL. Immunodominant epitopes in herpes simplex virus type 2 glycoprotein D are recognized by CD4 lymphocytes from both HSV-1 and HSV-2 seropositive subjects. <i>J Immunol</i> . 2008 Nov 1;181(9):6604-15. PubMed PMID: 18941251.	0	7
2008	Tools/ Analysis Resource	Lundegaard C., K. Lamberth, M. Harndahl, S. Buus, O. Lund, M. Nielsen. (2008). "NetMHC-3.0: accurate web accessible predictions of human, mouse and monkey MHC class I affinities for peptides of length 8-11." <i>Nucleic Acids Res</i> . 2008 36:W509-12. PMID: 18996943.	40	694
2008	Tools/ Analysis Resource	Nielsen, M., C. Lundegaard, T. Blicher, B. Peters, A. Sette, S. Justesen, S. Buus, O. Lund. (2008). "Quantitative Predictions of Peptide Binding to Any HLA-DR Molecule of Known Sequence: NetMHCIIpan." <i>PLoS Computational Biology</i> 4(7). PMID: 18604266.	18	229
2008	Tools/ Analysis Resource	Ponomarenko J, H. H. Bui, W. Li, N. Fusseder, P. E. Bourne, A. Sette, B. Peters. (2008). "ElliPro: a new structure-based tool for the prediction of antibody epitopes." <i>BMC Bioinformatics</i> , 9(514). PMID: 19055730.	204	1037
2008	Tools/ Analysis Resource	Ponomarenko J. V., M. H. C. van Regenmortel (2008) "B-cell epitope prediction." <i>Structural Bioinformatics</i> , Ed: Bourne PE, Gu J. Wiley-Liss; 2 edition.	6	69
2008	General	Vita R., B. Peters, A. Sette. (2008). "The Curation Guidelines of the Immune Epitope Database and Analysis Resource." <i>Cytometry A</i> 73(11): 1066-1070. PMID: 18688821	0	26
2008	Tools/ Analysis Resource	Wang, P., J. Sidney, C. Dow, B. Mothe, A. Sette, B. Peters. (2008). "A Systematic Assessment of MHC Class II Peptide Binding Predictions and Evaluation of a Consensus Approach." <i>PLoS Computational Biology</i> 4(4). PMID: 18389056	63	746
2008	General	Zarebski L. M., K. Vaughan, J. Sidney, B. Peters, H. Grey, K. D. Janda, A. Casadevall, A. Sette. (2008). "Analysis of epitope information related to Bacillus anthracis and Clostridium botulinum." <i>Expert Rev Vaccines</i> 7(1): 55-74. PMID: 18251694	2	25
2008	Tools/ Analysis Resource	Zhang, Q., P. Wang, et al. (2008). "Immune epitope database analysis resource (IEDB-AR)." <i>Nucleic Acids Research</i> 36: W513-W518. PMID: 18515843	28	294

2007	general	Sidney J, Peters B, Moore C, Pencille TJ, Ngo S, Masterman KA, Asabe S, Pinilla C, Chisari FV, Sette A. Characterization of the peptide-binding specificity of the chimpanzee class I alleles A 0301 and A 0401 using a combinatorial peptide library. Immunogenetics. 2007 Sep;59(9):745-51. Epub 2007 Aug 16. PubMed PMID: 17701407.	0	4
2007	Tools/ Analysis Resource	Beaver JE, Bourne PE, Ponomarenko JV. EpitopeViewer: a Java application for the visualization and analysis of immune epitopes in the Immune Epitope Database and Analysis Resource (IEDB). Immunome Res. 2007 Feb 21;3:3. PMID: 17313688	1	43
2007	General	Blythe MJ, Zhang Q, Vaughan K, de Castro R Jr, Salimi N, Bui HH, Lewinsohn DM, Ernst JD, Peters B, Sette A. An analysis of the epitope knowledge related to Mycobacteria. Immunome Res. 2007 Dec 14;3(1):10. PMID: 18081934	0	54
2007	General	Bui HH, Peters B, Assarsson E, Mbawuike I, Sette A. Ab and T cell epitopes of influenza A virus, knowledge and opportunities. Proc Natl Acad Sci U S A. 2007 Jan 2;104(1):246-51. PMID: 17200302	6	182
2007	Tools/ Analysis Resource	Bui HH, Sidney J, Li W, Fusseder N, Sette A. Development of an epitope conservancy analysis tool to facilitate the design of epitope-based diagnostics and vaccines. BMC Bioinformatics. 2007 Sep 26;8(1):361. PMID: 17897458.	64	382
2007	General	Ernst JD, Lewinsohn DM, Behar S, Blythe M, Schlesinger LS, Kornfeld H, Sette A. Meeting Report: NIH Workshop on the Tuberculosis Immune Epitope Database. Tuberculosis (Edinb). 2007 Dec 6. PMID: 18068490	0	29
2007	Tools/ Analysis Resource	Greenbaum JA, Andersen PH, Blythe M, Bui HH, Cachau RE, Crowe J, Davies M, Kolaskar AS, Lund O, Morrison S, Mumey B, Ofra Y, Pellequer JL, Pinilla C, Ponomarenko JV, Raghava GP, van Regenmortel MH, Roggen EL, Sette A, Schlessinger A, Sollner J, Zand M, Peters B. Towards a consensus on datasets and evaluation metrics for developing B-cell epitope prediction tools. J Mol Recognit. 2007 Mar-Apr;20(2):75-82. PMID: 17205610	16	218
2007	Tools/ Analysis Resource	Lundegaard C, Lund O, Kesmir C, Brunak S, Nielsen M. Modeling the adaptive immune system: predictions and simulations. Bioinformatics. 2007 Dec 15;23(24):3265-75. PMID: 18045832	6	115
2007	Tools/ Analysis Resource	Nielsen M, Lundegaard C, Blicher T, Lamberth K, Harndahl M, Justesen S, Røder G, Peters B, Sette A, Lund O, Buus S. NetMHCpan, a method for quantitative predictions of peptide binding to any HLA-A and -B locus protein of known sequence. PLoS ONE. 2007 Aug 29;2(8):e796. PMID: 17726526	39	500
2007	General	Peters B, Sette A. Integrating epitope data into the emerging web of biomedical knowledge resources. Nat Rev Immunol. 2007 Jun;7(6):485-90. PMID: 17479127	1	52
2007	Tools/ Analysis Resource	Ponomarenko JV, Bourne PE., Antibody-protein interactions: benchmark datasets and prediction tools evaluation. BMC Struct Biol. 2007 Oct 2;7(1):64. PMID: 17910770	12	207

2007	General	Sette A, Peters B., Immune epitope mapping in the post-genomic era: lessons for vaccine development. <i>Curr Opin Immunol.</i> 2007 Feb;19(1):106-10. PMID: 17113275	0	55
2007	General	Wang P, Morgan AA, Zhang Q, Sette A, Peters B. Automating document classification for the Immune Epitope Database. <i>BMC Bioinformatics.</i> 2007 Jul 26;8:269. PMID: 17655769	0	40
2006	Tools/ Analysis Resource	Bui HH, Sidney J, Dinh K, Southwood S, Newman MJ, Sette A. Predicting population coverage of T-cell epitope-based diagnostics and vaccines. <i>BMC Bioinformatics.</i> 2006 Mar 17;7(1):153. PMID: 16545123	105	534
2006	Tools/ Analysis Resource	Peters B, Bui HH, Frankild S, Nielson M, Lundegaard C, Kostem E, Basch D, Lamberth K, Harndahl M, Fleri W, Wilson SS, Sidney J, Lund O, Buus S, Sette A., A community resource benchmarking predictions of peptide binding to MHC-I molecules, <i>PLoS Comput Biol.</i> 2006 Jun 9;2(6):e65. Epub 2006 Jun 9. PMID: 16789818	4	279
2006	General	Salimi N, R. Vita. (2006). "The biocurator: connecting and enhancing scientific data". <i>PLoS Comput Biol</i> 2(10) :e125. PMID: 17069454	0	30
2006	General	Sette, A., H. H. Bui, S. Buus, W. Fleri, R. Kubo, O. Lund, D. Nemazee, J. V. Ponomarenko, M. Sathiamurthy, S. Stewart, S. Way, S. S. Wilson, B. Peters. (2006). "The Immune Epitope Database and Analysis Resource." <i>Pattern Recognition in Bioinformatics: International Workshop, Hong Kong, China, August 20, 2006 : Proceedings, Springer, Jagath C. Rajapakse, Limsoon Wong, Raj Acharya (Eds). ISBN: 3540374469 9783540374466</i>	0	3
2006	General	Vita R, K. Vaughan, L. Zarebski, N. Salimi, W. Fleri , H. Grey, M. Sathiamurthy, J. Mokili, H. H. Bui, P. E. Bourne, J. V. Ponomarenko, R. de Castro Jr, R. K. Chan, J. Sidney, S. S. Wilson, S. Stewart, S. Way, B. Peters, A. Sette. (2006) "Curation of complex, context-dependent immunological data." <i>BMC Bioinformatics.</i> 12(7): 341. PMID: 16836764	0	21
2005	General	Sette A, Sidney J, Bui HH, del Guercio MF, Alexander J, Loffredo J, Watkins DI, Mothé BR. Characterization of the peptide-binding specificity of Mamu-A*11 results in the identification of SIV-derived epitopes and interspecies cross-reactivity. <i>Immunogenetics.</i> 2005 Apr;57(1-2):53-68. Epub 2005 Mar 4. PubMed PMID: 15747117.	0	0
2005	Tools/ Analysis Resource	Peters B, Bui HH, Sidney J, Weng Z, Loffredo JT, Watkins DI, Mothé BR, Sette A. A computational resource for the prediction of peptide binding to Indian rhesus macaque MHC class I molecules. <i>Vaccine.</i> 2005 Nov 1;23(45):5212-24. Epub 2005 Aug 18. PubMed PMID: 16137805.	0	5
2005	Tools/ Analysis Resource	Peters B, Sette A. Generating quantitative models describing the sequence specificity of biological processes with the stabilized matrix method. <i>BMC Bioinformatics.</i> 2005 May 31;6:132. PMID: 15927070	29	490
2005	General	Peters, B., J. Sidney, P. Bourne, H. H. Bui, S. Buus, G. Doh, W. Fleri, M. Kronenberg, R.Kubo, O. Lund, D. Nemazee, J. V. Ponomarenko, M. Sathiamurthy, S. Schoenberger, S. Stewart, P. Surko, S. Way, S. Wilson, A. Sette. (2005). "The design and implementation of the immune epitope database	5	85

		and analysis resource." Immunogenetics 57(5): 326-336. PMID: 15895191		
2005	General	Peters, B., J. Sidney, P. Bourne, H. H. Bui, S. Buus, G. Doh, W. Fleri, M. Kronenberg, R. Kubo, O. Lund, D. Nemazee, J. V. Ponomarenko, M. Sathiamurthy, S. Schoenberger, S. Stewart, P. Surko, S. Way, S. Wilson, A. Sette. (2005). "The Immune Epitope Database and Analysis Resource: From Vision to Blueprint." PLoS Biology 3(3). PMID: 15760272	9	336
2005	General	Sathiamurthy, M., B. Peters, H. H. Bui, J. Sidney, J. Mokili, S. S. Wilson, W. Fleri, D. McGuinness, P. Bourne, A. Sette. (2005). "An ontology for immune epitopes: application to the design of a broad scope database of immune reactivities." Immunome Res 1(2): 1745-7580. PMID: 16305755	1	29
2005	General	Sette, A., W. Fleri, B. Peters, M. Sathiamurthy, H. H. Bui, S. Wilson. (2005). "A Roadmap for the Immunomics of Category A–C Pathogens." Immunity 22(2): 155-161. PMID: 15773067	3	68
2004	General	Sette A. (2004). "The immune epitope database and analysis resource: from vision to blueprint." Genome Inform Ser Workshop Genome Inform 15(2):299. PMID: 16312048	1	10
2004	General	Loffredo JT, Sidney J, Wojewoda C, Dodds E, Reynolds MR, Napoé G, Mothé BR, O'Connor DH, Wilson NA, Watkins DI, Sette A. Identification of seventeen new simian immunodeficiency virus-derived CD8+ T cell epitopes restricted by the high frequency molecule, Mamu-A*02, and potential escape from CTL recognition. J Immunol. 2004 Oct 15;173(8):5064-76. PubMed PMID: 15470050.	0	10
2003	General	Iwai LK, Yoshida M, Sidney J, Shikanai-Yasuda MA, Goldberg AC, Juliano MA, Hammer J, Juliano L, Sette A, Kalil J, Travassos LR, Cunha-Neto E. In silico prediction of peptides binding to multiple HLA-DR molecules accurately identifies immunodominant epitopes from gp43 of Paracoccidioides brasiliensis frequently recognized in primary peripheral blood mononuclear cell responses from sensitized individuals. Mol Med. 2003 Sep-Dec;9(9-12):209-19. PubMed PMID: 15208742.	6	14
In-Text / Informal Citations (data captured from 2012 only)			489	3,556
Total from Publications			4,468	24,994

3.2 Publications Citing the IEDB in 2022

In 2022, the IEDB or Analysis Resource received a total of 4,468 individual citations (3,979 total citations for IEDB publications and 489 additional inline citations). This includes citations received by each of the 187 papers written by the IEDB team over the past twenty years. This represents an increase of 36 citations over 4,432 from the previous year (2021).

The formal citation list was compiled by using Google Scholar and the ISI Web of Knowledge to find citations for each one of the 187 papers and excludes self-citations. In contrast, often scientific reports quote the IEDB not as a formal citation, but by simply stating in the text that the IEDB was used or listing the website address. Here we developed and implemented a plan to address in-text citations (informal mentions of the IEDB, rather than a direct citation). For this purpose, we utilized Publish or Perish, a software

program that retrieves citations from Crossref, Google Scholar, Microsoft Academic, Scopus, and Web of Science, and returns output that can easily be converted to an excel file. The tool can capture citations that the document specialist's citation analysis, which includes Google Scholar and Web of Science, may have missed, and can capture references that cite the IEDB in text but do not cite a specific IEDB-related publication.

The papers have been categorized by the type of IEDB paper they cited – General IEDB or Analysis Resource (AR). By reference, the General IEDB (G) papers received 1,161 citations in 2022 and the Analysis Resource (AR) papers received 2,818 citations. Additionally, there were 489 inline citations, which, in total, sums to the expected 4,468 citations for 2022.

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